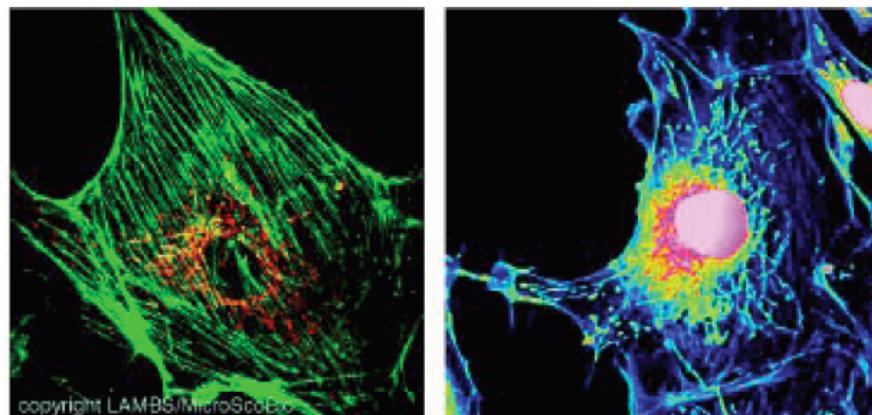




IFOM Milano, 14 Marzo 2006



INVESTIGAZIONI FOTONICHE



ALBERTO DIASPRO

DEPARTMENT OF PHYSICS, UNIVERSITY OF GENOA
LAMBS, MICROScobIO RESEARCH CENTER, IFOM

diaspro@fisica.unige.it

<http://www.lambs.it>

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Ilaria Testa

Mattia Pesce

Federica Morotti

Francesco Di Fato

Francesca Cella

Emiliano Ronzitti

Cristina Barmo

Johnatan Roberts



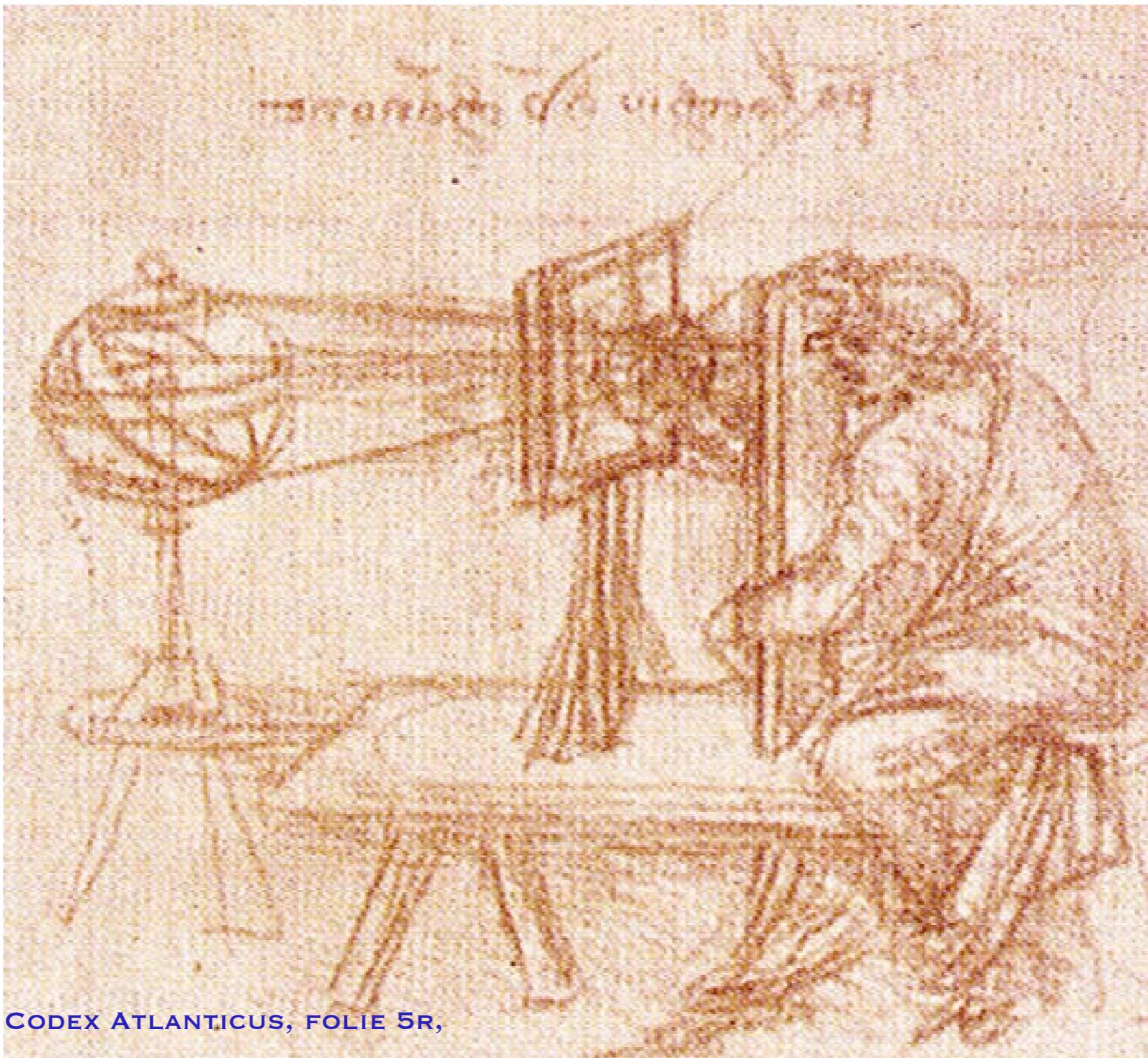
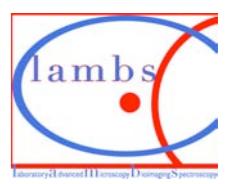
NANoMED_{Lab}
Nano technology for bioMedicine

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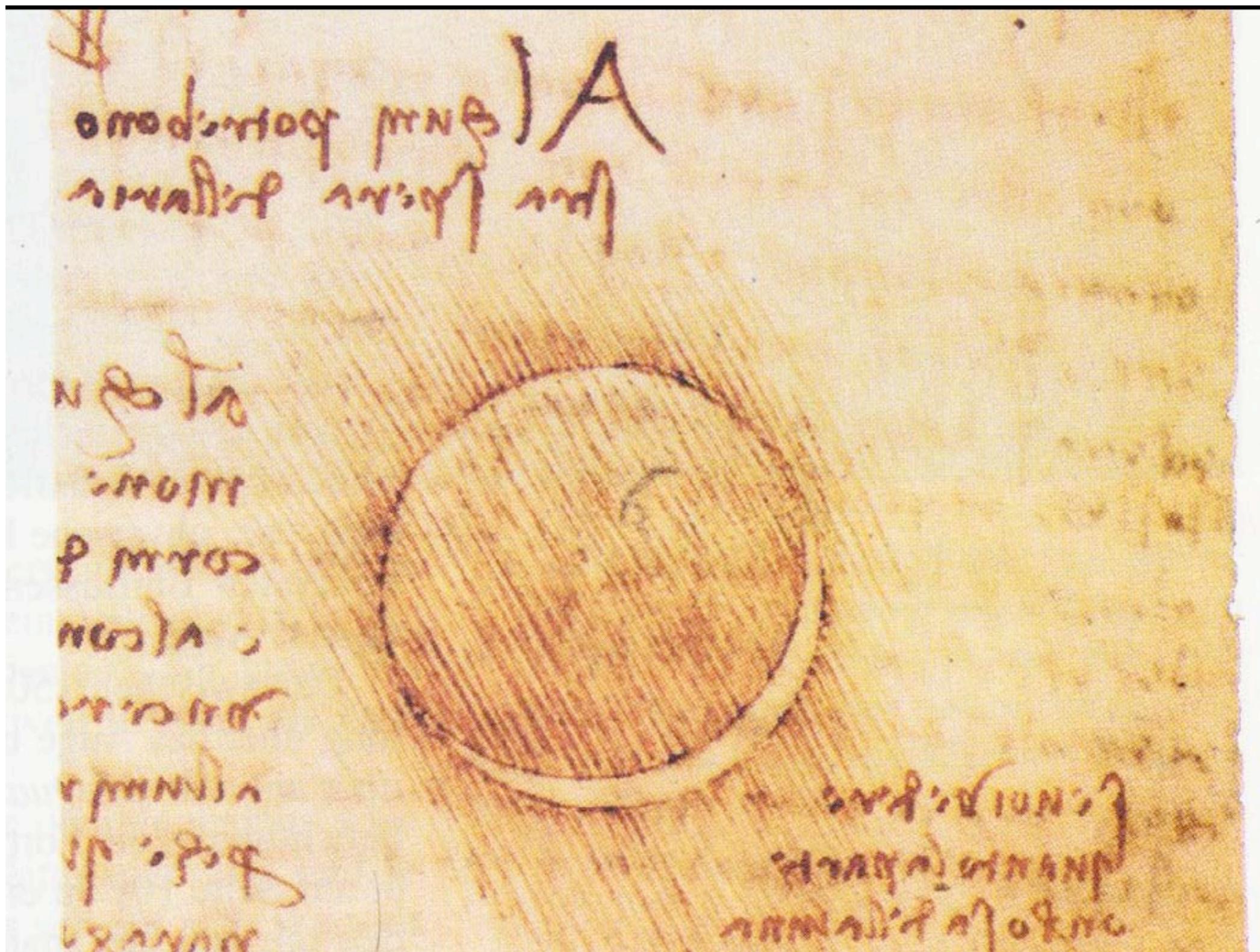
OBSERVING EVENTS



CODEX ATLANTICUS, FOLIE 5R,

SLIDE CREDIT: IRINA MAJOUЛ ROYAL HOLLOWAY UNIVERSITY OF LONDON

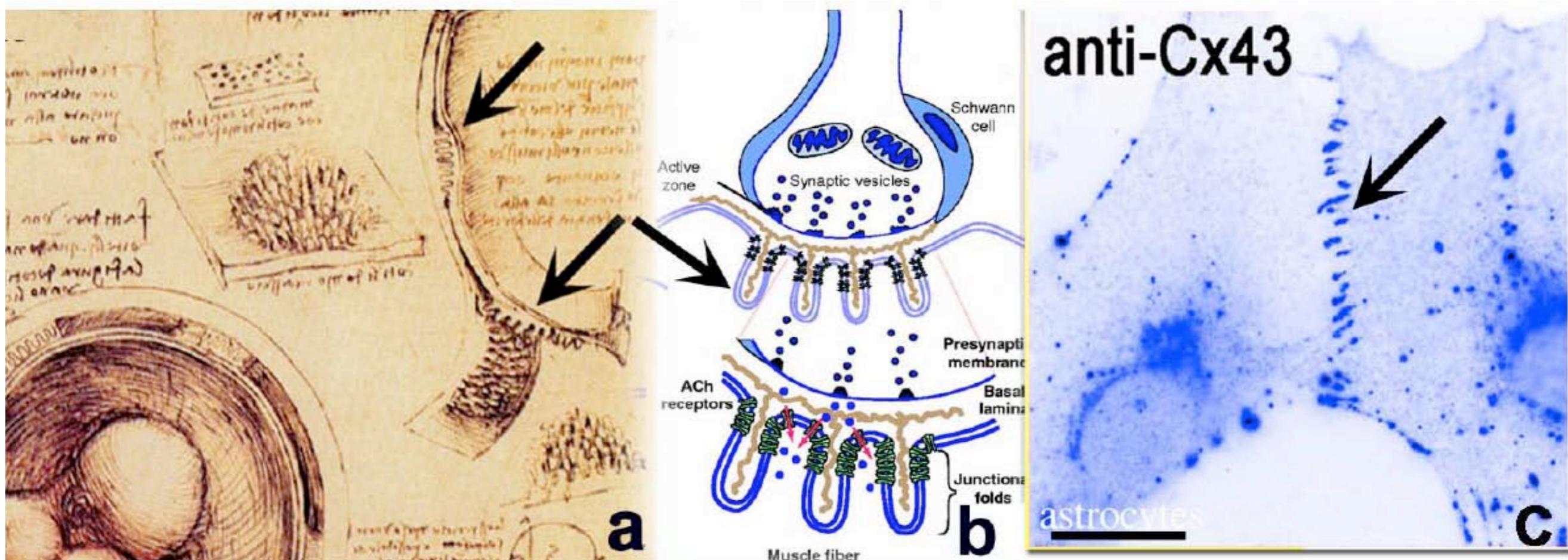
OBSERVING EVENTS



LEONARDO DA VINCI, IMAGE OF A MOON PHASE FROM THE CODEX LEICESTER: 35v, 2R

SLIDE CREDIT: IRINA MAJOUl ROYAL HOLLOWAY UNIVERSITY OF LONDON

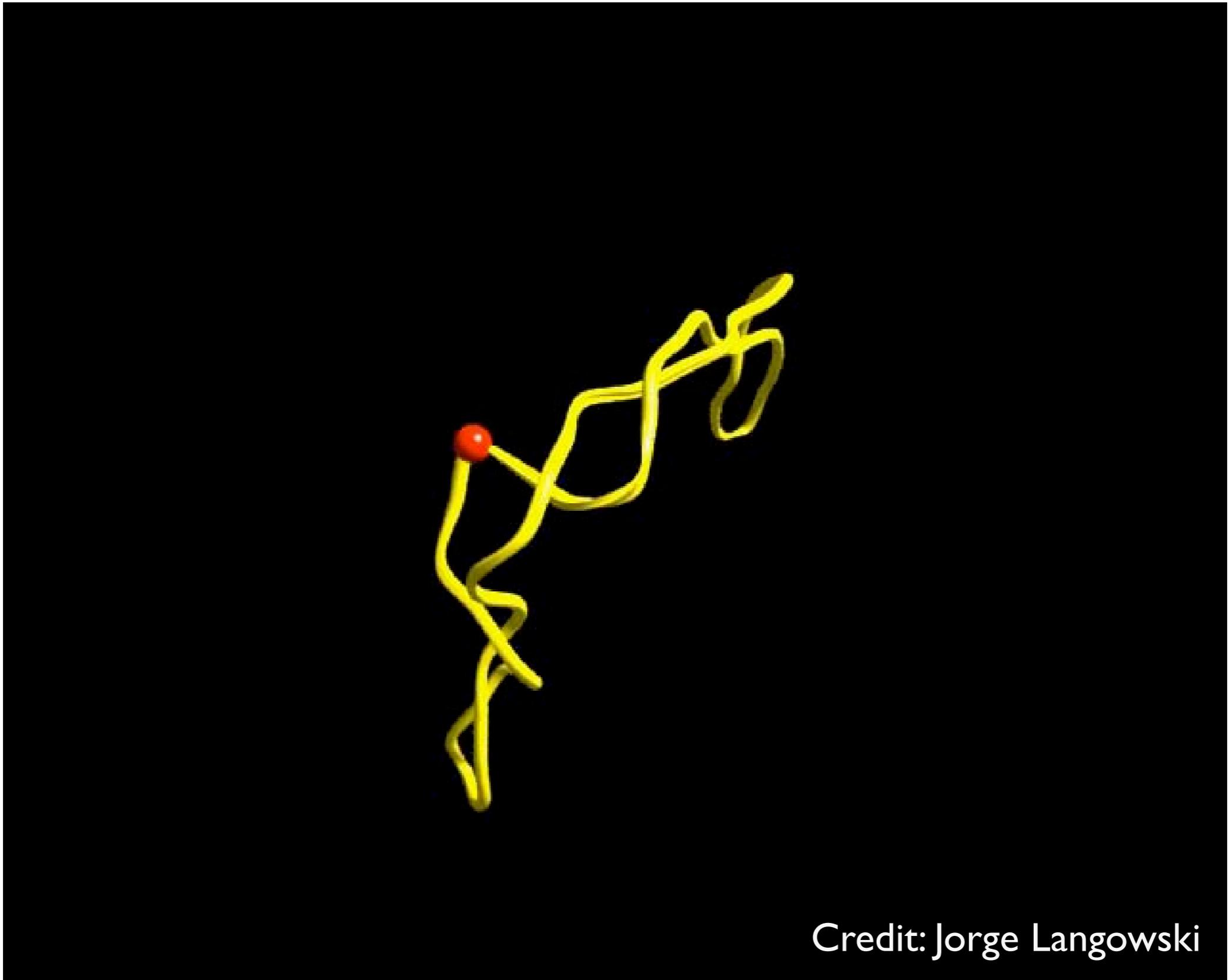
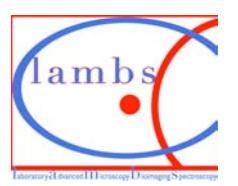
OBSERVING EVENTS



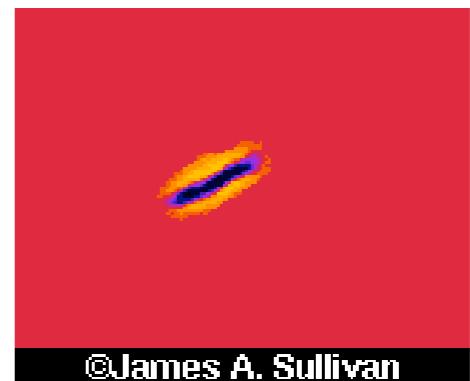
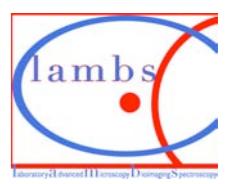
A) – TAKEN FROM AN ANATOMICAL STUDY OF LEONARDO DA VINCI 1508 - 1510 (MANUSCRIPT RL, 19102R, WINDSOR LIBRARY, UK). THE LEFT PANEL SHOWS THE INTERFACE BETWEEN DISSECTED TISSUES (WALL OF A WOMB) WHERE LEONARDO USED THE TOOLS AVAILABLE FOR HIM TO ANALYSE HOW TWO BIOLOGICAL SURFACES CREATE AN ADHESIVE JUNCTIONAL INTERFACE; B) – AN INSERT REPRESENTING JUNCTIONAL FOLDS OF A SYNAPTIC TERMINAL THAT CREATES INTEGRITY OF THE STRUCTURE (COHEN-CORY, 2002). C) – JUNCTIONAL CELL-CELL INTERFACE BETWEEN TWO CULTURED PRIMARY ASTROCYTES REVEALED WITH ANTIBODIES AGAINST A GAP JUNCTION PROTEIN, CONNEXIN-43 (BUTKEVICH ET AL., 2004).

SLIDE CREDIT: IRINA MAJOUЛ ROYAL HOLLOWAY UNIVERSITY OF LONDON

OBSERVING EVENTS

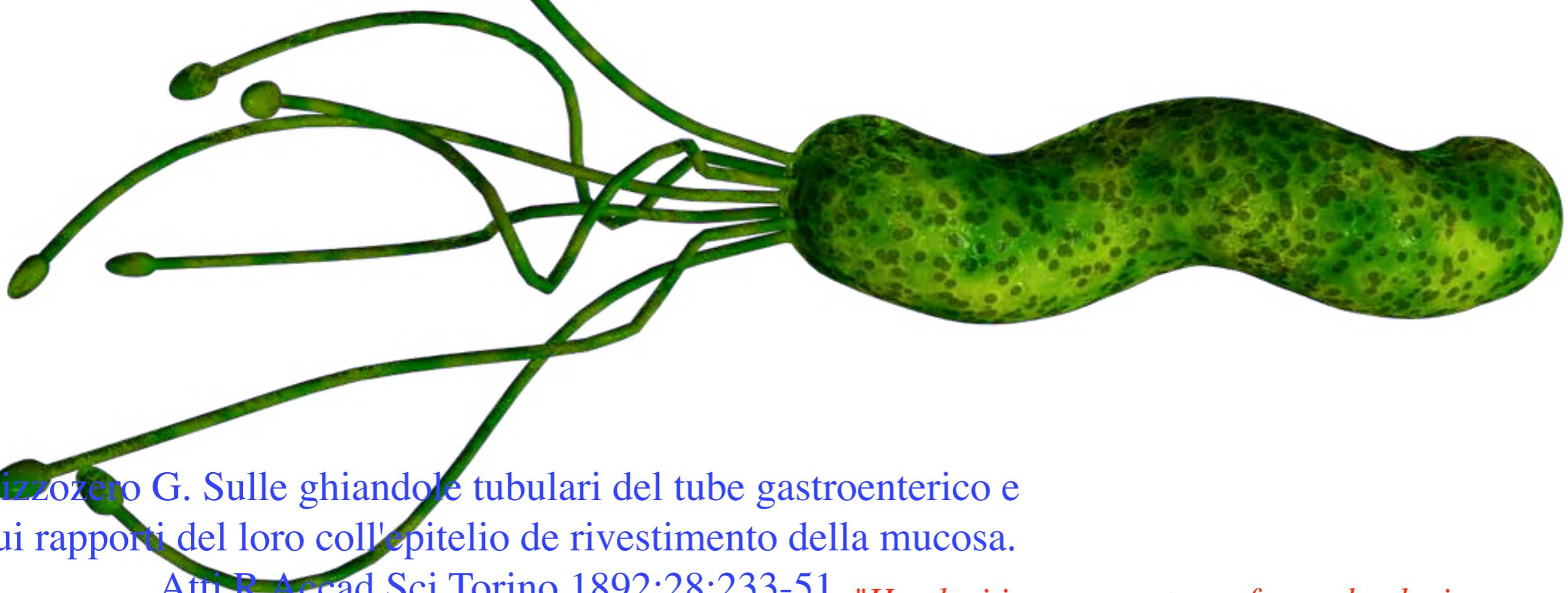


OBSERVING EVENTS



The Nobel Prize in Physiology or Medicine 2005

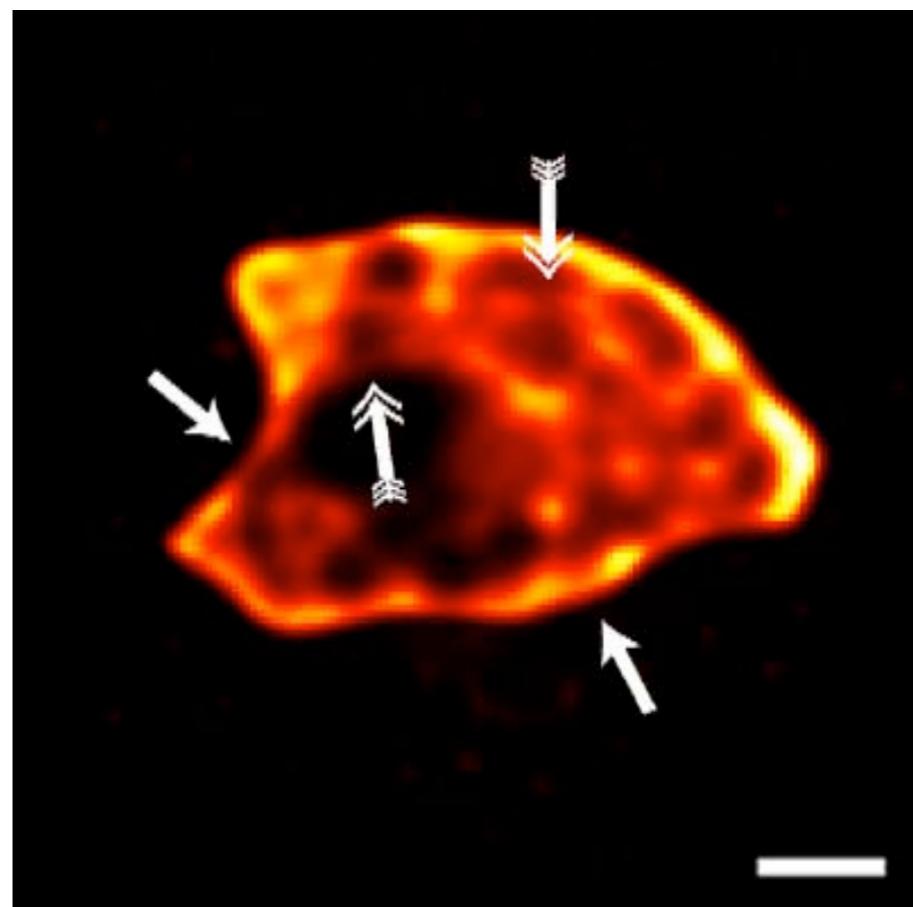
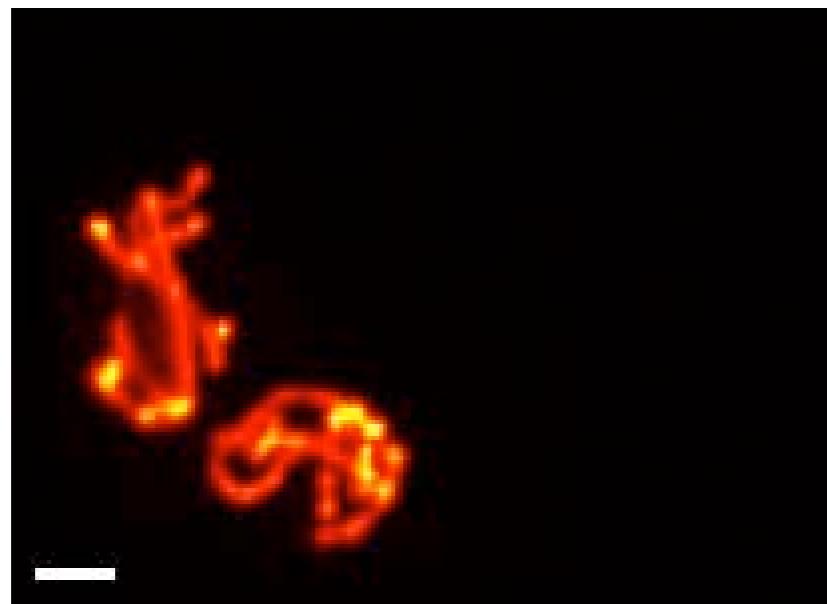
Helicobacters were discovered in Italy in 1892: An episode in the scientific life of an eclectic pathologist, Giulio Bizzozero



Bizzozero G. Sulle ghiandole tubulari del tube gastroenterico e sui rapporti del loro coll'epitelio de rivestimento della mucosa.

Atti R Accad Sci Torino 1892;28:233-51 "H.pylori image courtesy of www.hpylori.com.au"

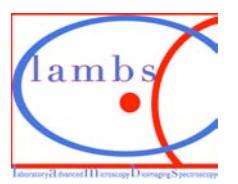
OBSERVING EVENTS



Credit: Stefan Hell, JCS web site

Journal of Cell Science 116 (10), 2005
Spatial and temporal dynamics of budding yeast
mitochondria lacking the division component Fis1p

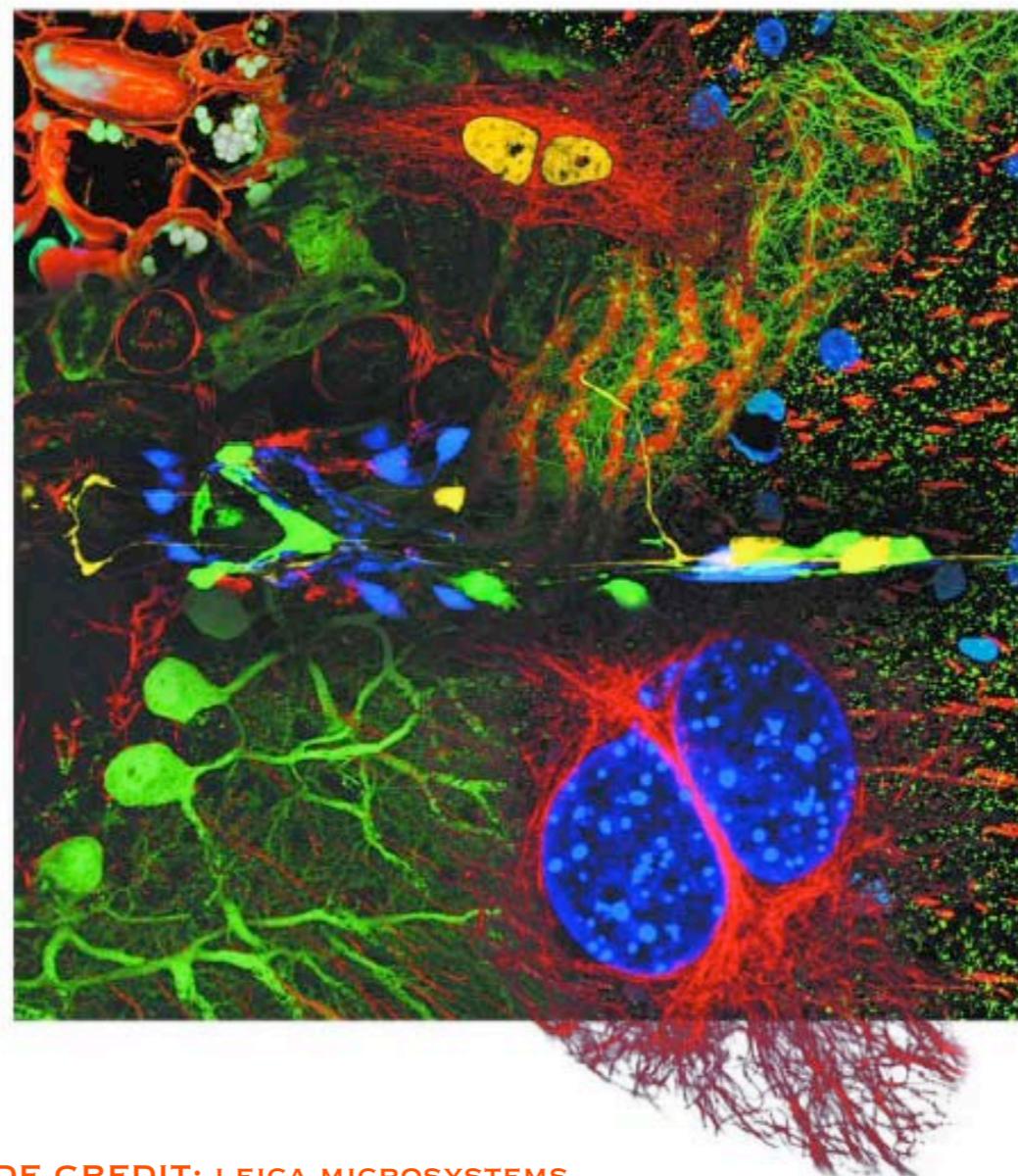
CAPTURING EVENTS



MICROSCOPY

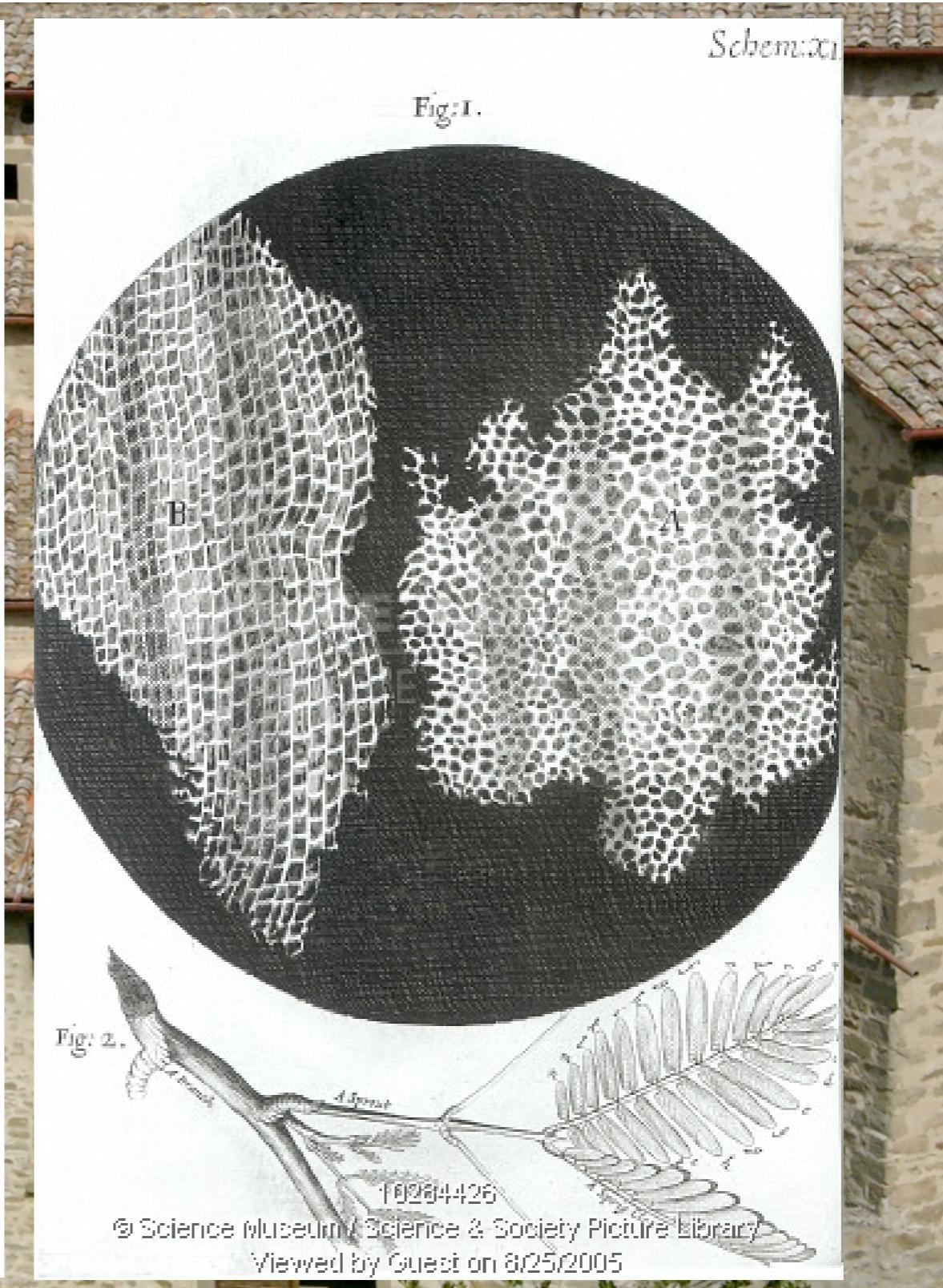
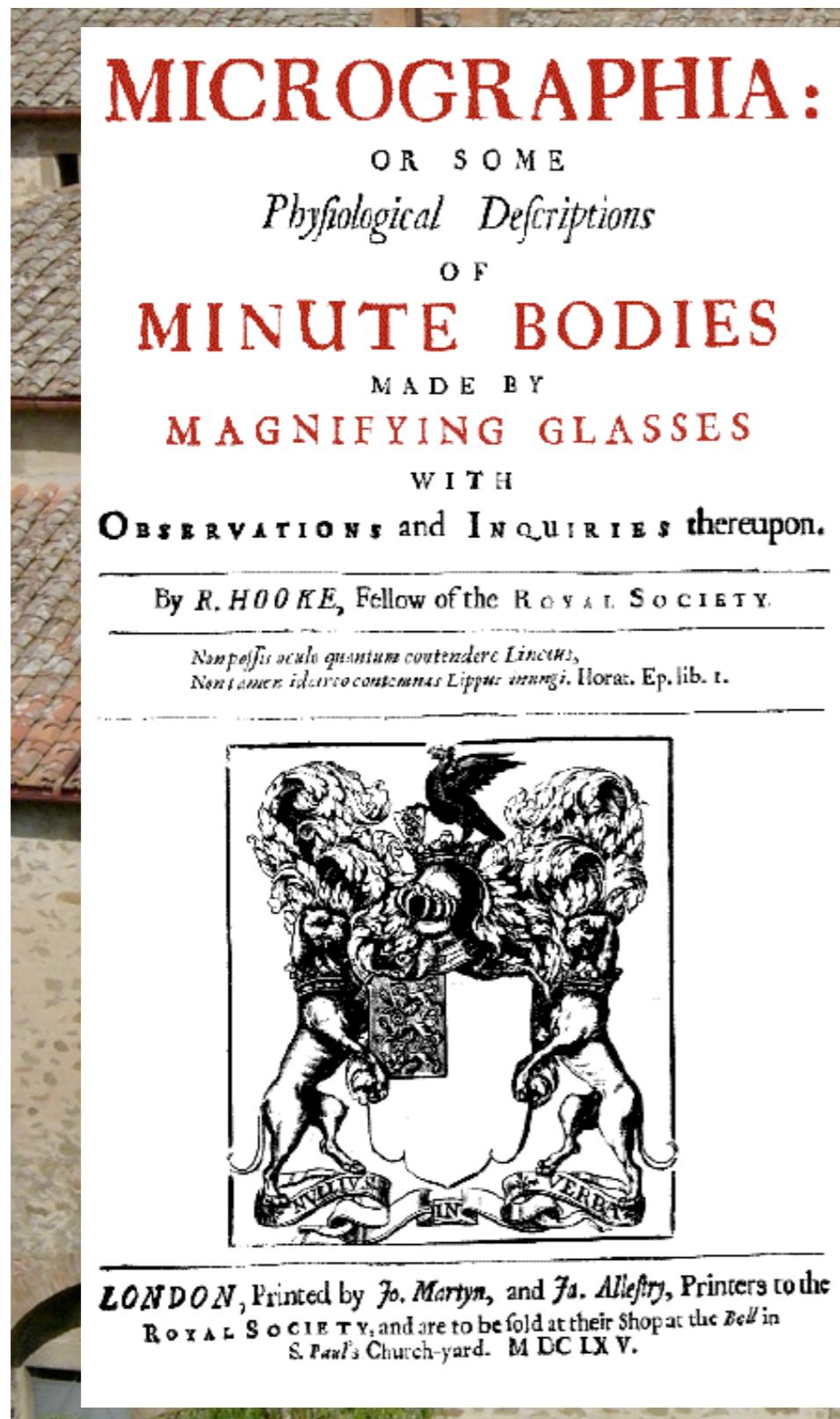
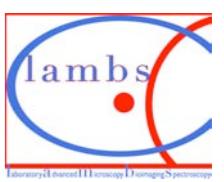
A microscope is an instrument magnifying objects by means of a specific interaction – more commonly by means of lenses – so as to capture details invisible to the naked eye.

(Oxford dictionary, after Colin Sheppard)

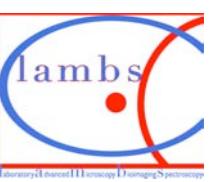


SLIDE CREDIT: LEICA MICROSYSTEMS

OPTICAL MICROSCOPY



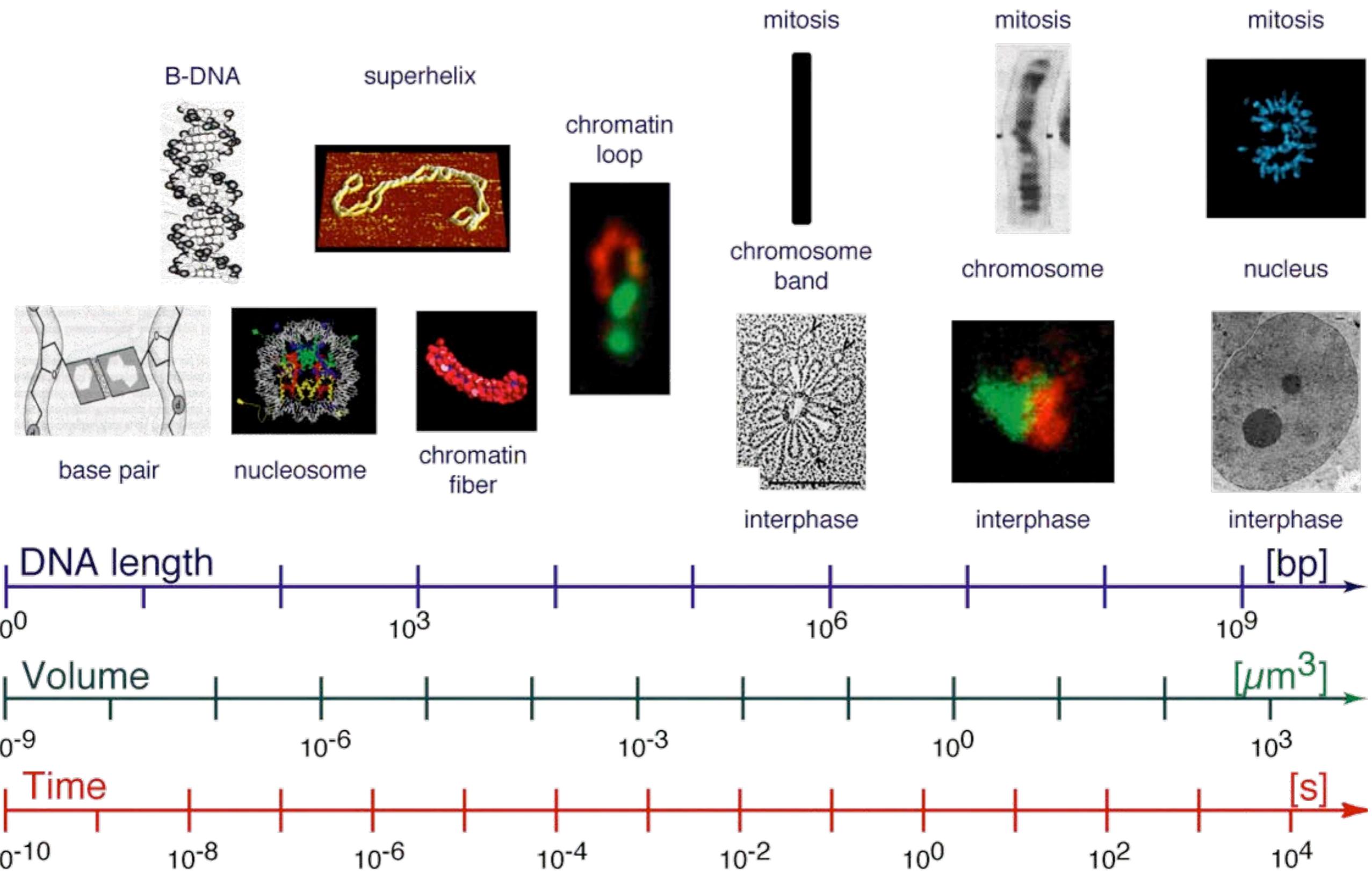
OPTICAL MICROSCOPY



copyright Darwin/Elvio Paoni

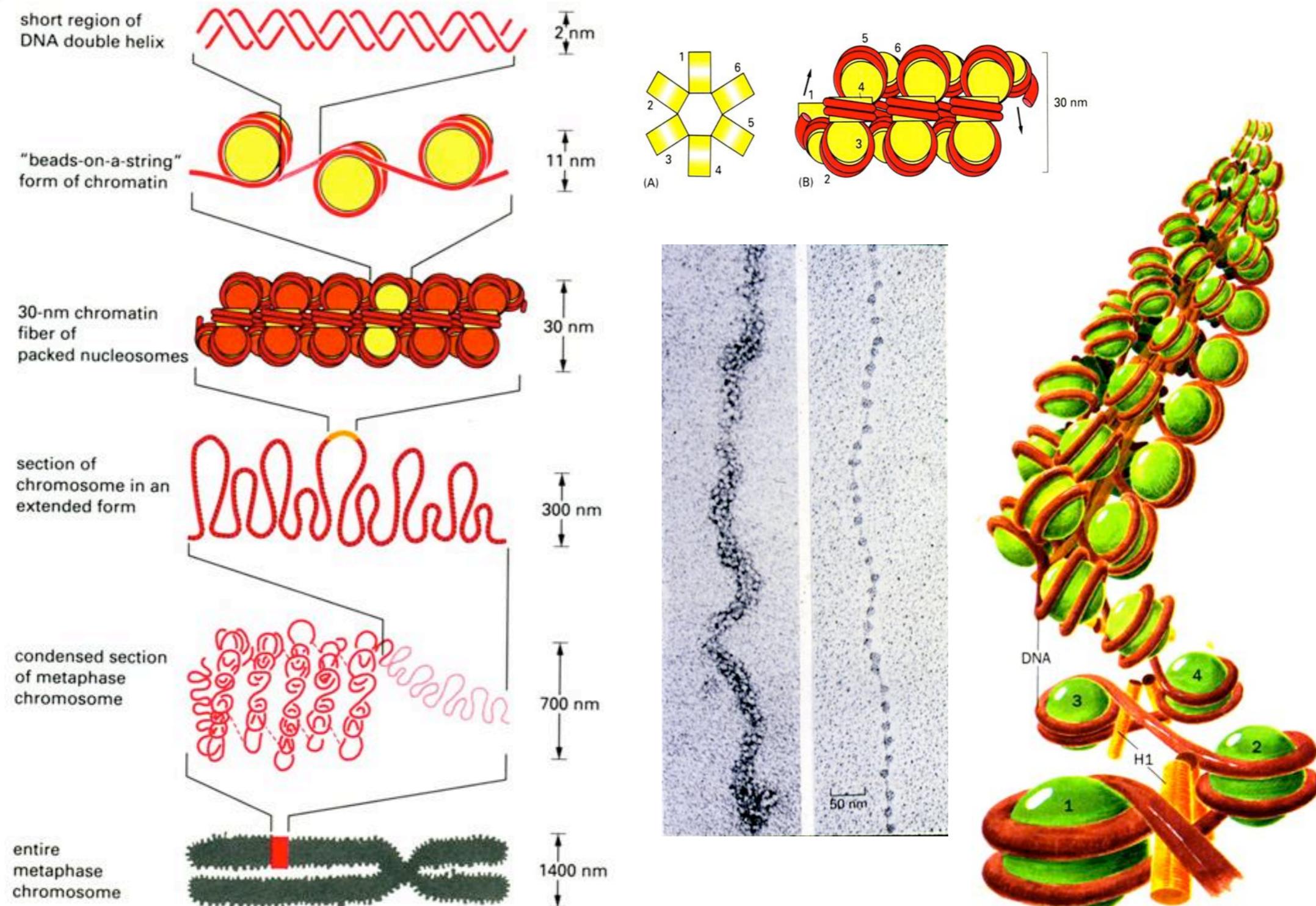


THE BIOLOGICAL SCENARIO

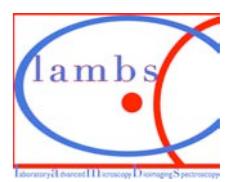


Slide credit: Jörg Langowski, DKFZ, Heidelberg

THE BIOLOGICAL SCENARIO



RESOLUTION OBSESSION



HUMAN EYE

100 000 nm

OPTICAL MICROSCOPE

200 nm

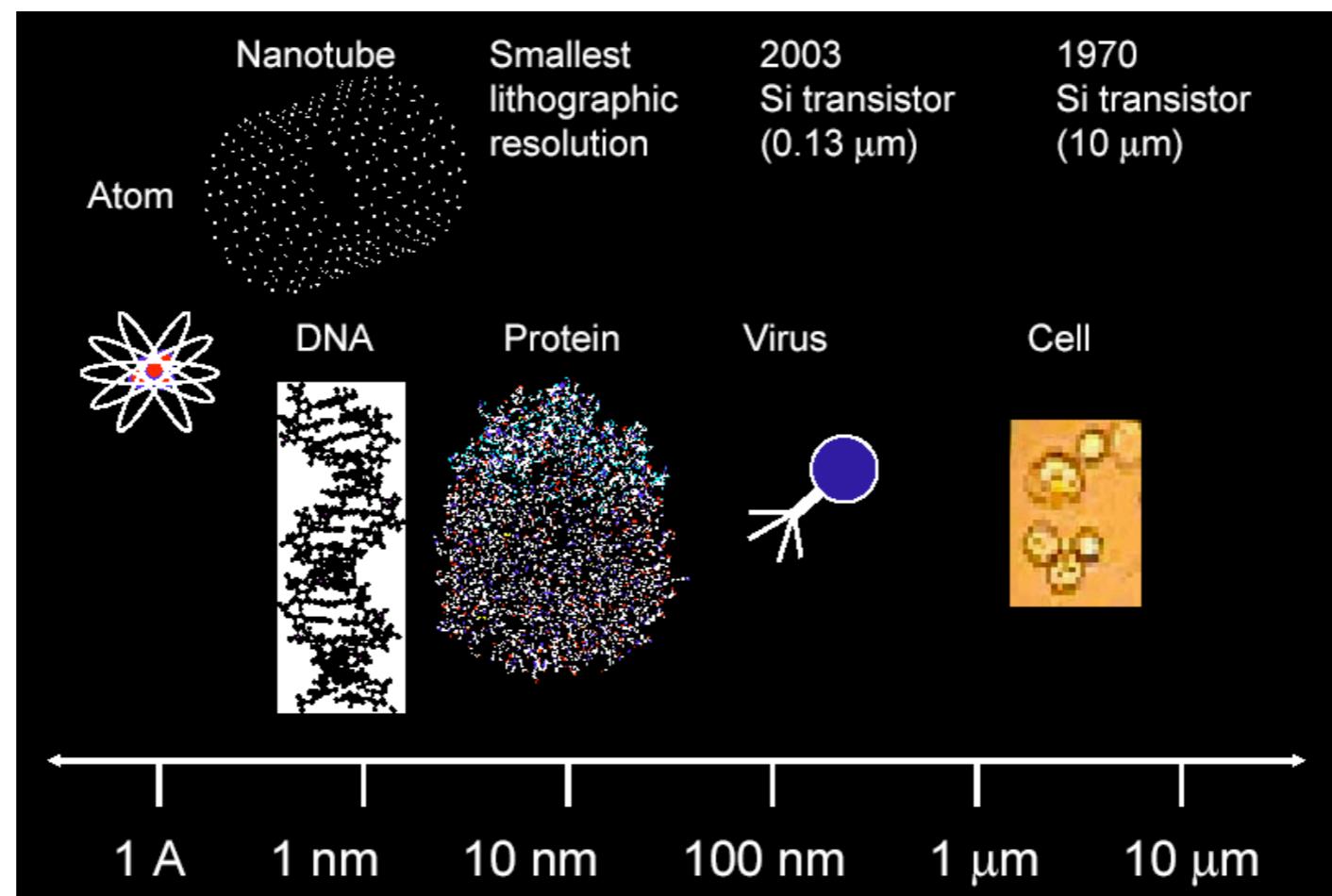
ELECTRON MICROSCOPE

0.4 nm

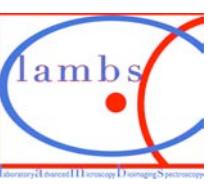
3 nm

SCANNING PROBE MICROSCOPE

0.1-10 nm

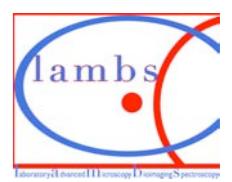


OPTICAL MICROSCOPY IMAGING



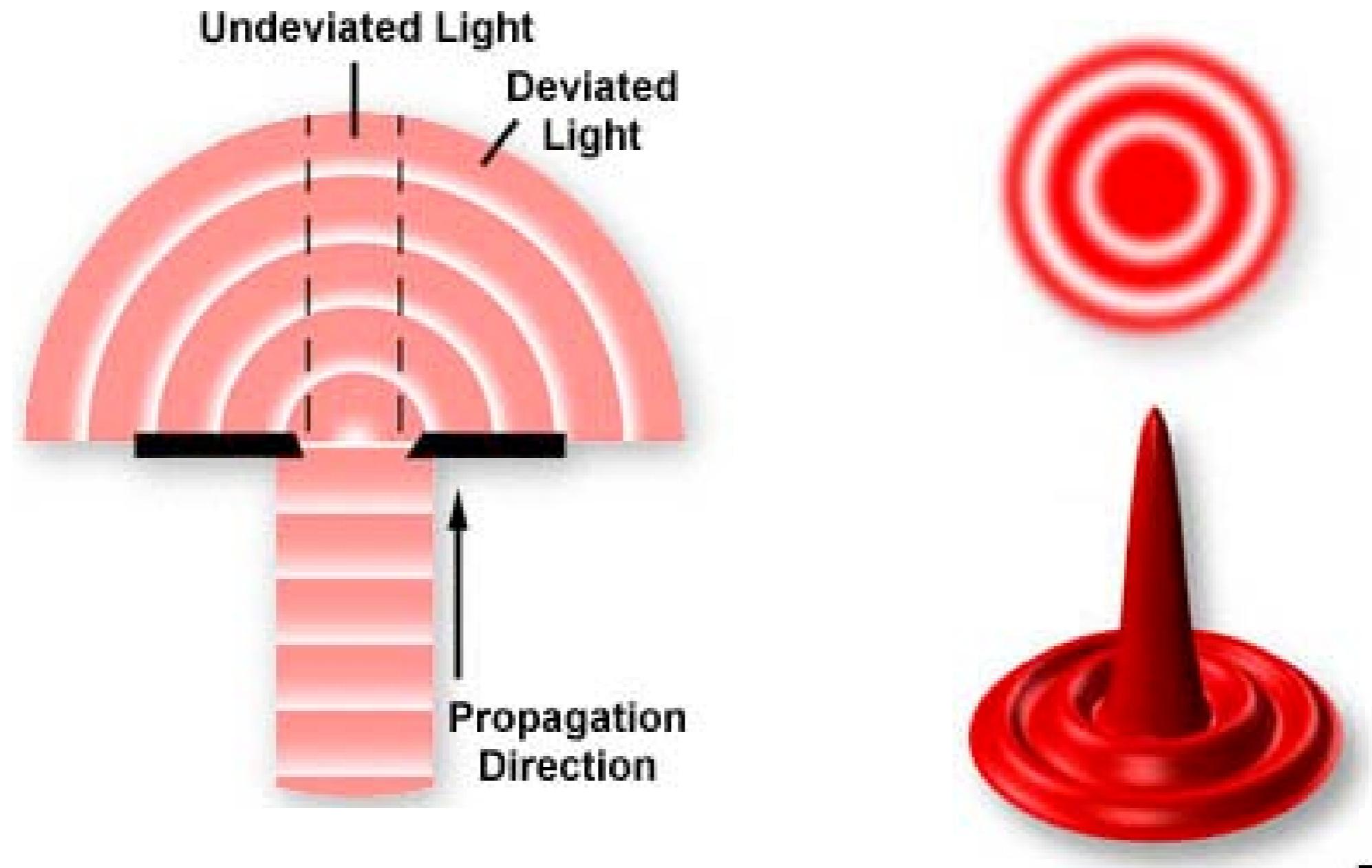
Georges Seurat (1859-1891), Una domenica pomeriggio all'isola della Grande Jatte, 1883-85

LENS EFFECT

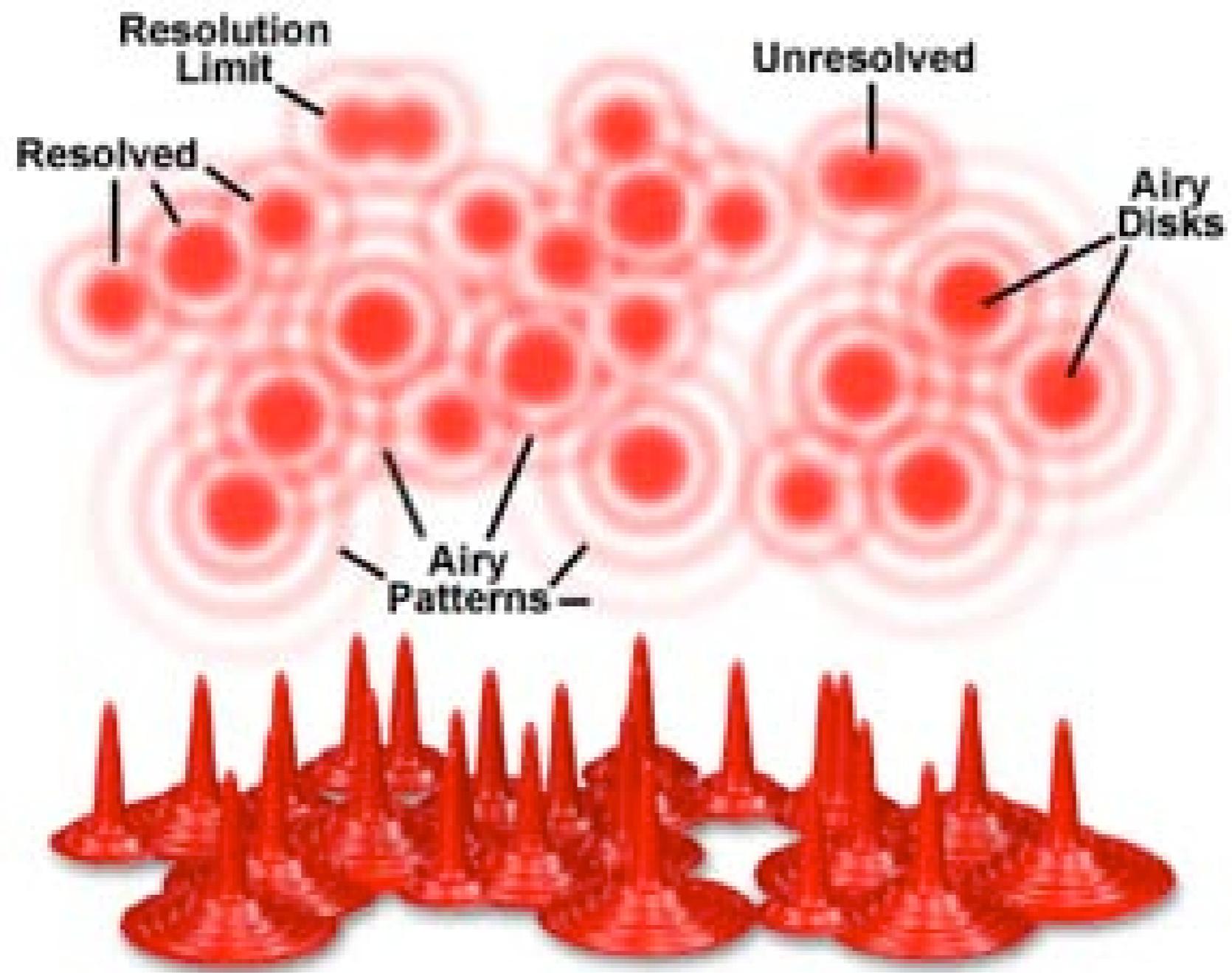


POINTLIKE OBJECT VS. QUASI POINTLIKE IMAGE

OPTICAL RESOLUTION

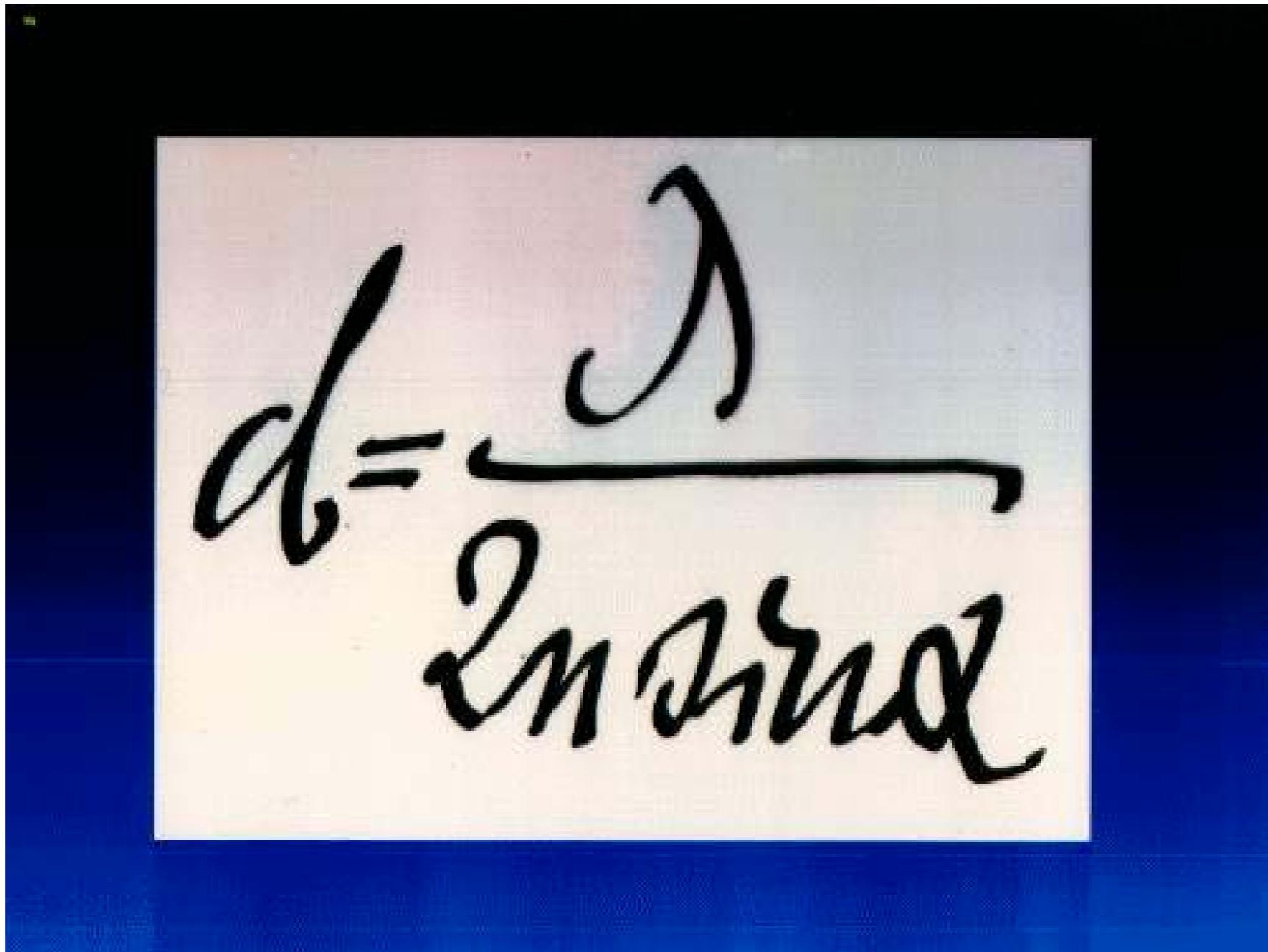
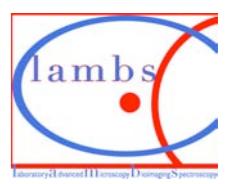


OPTICAL RESOLUTION



Vignette credit: <http://www.microscopyu.com/tutorials/>

OPTICAL RESOLUTION



ABBE, E (1873) ARCHIVE F. MIKROSKOP.ANAT. 9, 413-420

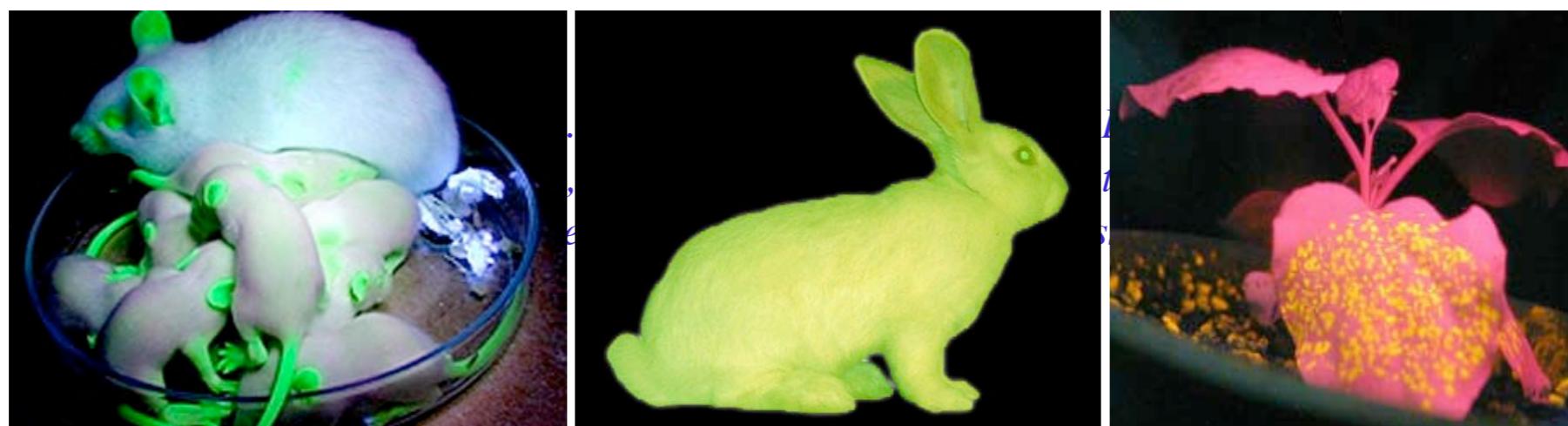
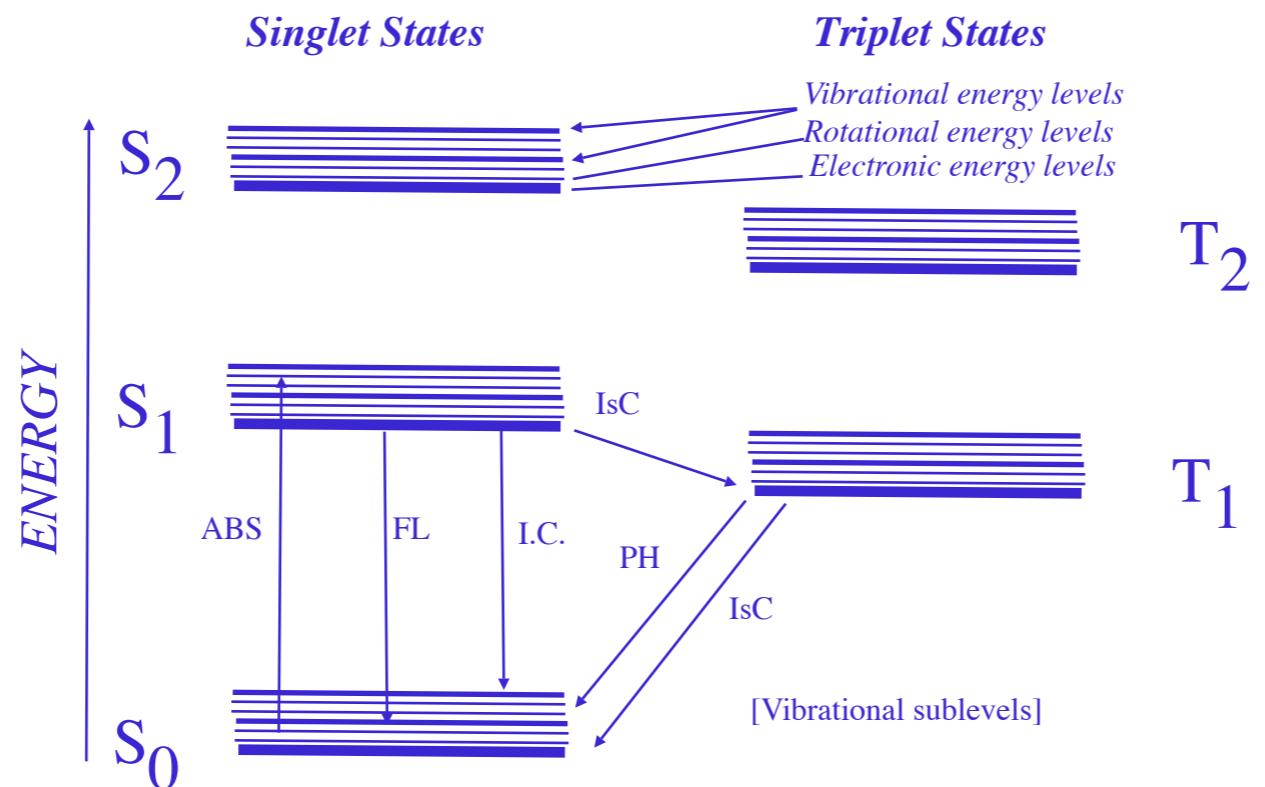
OPTICAL CONTRAST



Giudizio Universale: immagine nel VIS (sinistra) e in fluorescenza indotta UV (destra): l'azzurro di lapislazzuli, utilizzato da Michelangelo, e' riconoscibile dalla presenza di una fluorescenza bianco-verdastra

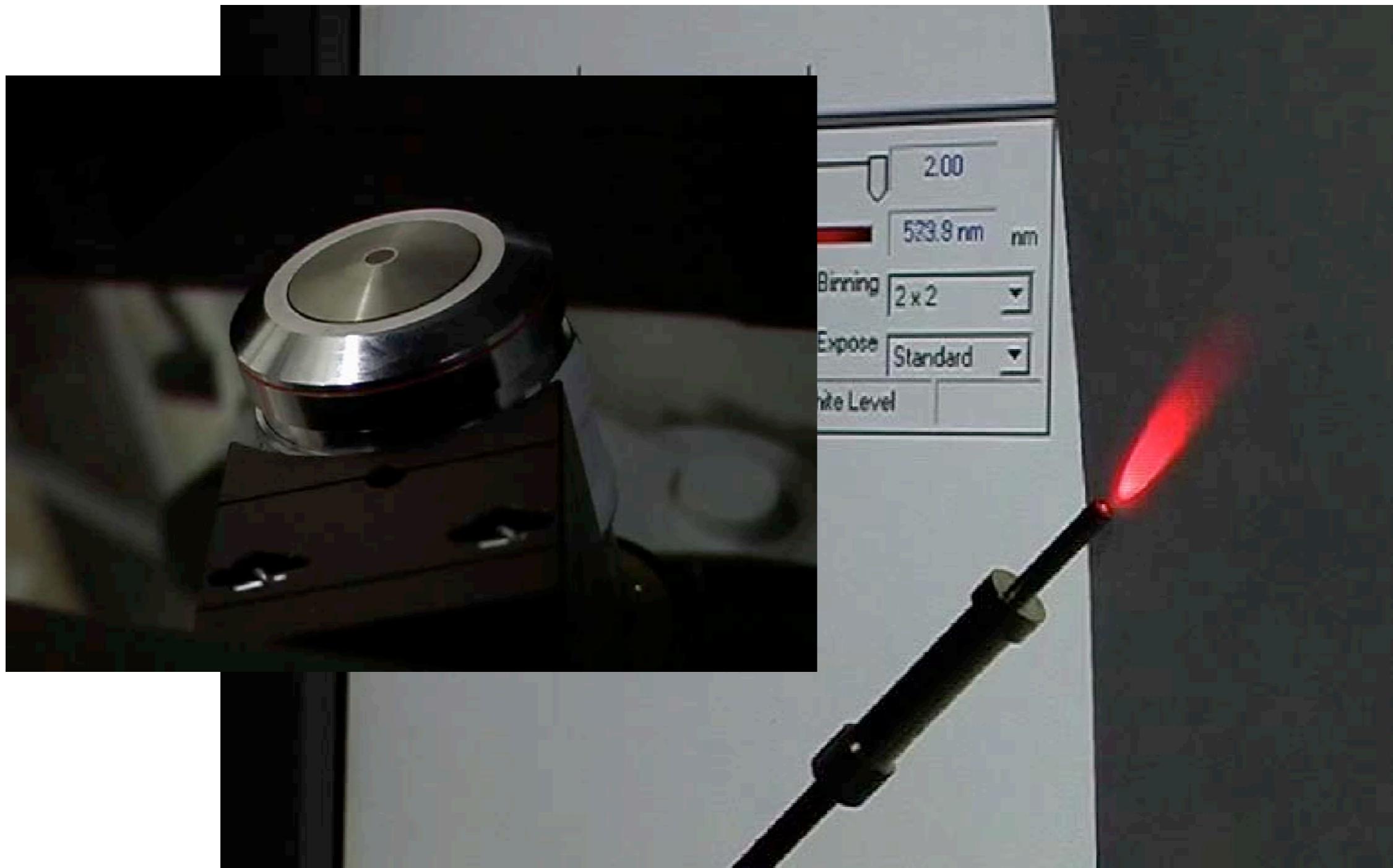
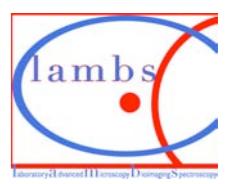
FLUORESCENCE

Perrin-Jablonski Diagram



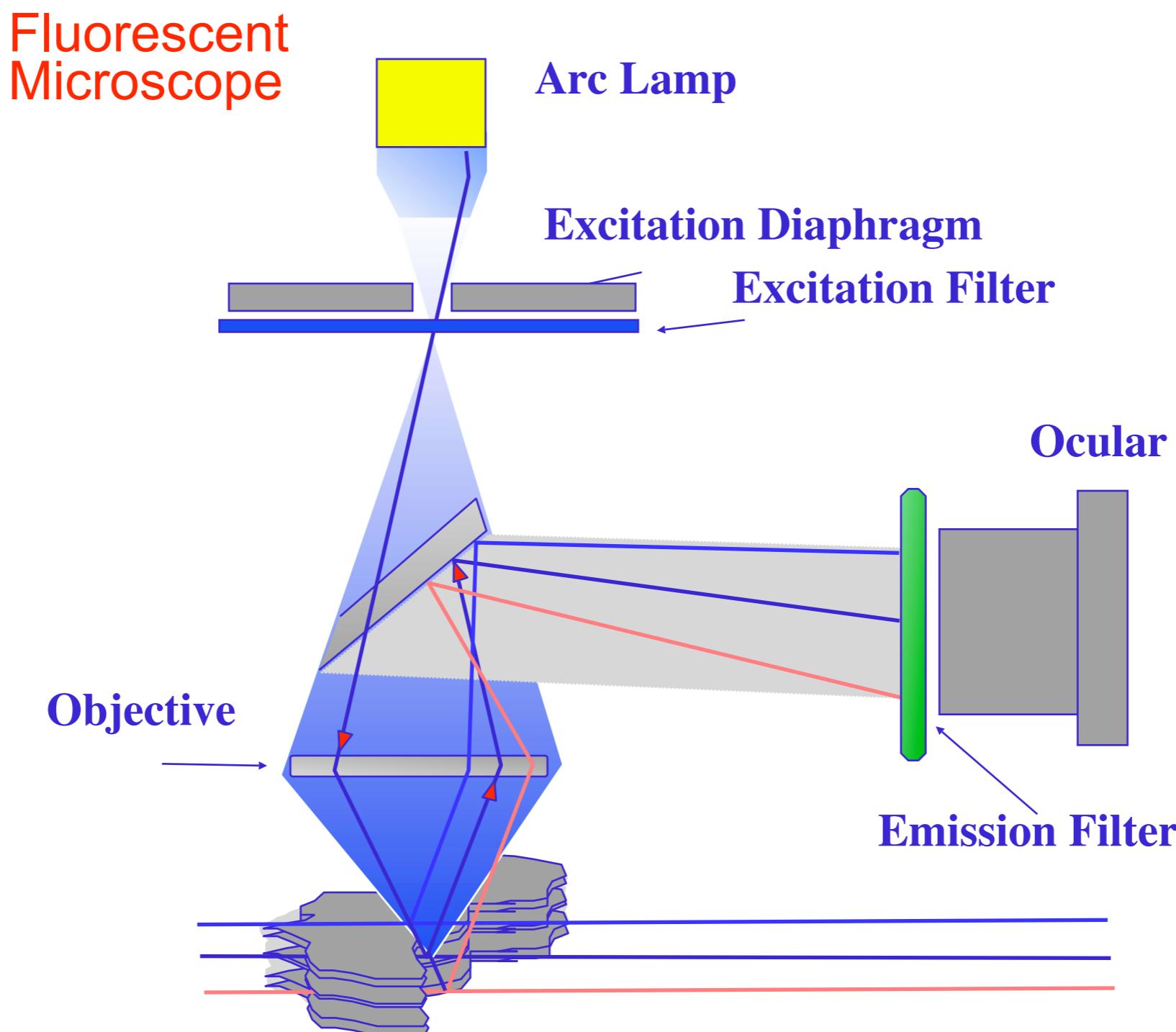
Slide Credit: Paul Robinson, Purdue and Paolo Bianchini, LAMBS

THE OPTICAL MICROSCOPE



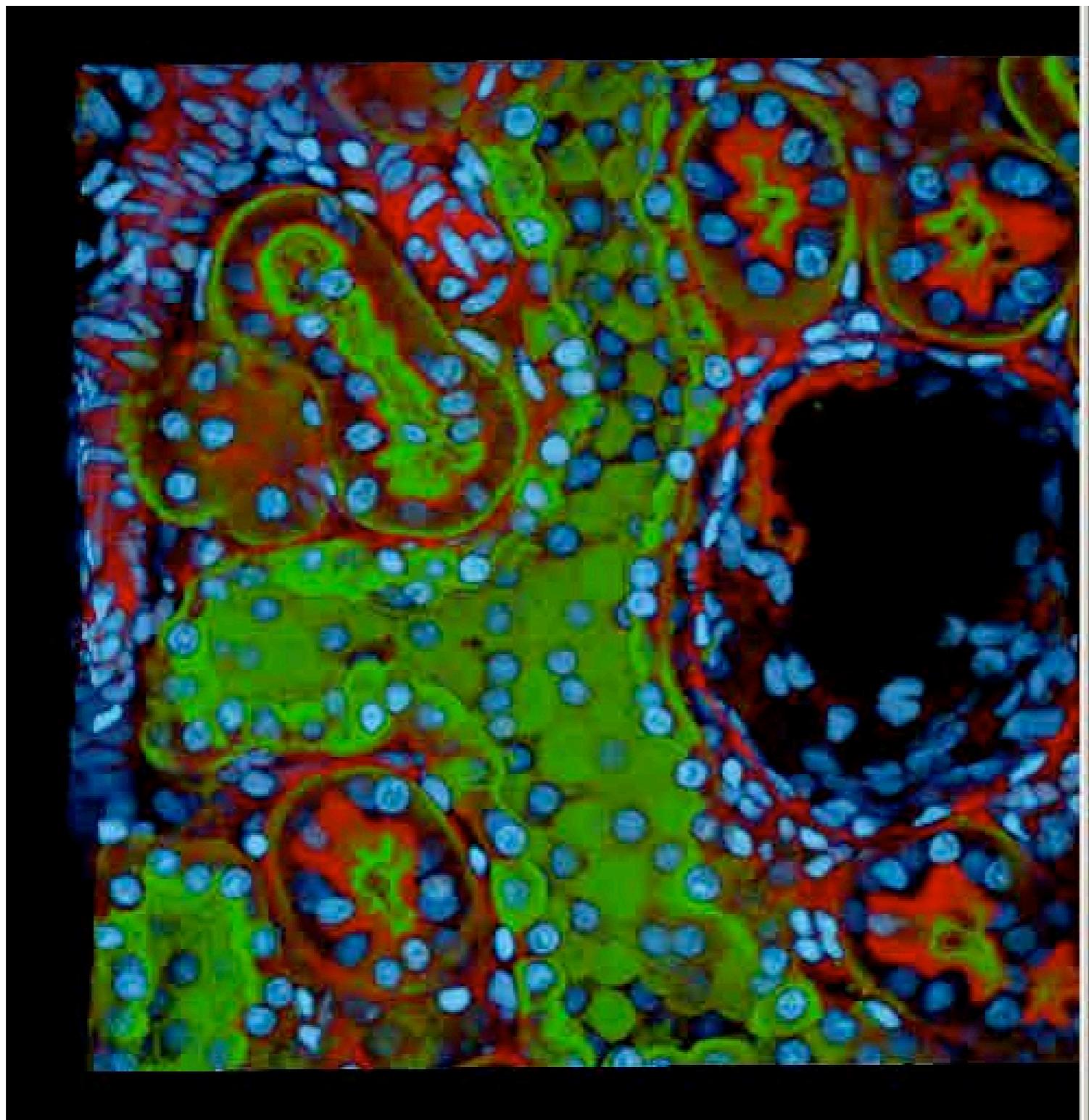
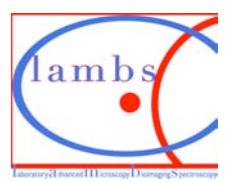
Credit: Leica Microsystems, Mannheim, Germany

THE OPTICAL MICROSCOPE



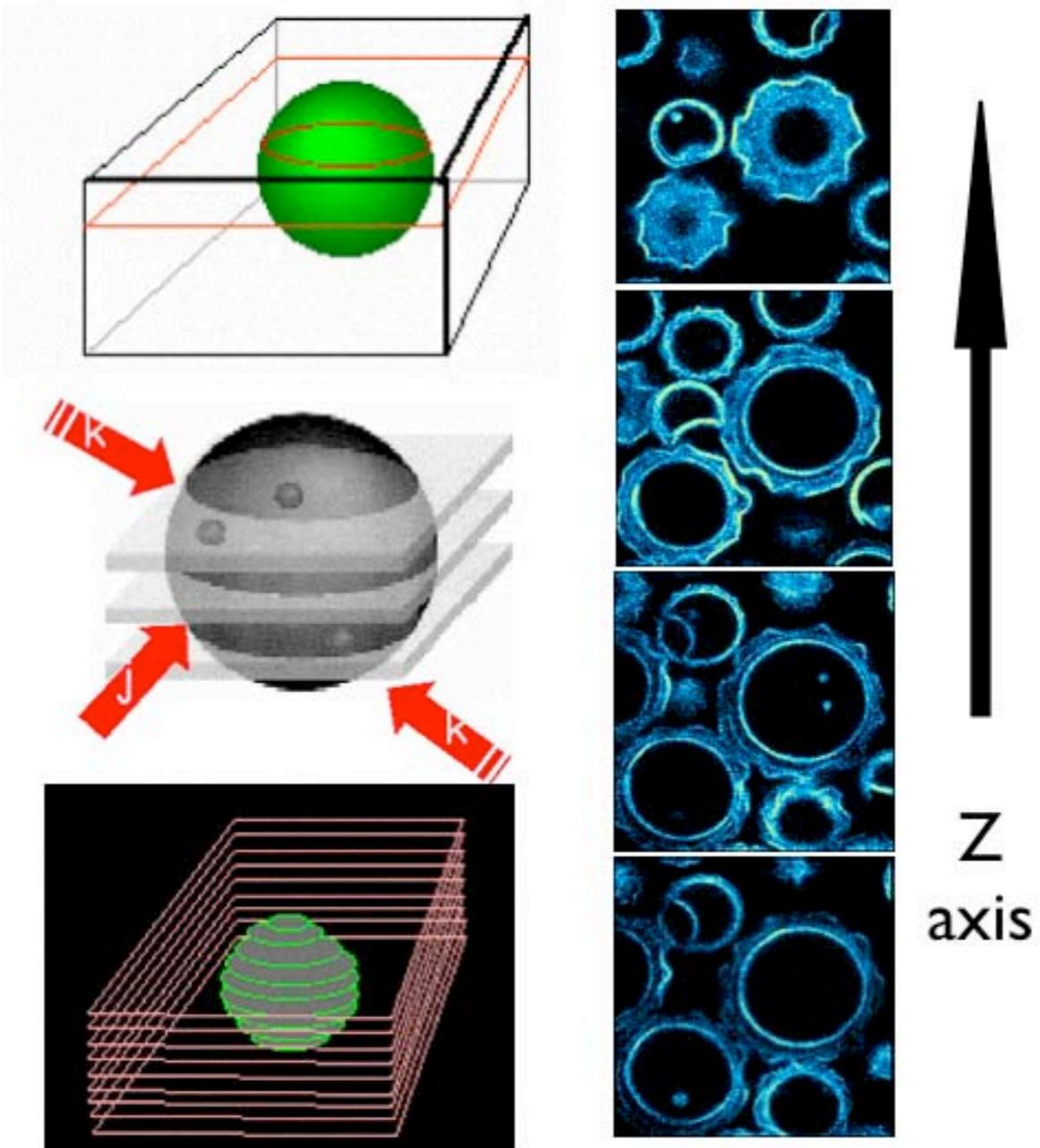
(COURTESY: PAUL ROBINSON, PURDUE UNIVERSITY)

3D OPTICAL MICROSCOPY



INDIANA
CENTER
for
BIOLOGICAL
MICROSCOPEY

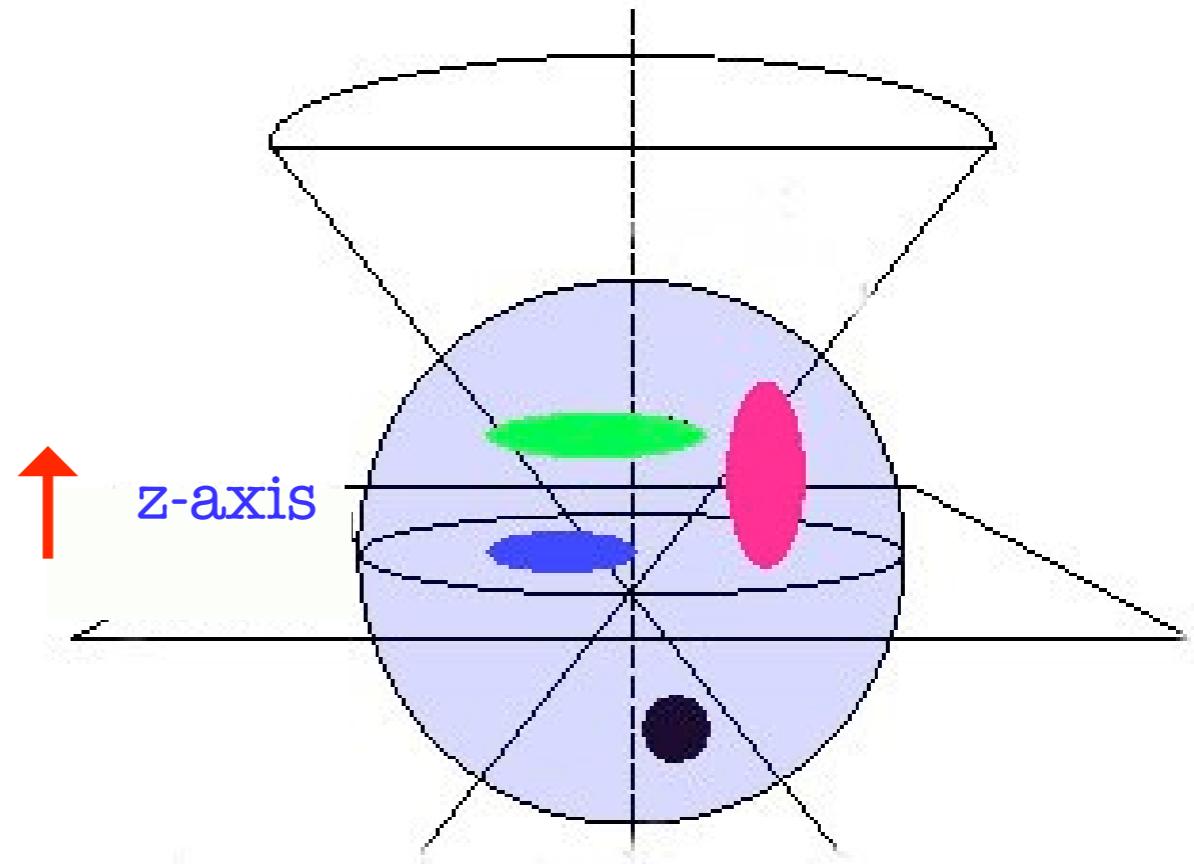
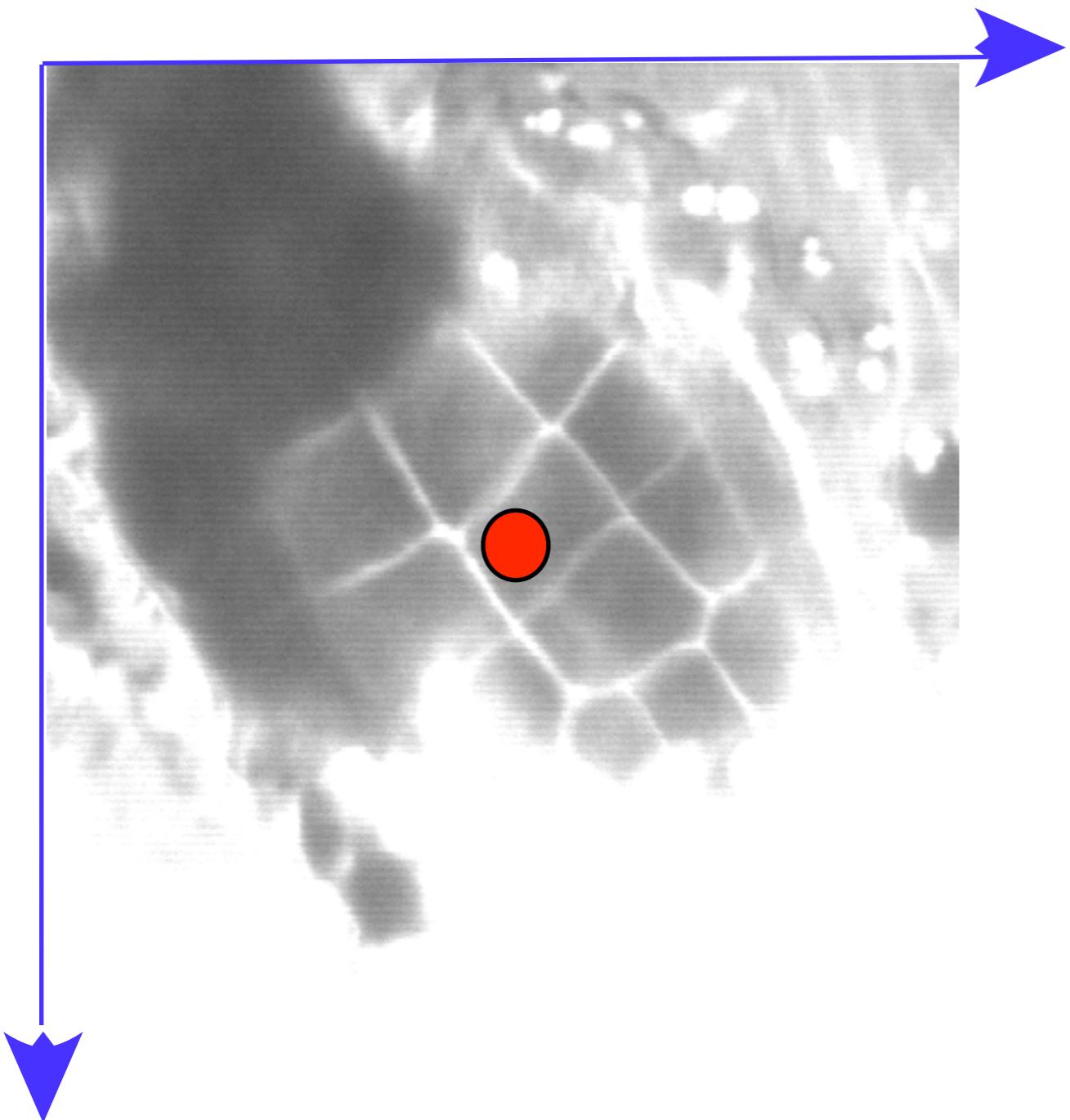
OPTICAL SECTIONING



B. BIANCO, A. DIASPRO, CELL BIOPHYSICS, 15 (3), PP.189-200, 1989.
A. DIASPRO, ET AL., IMAGE VISION AND COMPUTING, 8 (2), PP.130-141, 1990.

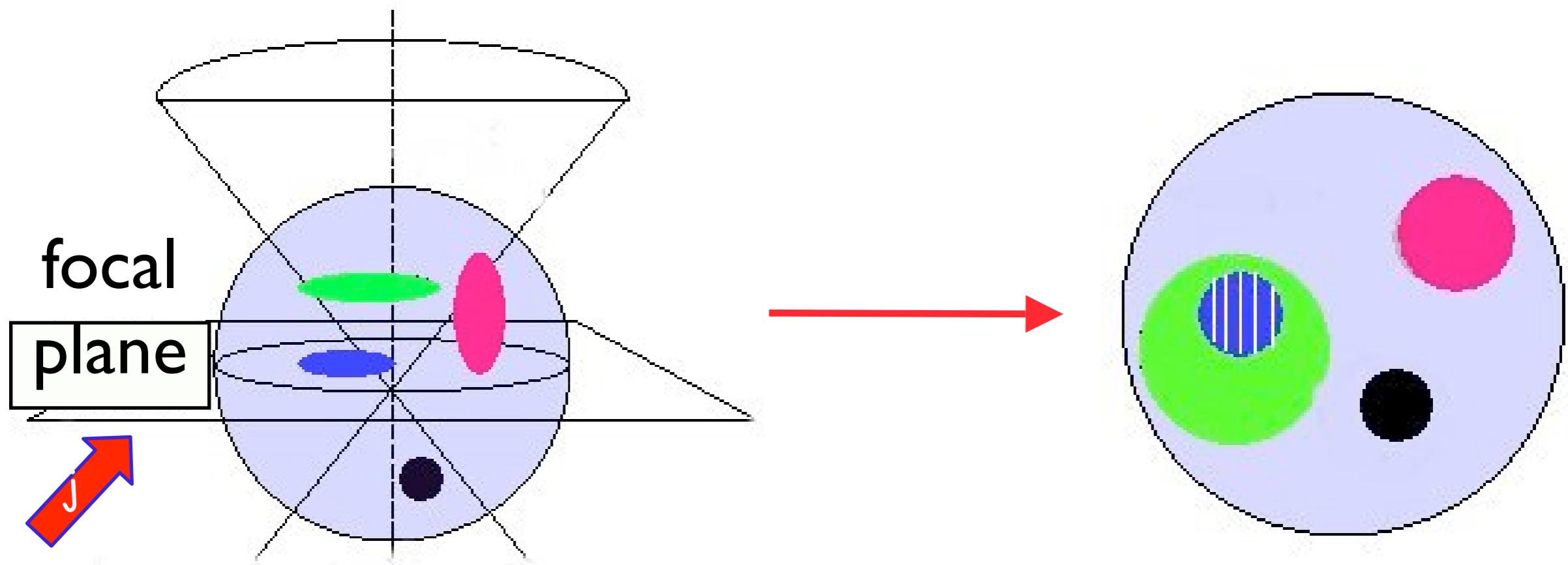
A.DIASPRO, G.CHIRICO, M.COLLI (2004) QUART. REV. .BIOPHYS.,VOL.38, NR.2, PP.1-72 (2006).

NAVIGATING INTO CELLS AND TISSUES



Vignette credit: Giuseppe Vicidomini, LAMBS-MicroScoBio, Genova

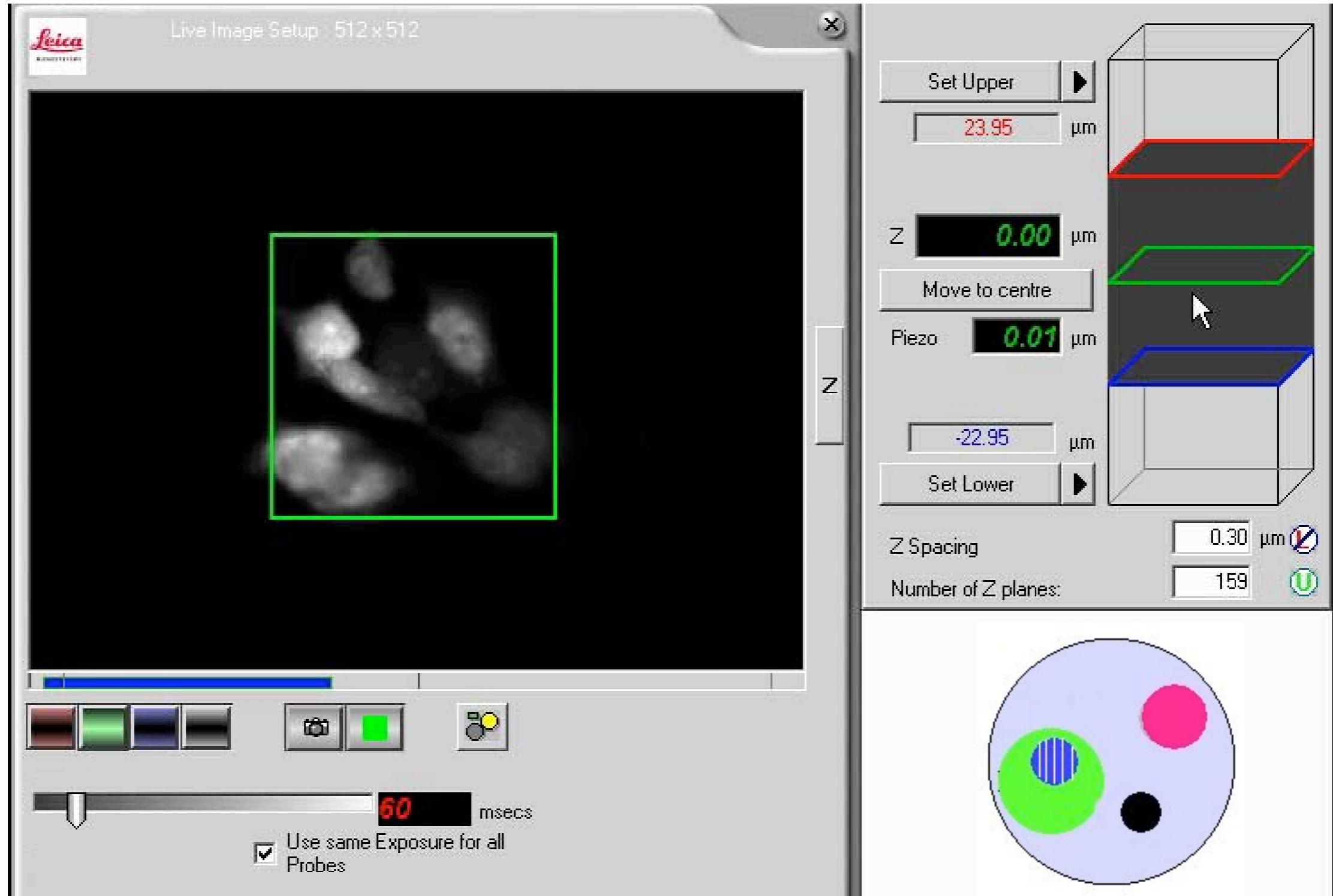
OPTICAL SECTIONING



B. Bianco, A. Diaspro, Cell Biophysics, 15 (3), pp.189-200, 1989.
A. Diaspro, et al., Image Vision and Computing, 8 (2), pp.130-141, 1990.

Slide credit: Giuseppe Vicedomini, LAMBS-MicroScoBio, Genova

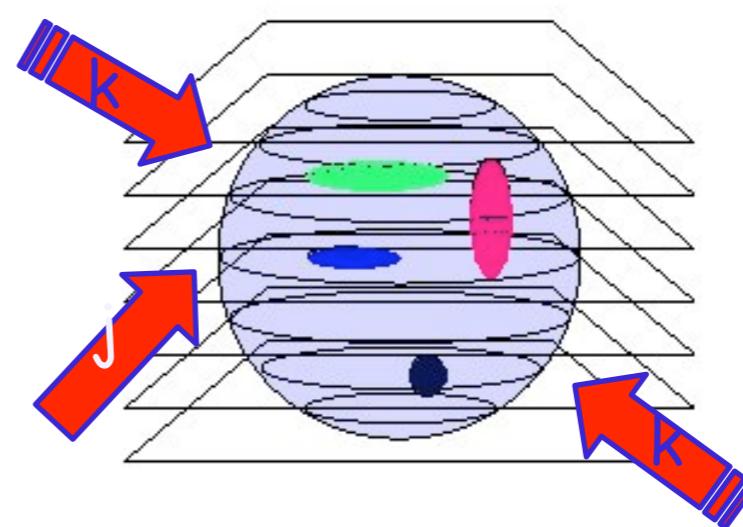
OPTICAL SECTIONING



OPTICAL SECTIONING

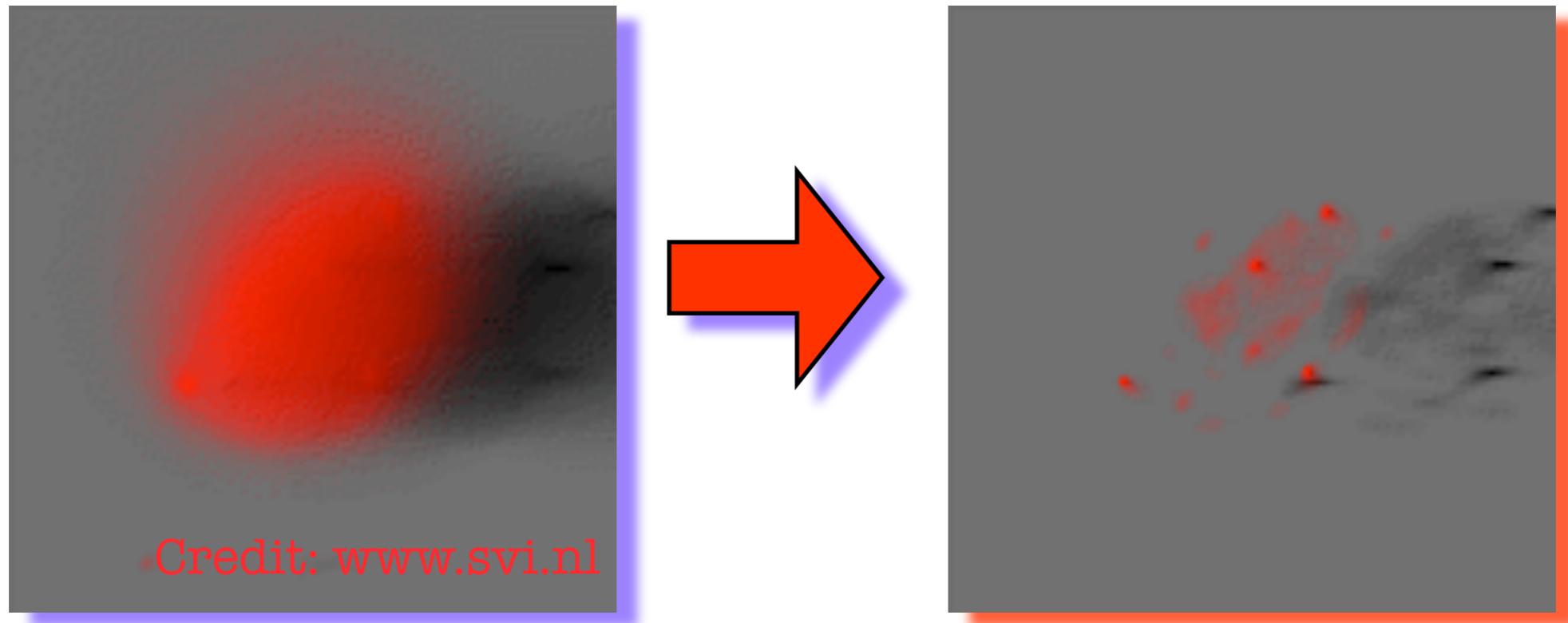
Within the optical sectioning scheme the situation is that the observed image O at a plane j is produced by the true fluorescence distribution at plane j , distorted by the microscope through S , plus contributions from adjacent k planes and noise N .

$$O_j = I_j S_j + \sum_{k \neq j} I_k S_k + N$$



Solving the equation set...

$$O_j = I_j S_j + \sum_{k \neq j} I_k S_k + N$$



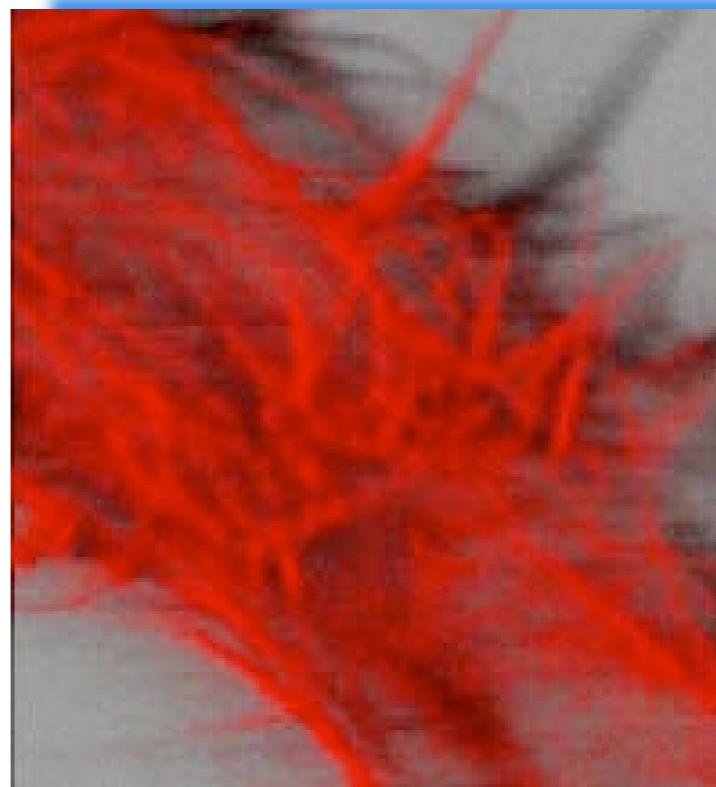
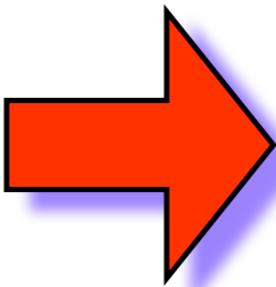
$$h(x, y, z)^* f(x, y, z) + n(x, y, z) + b(x, y, z) = g(x, y, z)$$

www.powermicroscope.com

BONETTO P., BOCCACCI P., SCARITO M., DAVOLIO M., EPIFANI M., VICIDOMINI G., TACCHETTI C., RAMOINO P., USAI C., DIASPRO A.
(2004) MICROSCOPY RESEARCH AND TECHNIQUE. vol. 64, pp. 196-203.

Solving the equation set...

$$O_j = I_j S_j + \sum_{k \neq j} I_k S_k + N$$



$$h(x, y, z)^* f(x, y, z) + n(x, y, z) + b(x, y, z) = g(x, y, z)$$

www.powermicroscope.com

BONETTO P., BOCCACCI P., SCARITO M., DAVOLIO M., EPIFANI M., VICIDOMINI G., TACCHETTI C., RAMOINO P., USAI C., DIASPRO A.
(2004) MICROSCOPY RESEARCH AND TECHNIQUE. vol. 64, pp. 196-203.

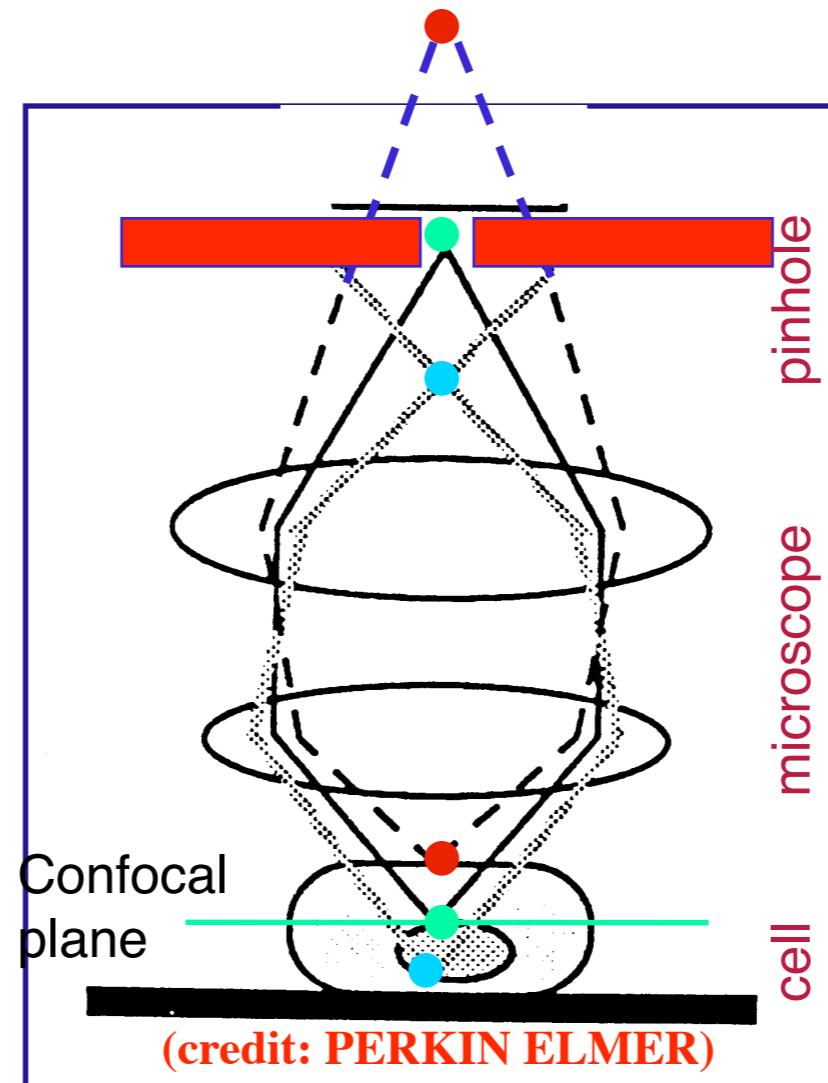
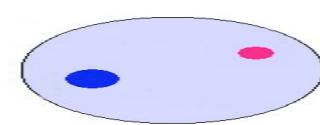
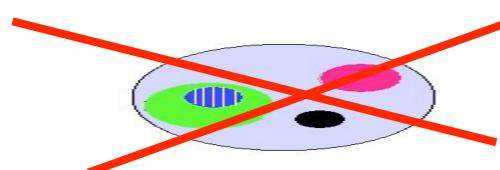
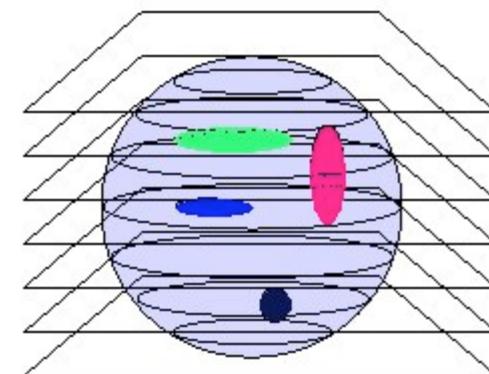
CONFOCAL MICROSCOPY

Solving the equation set...by means of a confocal set-up

$$\sum I_k S_k \rightarrow 0$$

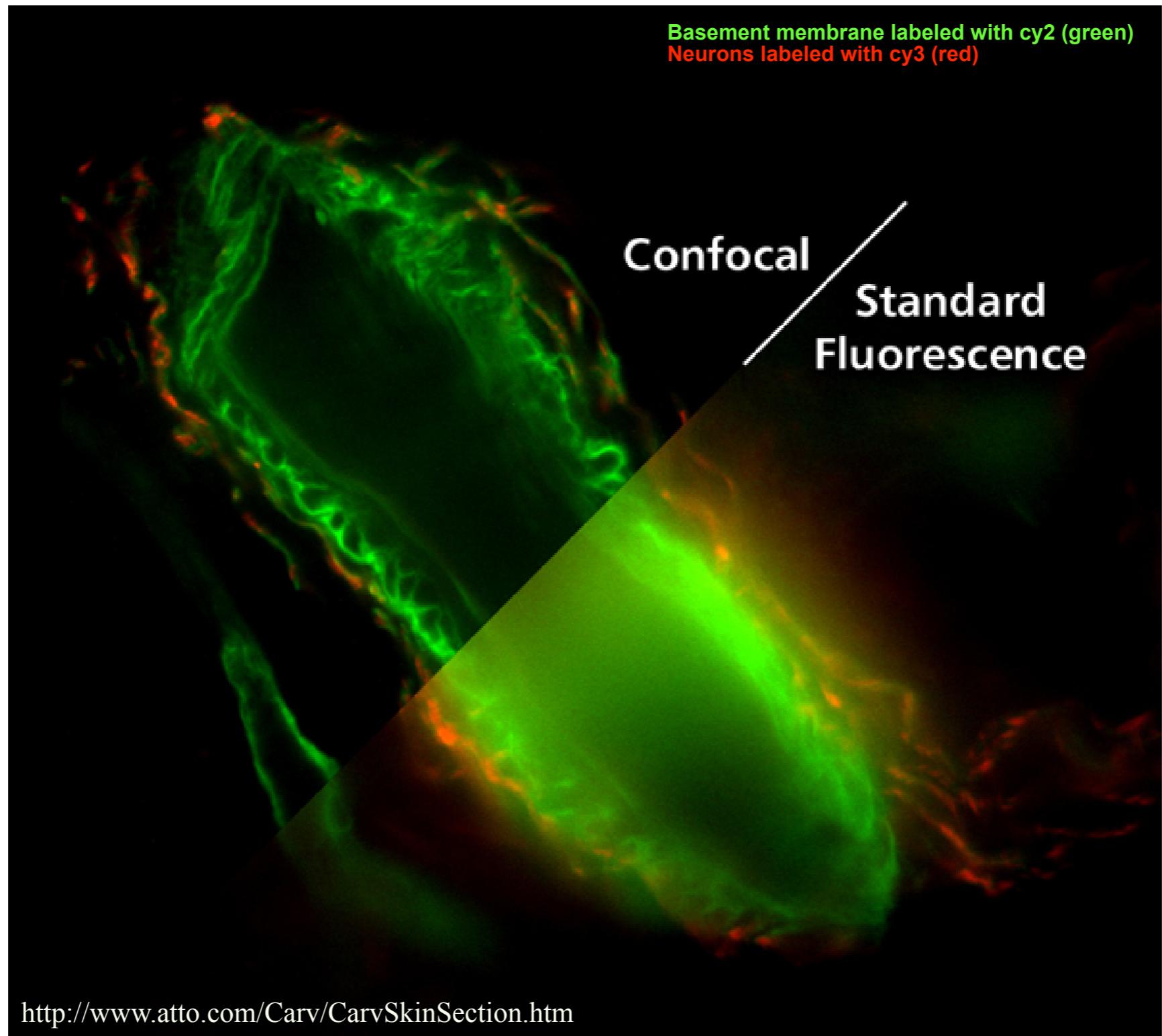
$$S_j \rightarrow 1$$

$$O_j \rightarrow I_j$$

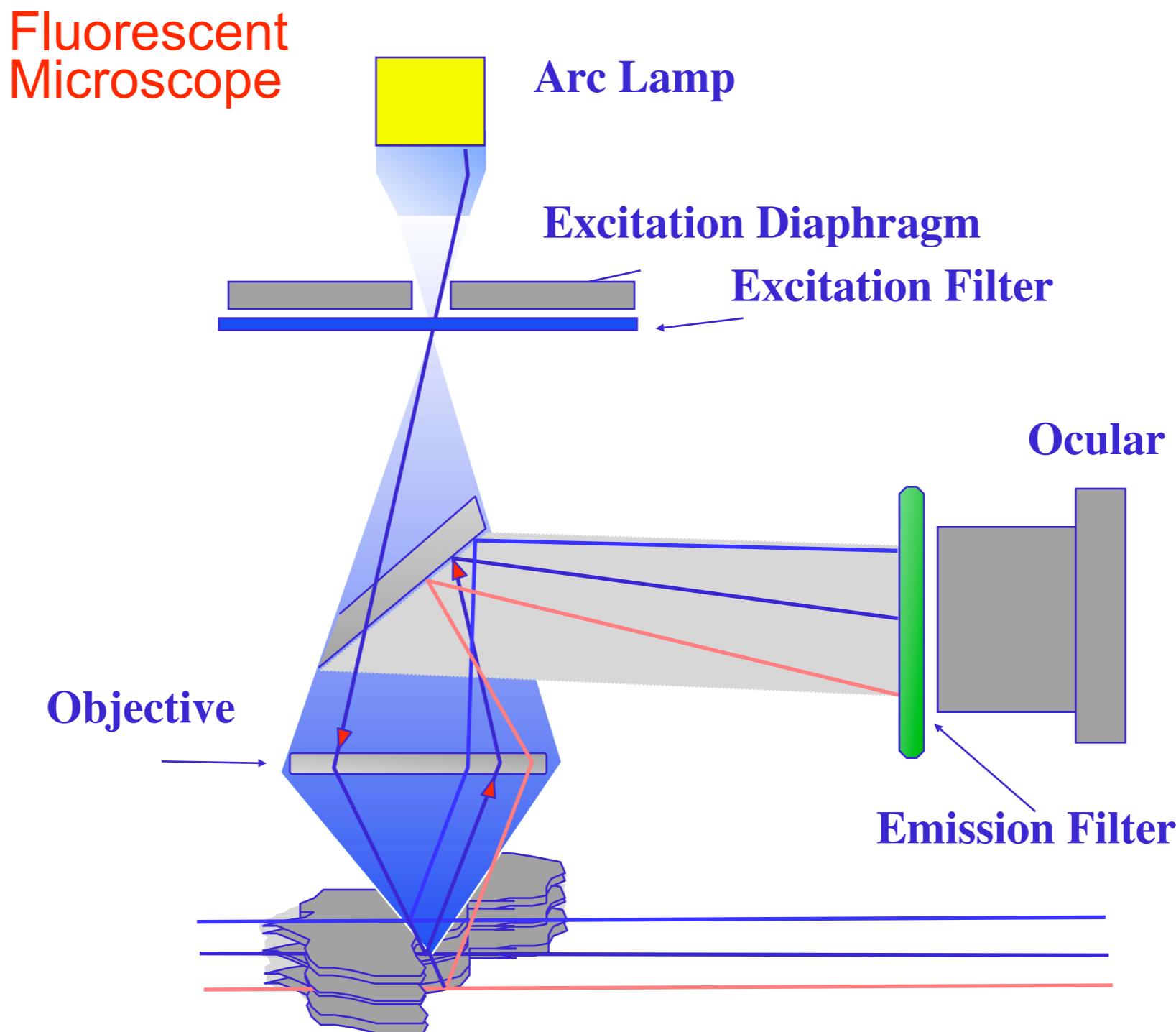


$$O_j = I_j S_j + \sum_{k \neq j} I_k S_k + N$$

CONFOCAL MICROSCOPY

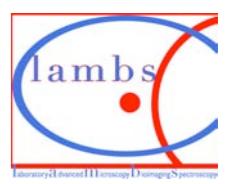


THE OPTICAL MICROSCOPE

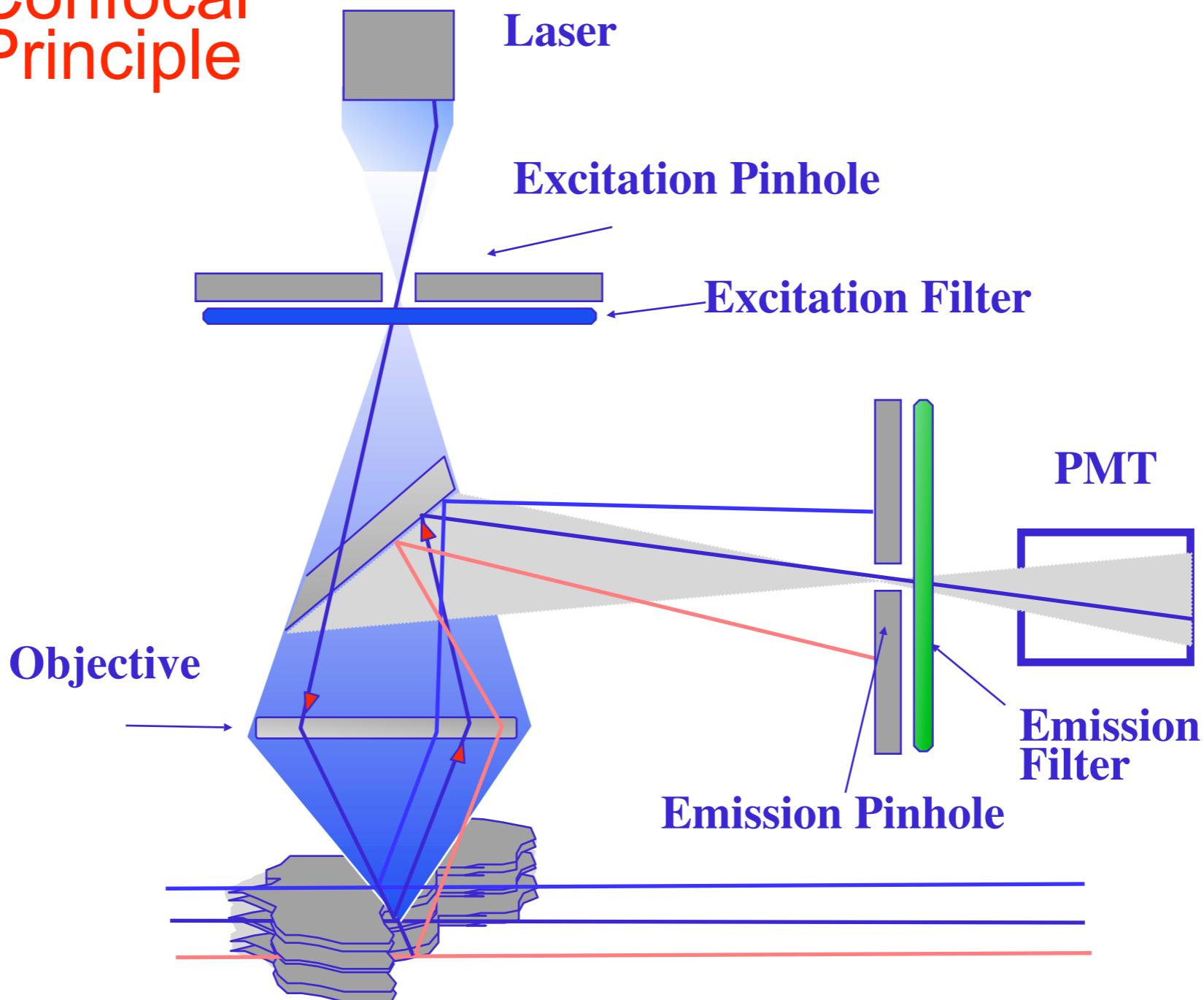


(COURTESY: PAUL ROBINSON, PURDUE UNIVERSITY)

THE OPTICAL MICROSCOPE



Confocal Principle



(COURTESY: PAUL ROBINSON, PURDUE UNIVERSITY)

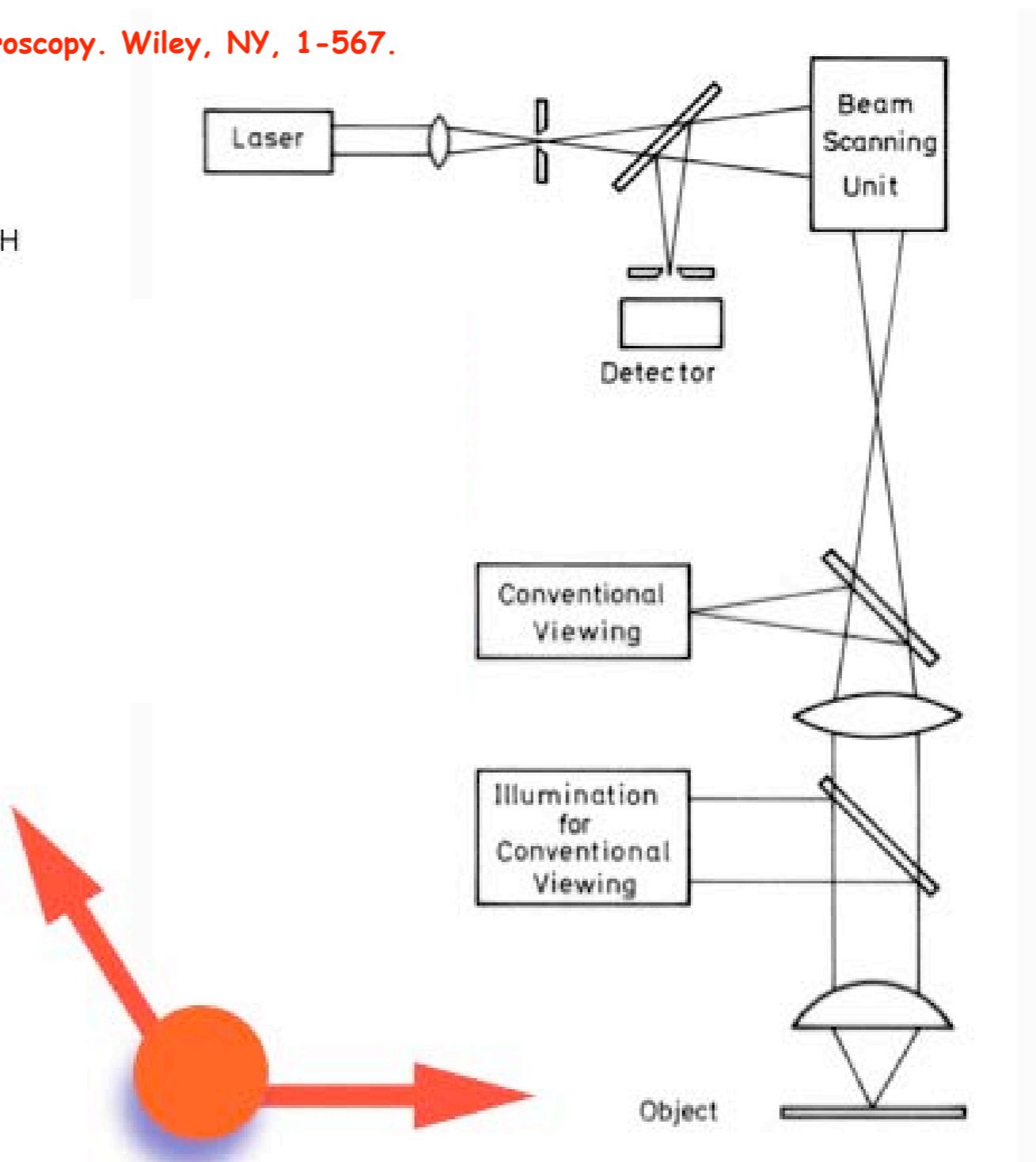
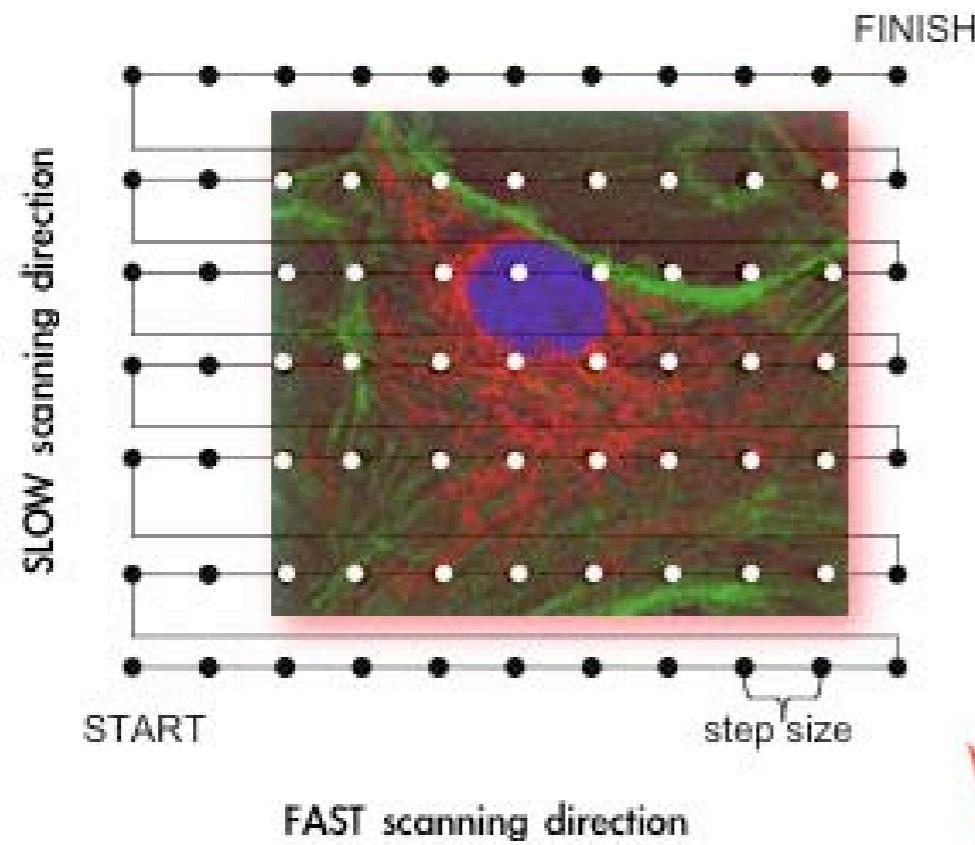
THE BILL...



(GOYA, Blindman's buff; credit:BRUNO SAMORI')

POINT BY POINT IMAGING

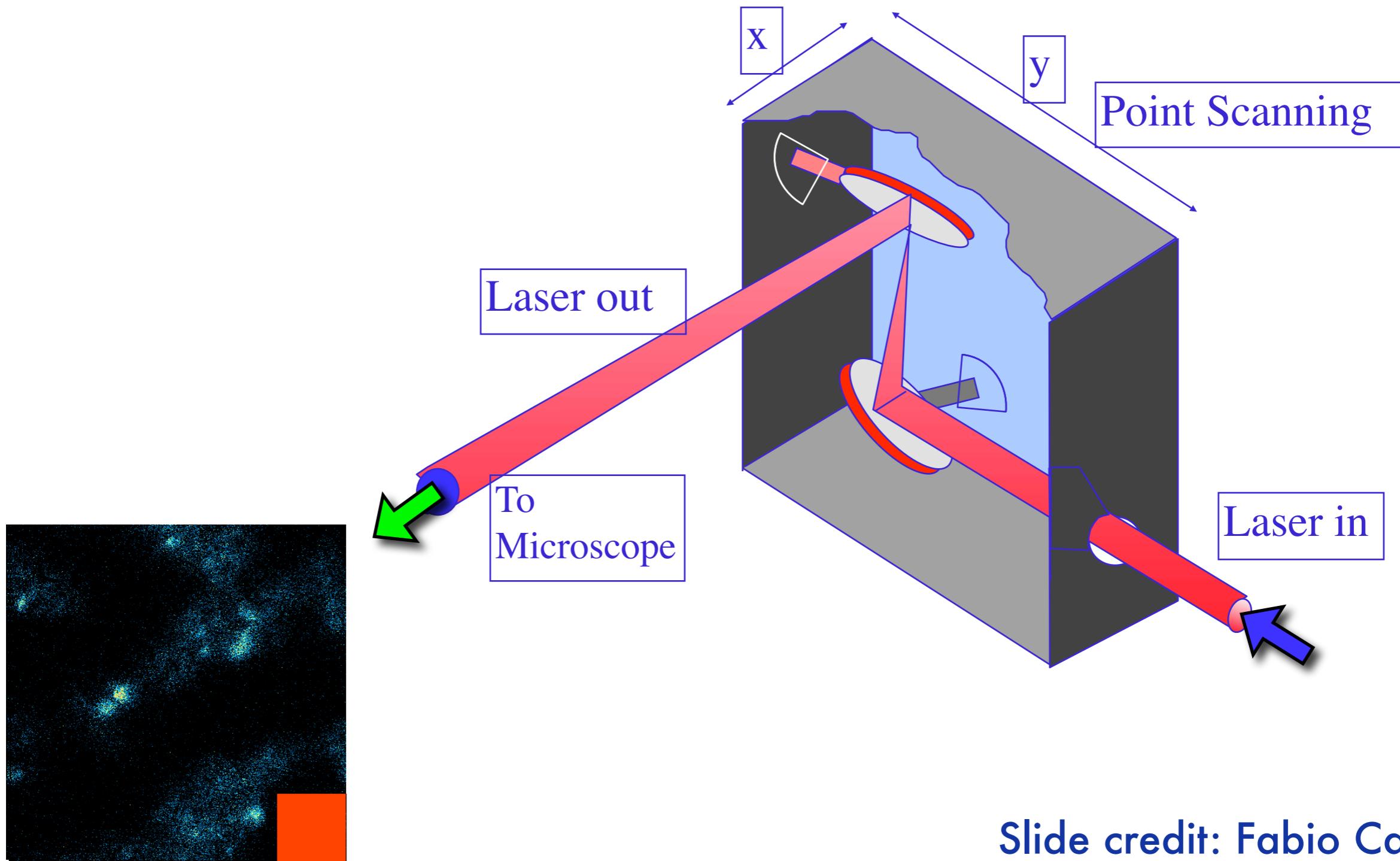
A. Diaspro (ed.), (2001) "Confocal and Two-photon Microscopy. Wiley, NY, 1-567.



POINT BY POINT IMAGING

LIKE A FLORIAN'S COFFE IN VENICE

2 ms/line

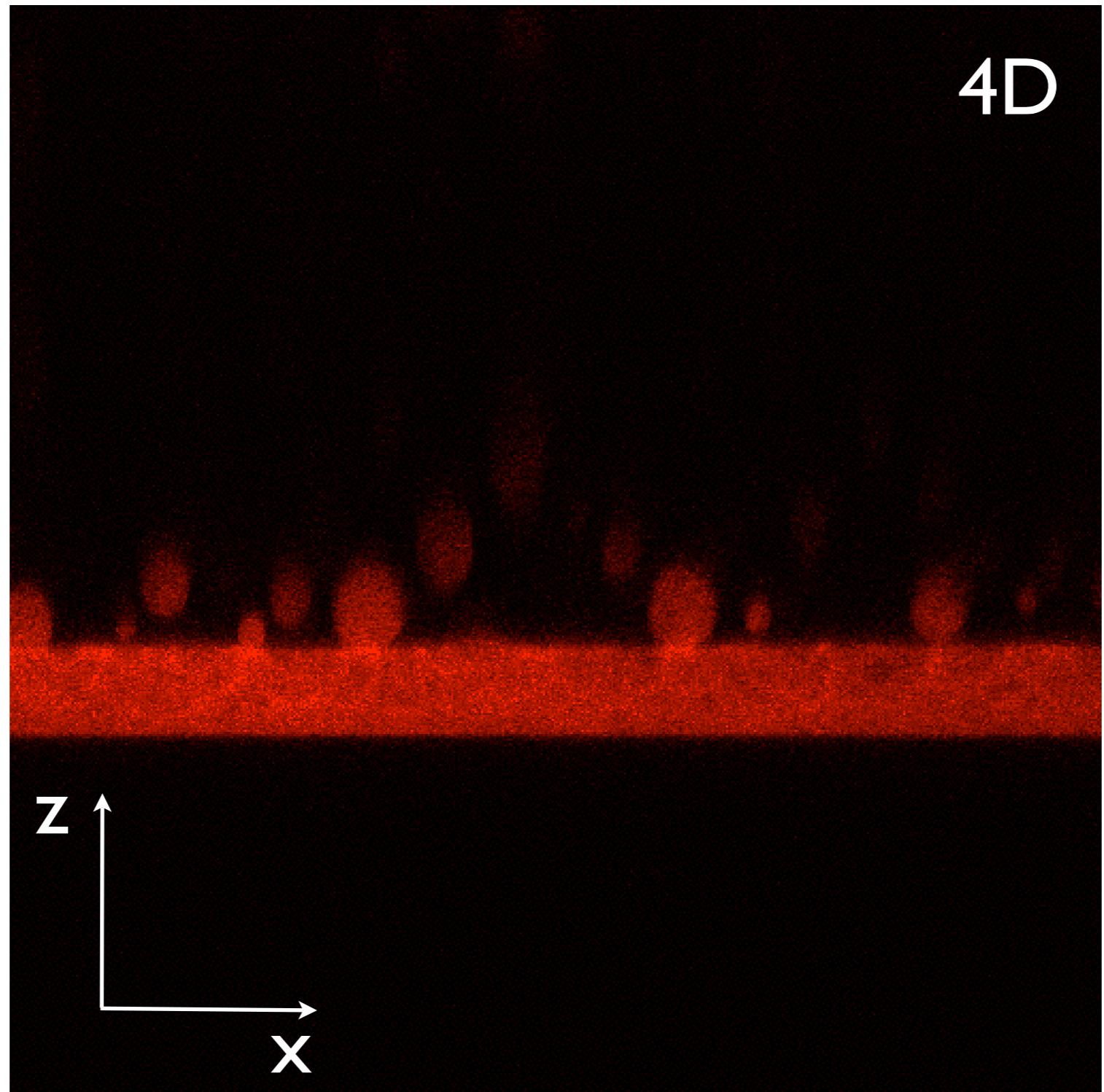


Slide credit: Fabio Cannone

POINT BY POINT IMAGING

LIKE A FLORIAN'S COFFE IN VENICE

2 ms/line



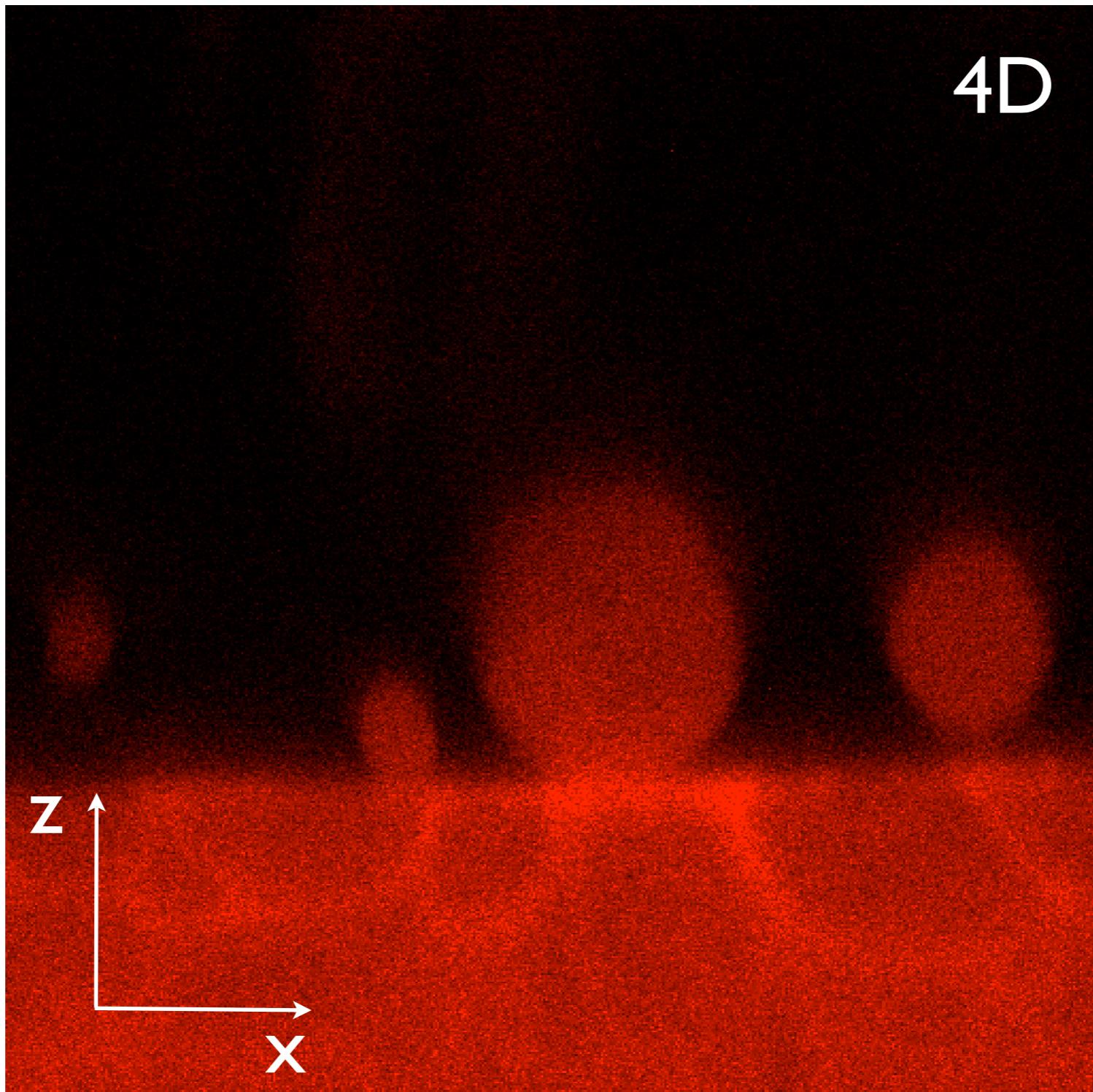
Slide credit:Davide Mazza, LAMBS-MicroScoBio, Genova

POINT BY POINT IMAGING

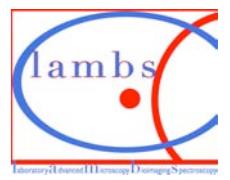
LIKE A FLORIAN'S COFFE IN VENICE

2 ms/line

4D

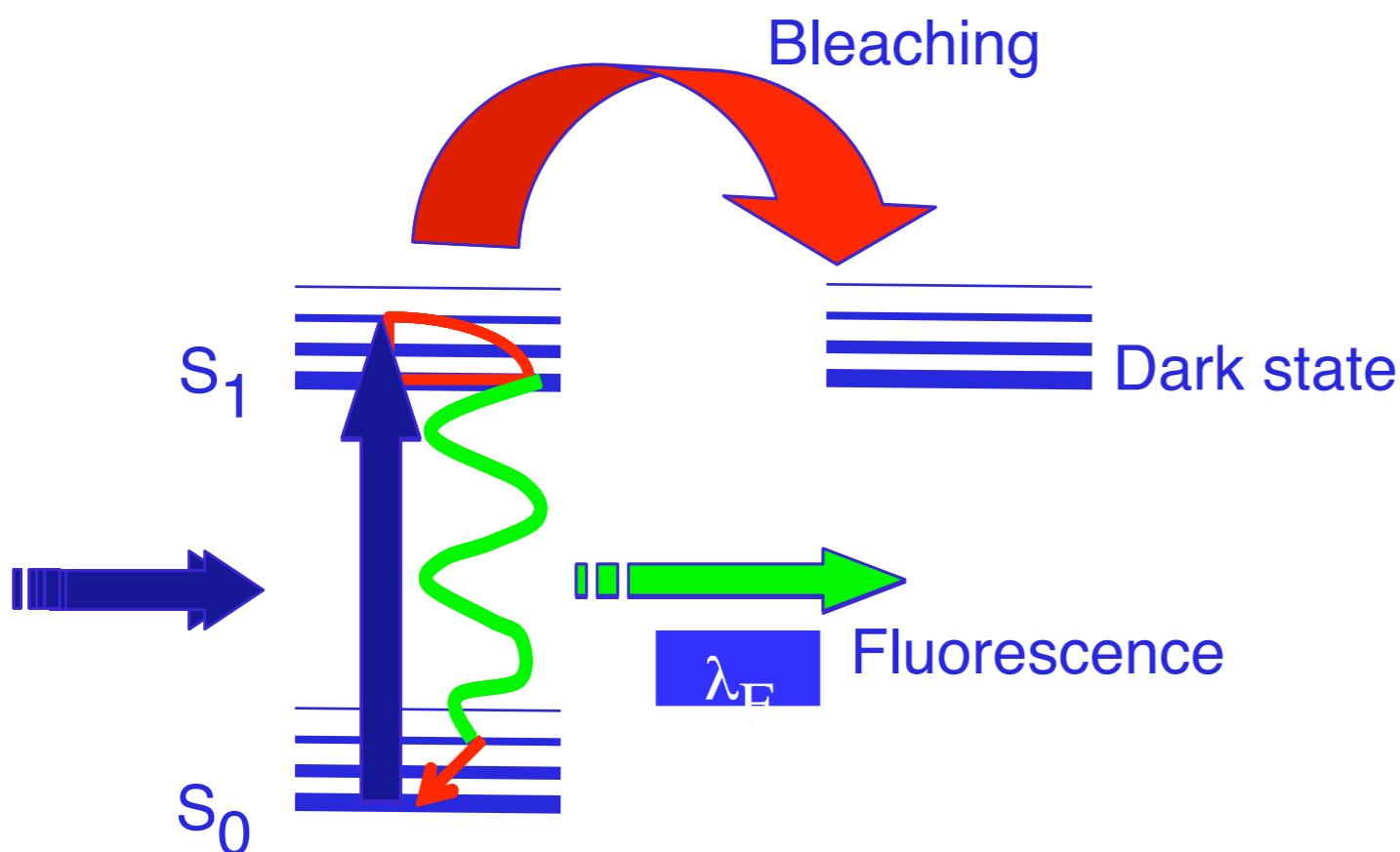


POINT BY POINT IMAGING



Giuseppe Pellizza da Volpedo (1868-1907)
Panni al sole (1894), olio su tela, 87x131 cm, Domodossola, collezione privata

PHOTOBLEACHING PROBLEM

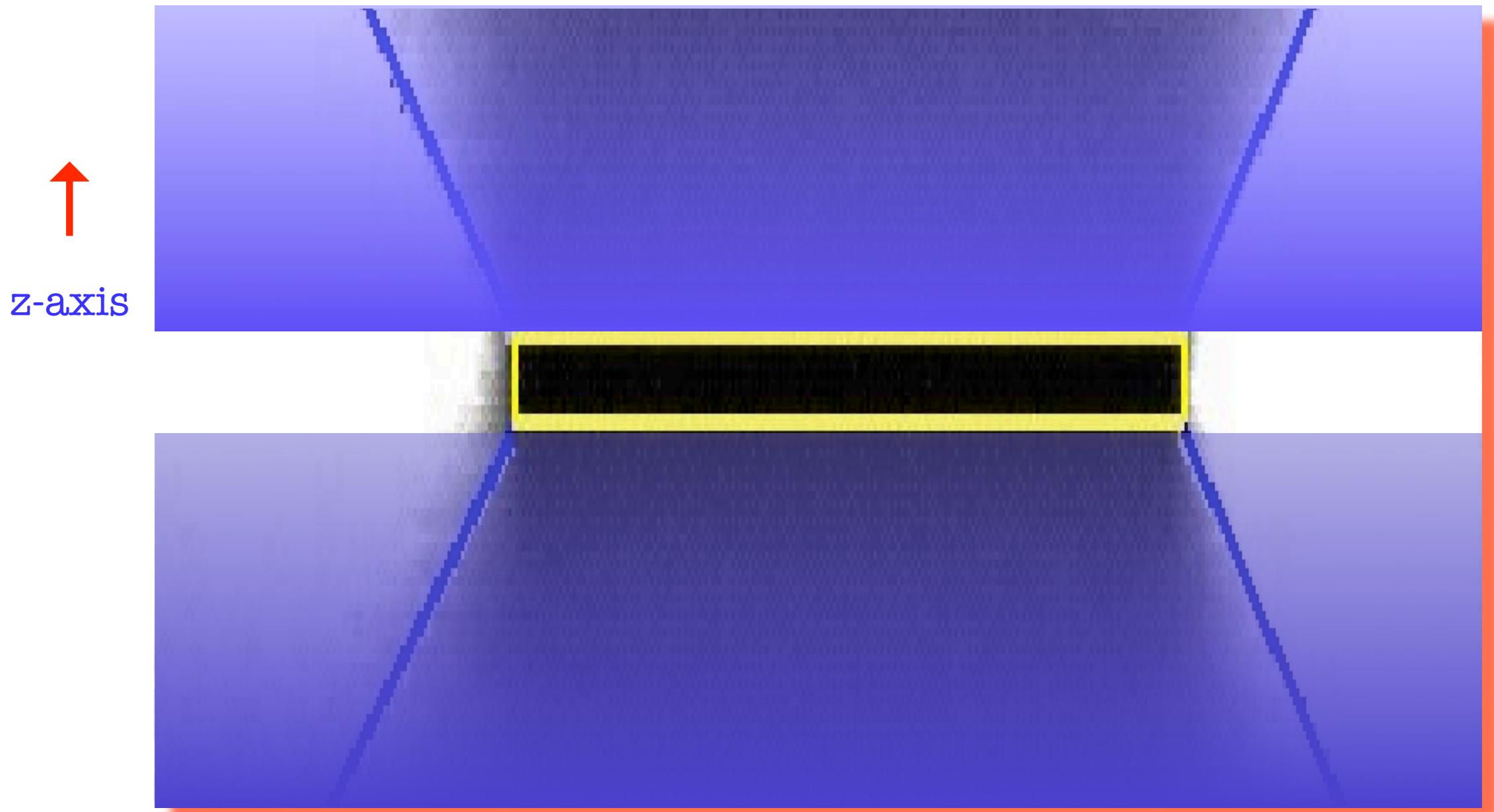
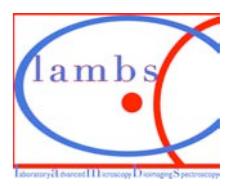


- Photochemical lifetime: fluorescein will undergo 30-40,000 emissions before bleaching.
- At low excitation intensities, pb occurs but at lower rate.
- Bleaching is often photodynamic-- involves light and oxygen.

Diaspro A. et al., Photobleaching, in Handbook of Confocal Microscopy (J.Pawley ed.), Plenum, NEW edition, 2006

Slide credit: Giberto Chirico, Enrico Gratton

PHOTOBLEACHING PROBLEM



Diaspro A. et al., Photobleaching, in Handbook of Confocal Microscopy (J.Pawley ed.), Plenum, NEW edition, 2006

Image credit:David Piston, Vanderbilt University

TWO-PHOTON EXCITATION MICROSCOPY



A.Diaspro (ed.), (2001) "Confocal and Two-photon Microscopy. Wiley, NY, 1-567.

TWO-PHOTON EXCITATION MICROSCOPY



A.Diaspro (ed.), (2001) "Confocal and Two-photon Microscopy. Wiley, NY, 1-567.

TWO-PHOTON EXCITATION MICROSCOPY



Giuseppe Pellizza da Volpedo (1868–1907)

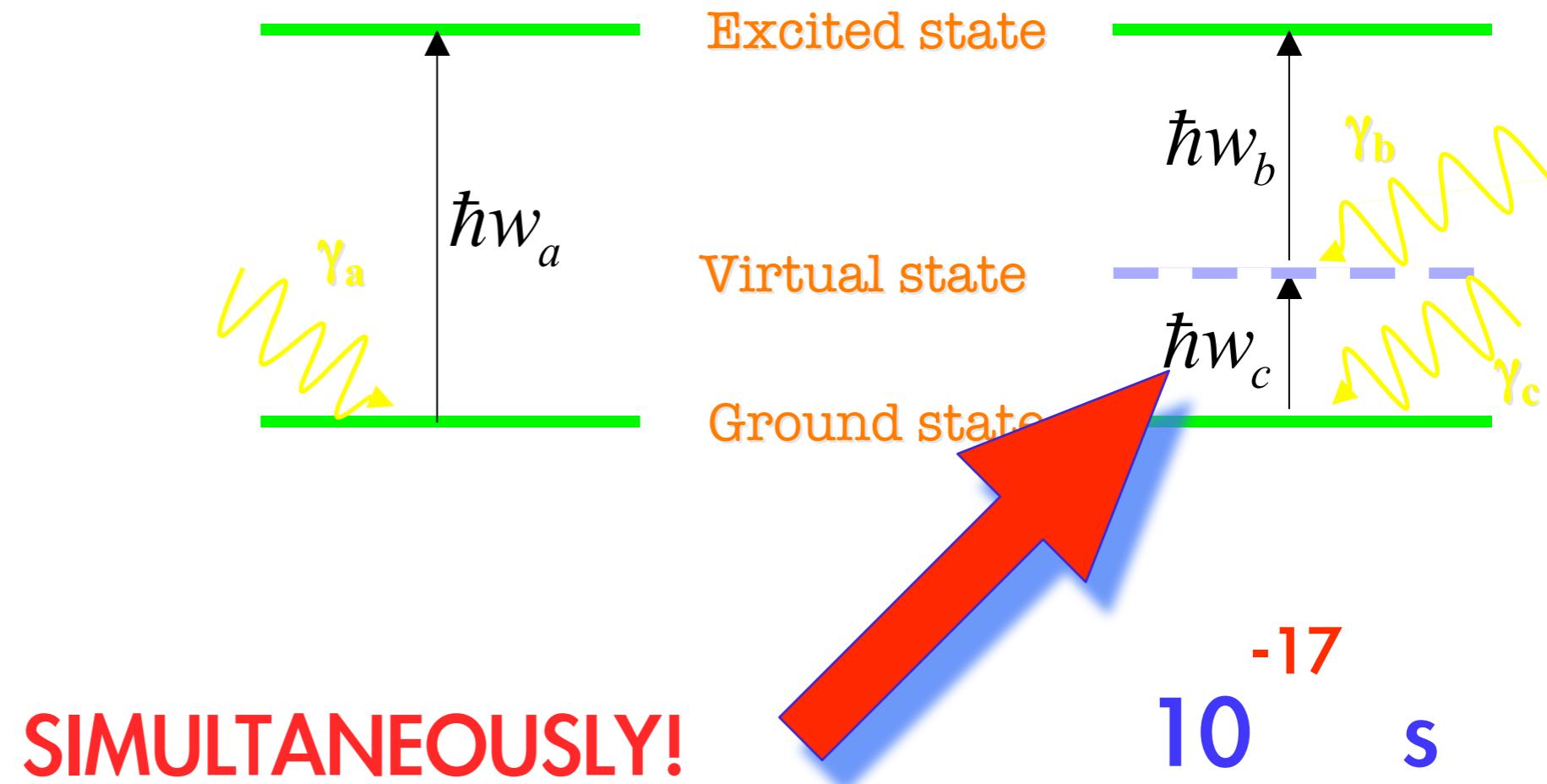
Il quarto stato (1901), olio su tela, 293x545 cm, Milano, Civica Galleria d'Arte Moderna

TWO-PHOTON EXCITATION MICROSCOPY



(Credit: CLAUDIA DIASPRO from JOVANOTTI'S POP CONCERT)

TWO-PHOTON EXCITATION MICROSCOPY



(credit: EMILIO SEGRE' archive)

Maria Göppert-Mayer predicted that an atom or a molecule could interact with two photons simultaneously by absorbing them in the very same quantum event.

TWO-PHOTON EXCITATION MICROSCOPY

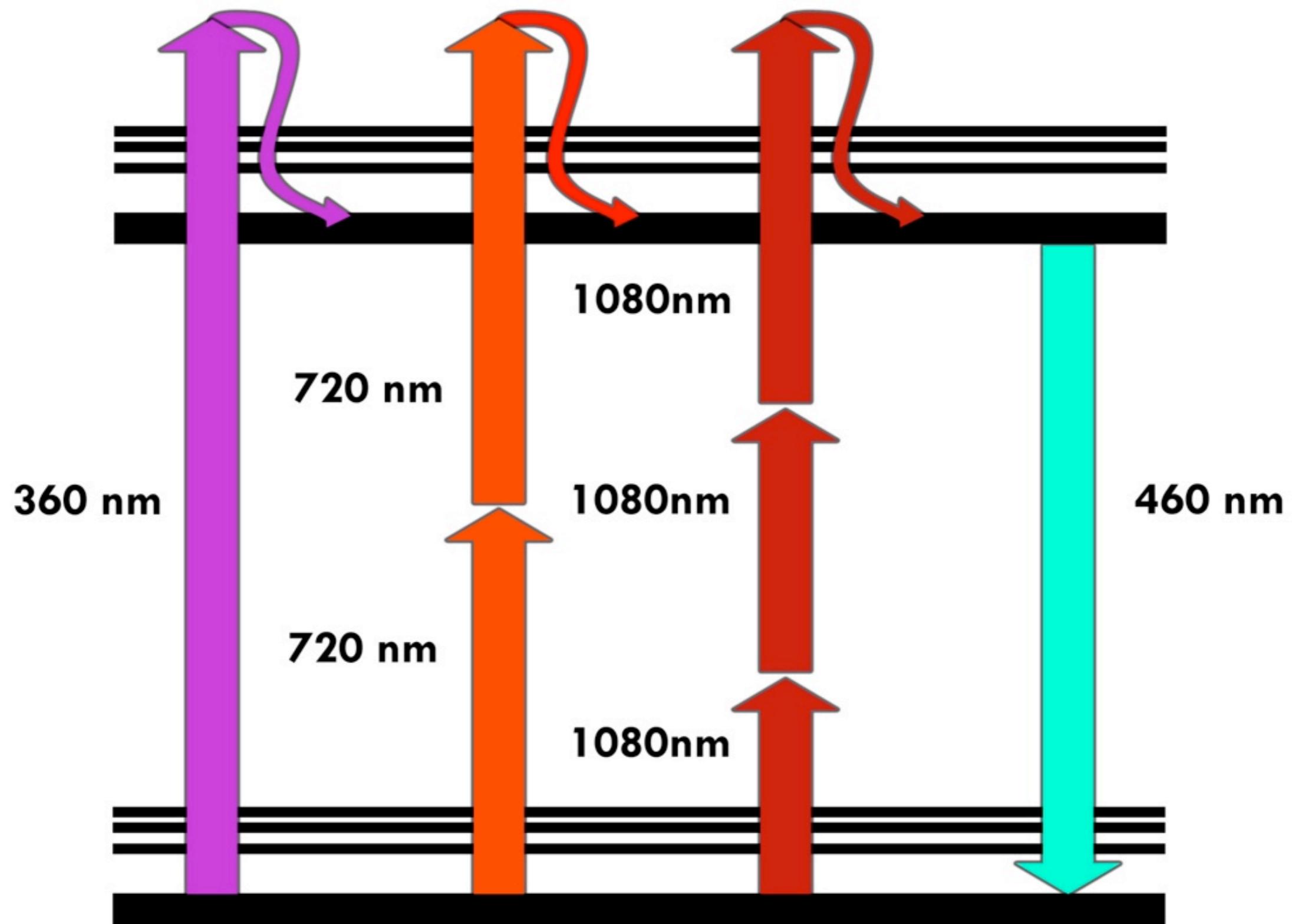


$$E_a = E_b + E_c$$

$$\lambda_a = \left(\frac{1}{\lambda_b} + \frac{1}{\lambda_c} \right)^{-1}$$



TWO-PHOTON EXCITATION MICROSCOPY



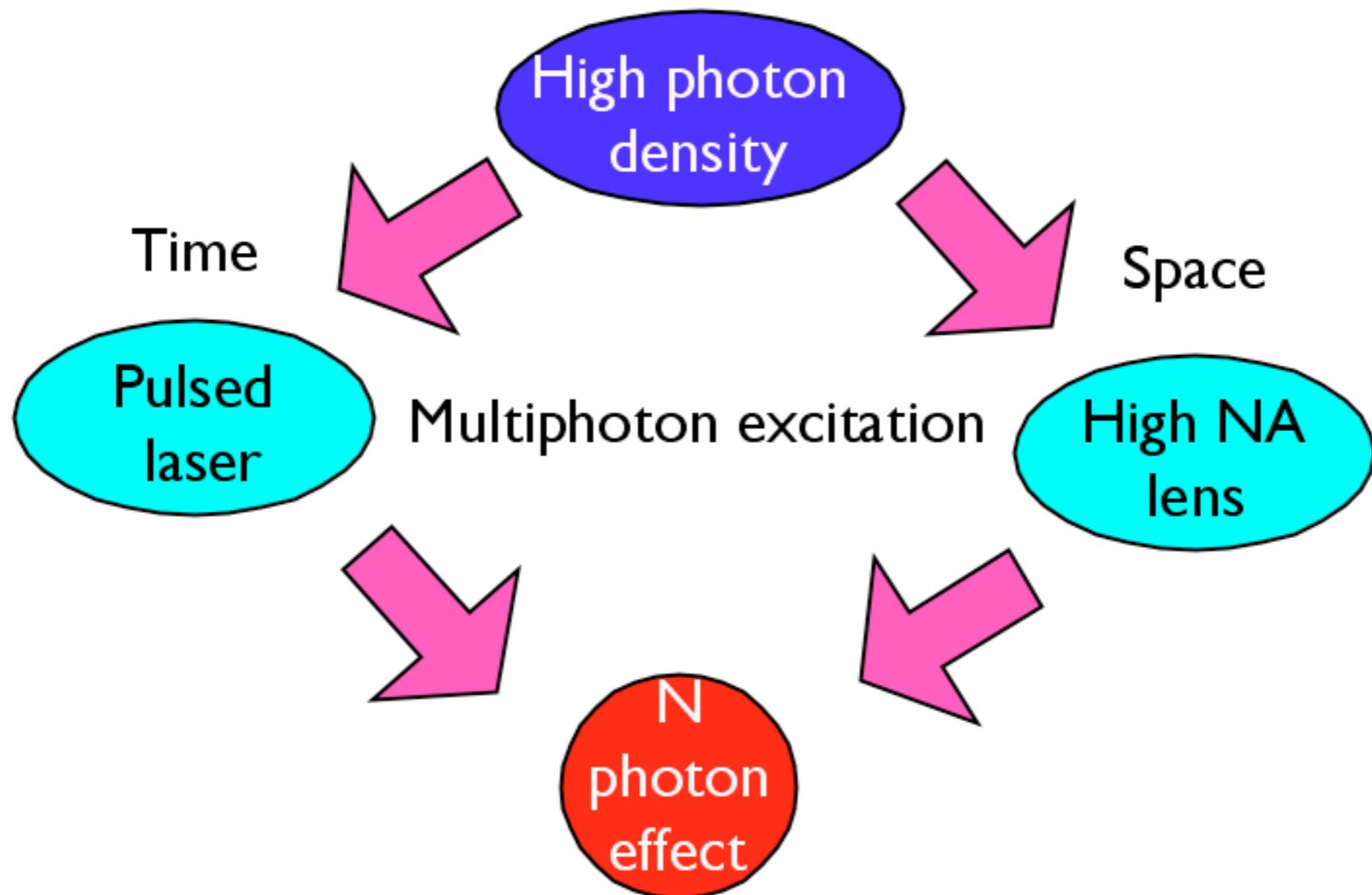
TWO-PHOTON EXCITATION MICROSCOPY



IN BRIGHT SUNLIGHT, A MOLECULE OF RHODAMINE B, AN EXCELLENT 1- OR 2- PHOTON ABSORBER, ABSORBS A PHOTON THROUGH A 1-PHOTON PROCESS ABOUT ONCE A SECOND, A PHOTON PAIR BY 2-PHOTON ABSORPTION EVERY 10 MILLION YEARS...

Denk and Svoboda (1997) Neuron, 18:351

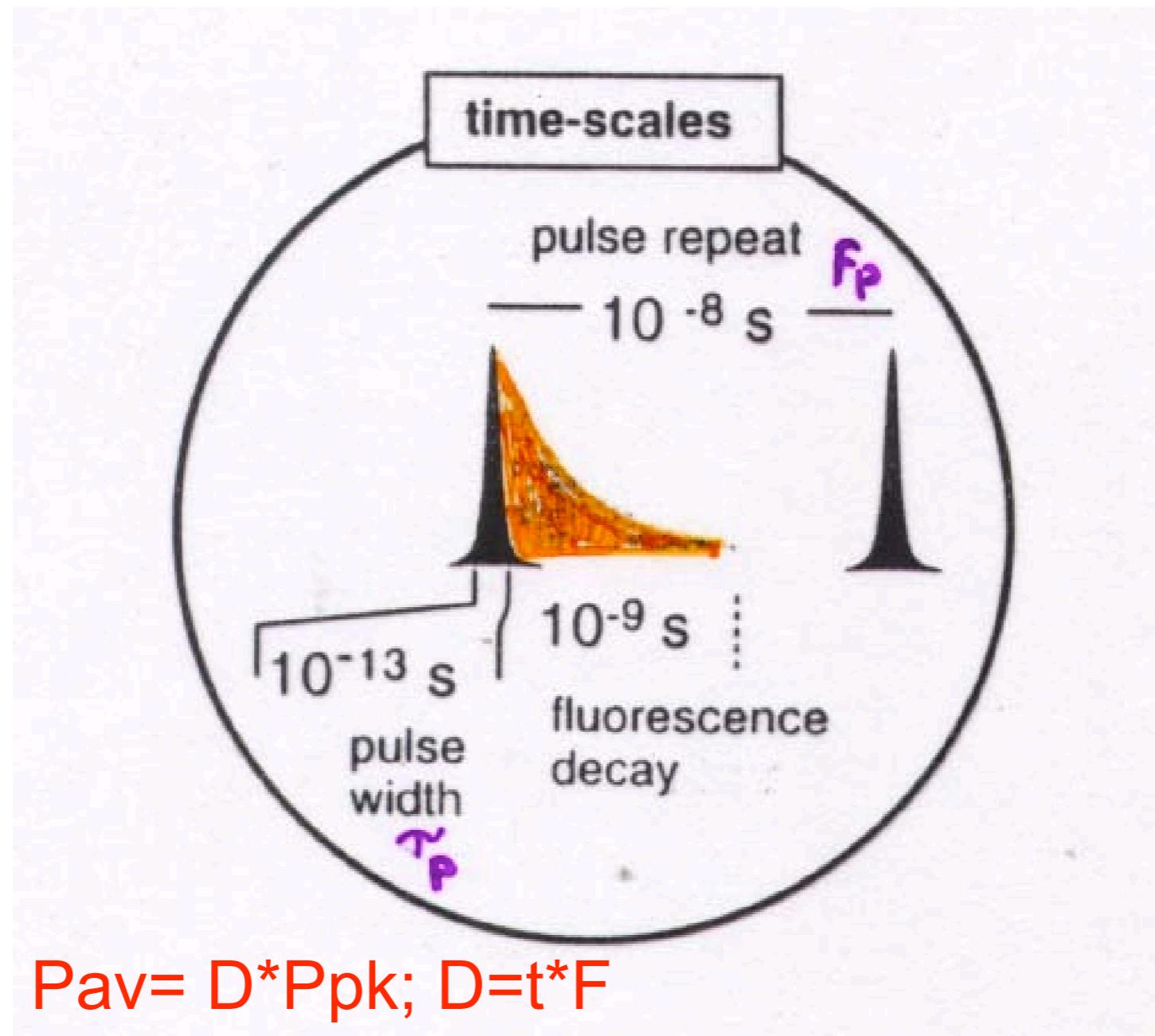
TWO-PHOTON EXCITATION MICROSCOPY



A.DIASPRO, G.CHIRICO, M.COLLI (2004) QUART. REV. BIOPHYS., VOL.38, NR.2, PP.1-72 (2006).

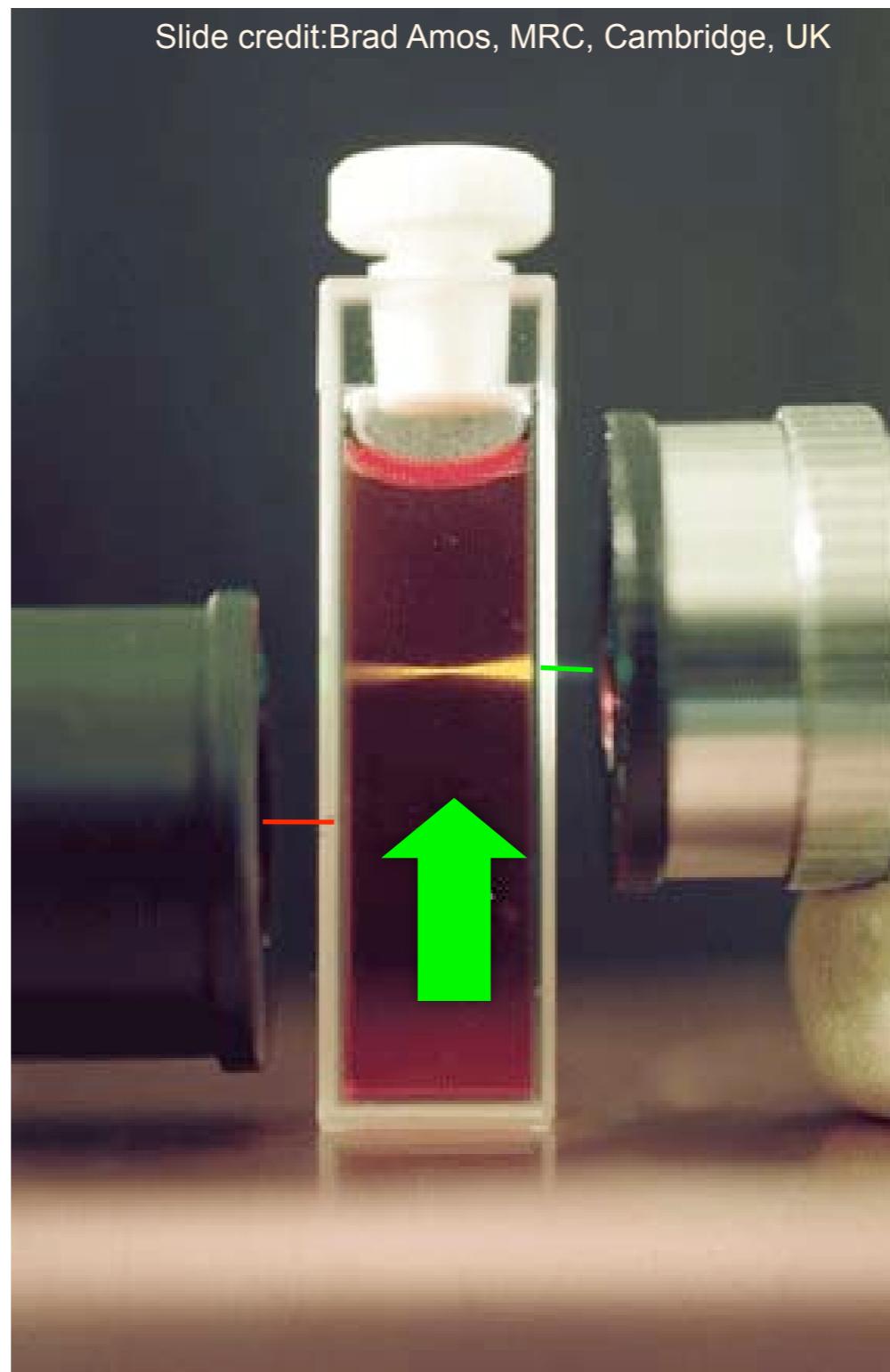
TWO-PHOTON EXCITATION MICROSCOPY

CORE ELEMENTS ARE LASER SOURCES

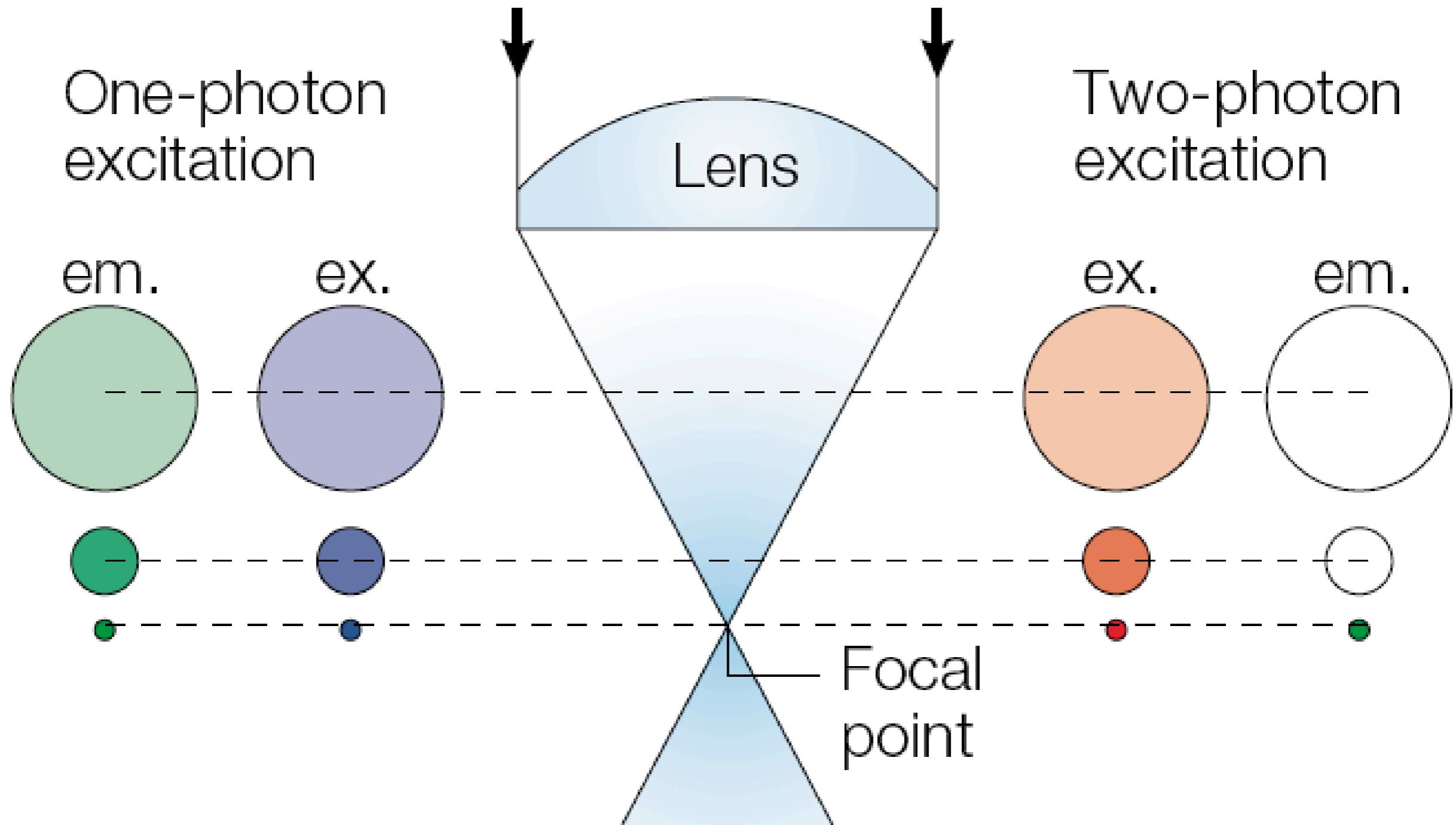


TWO-PHOTON EXCITATION MICROSCOPY

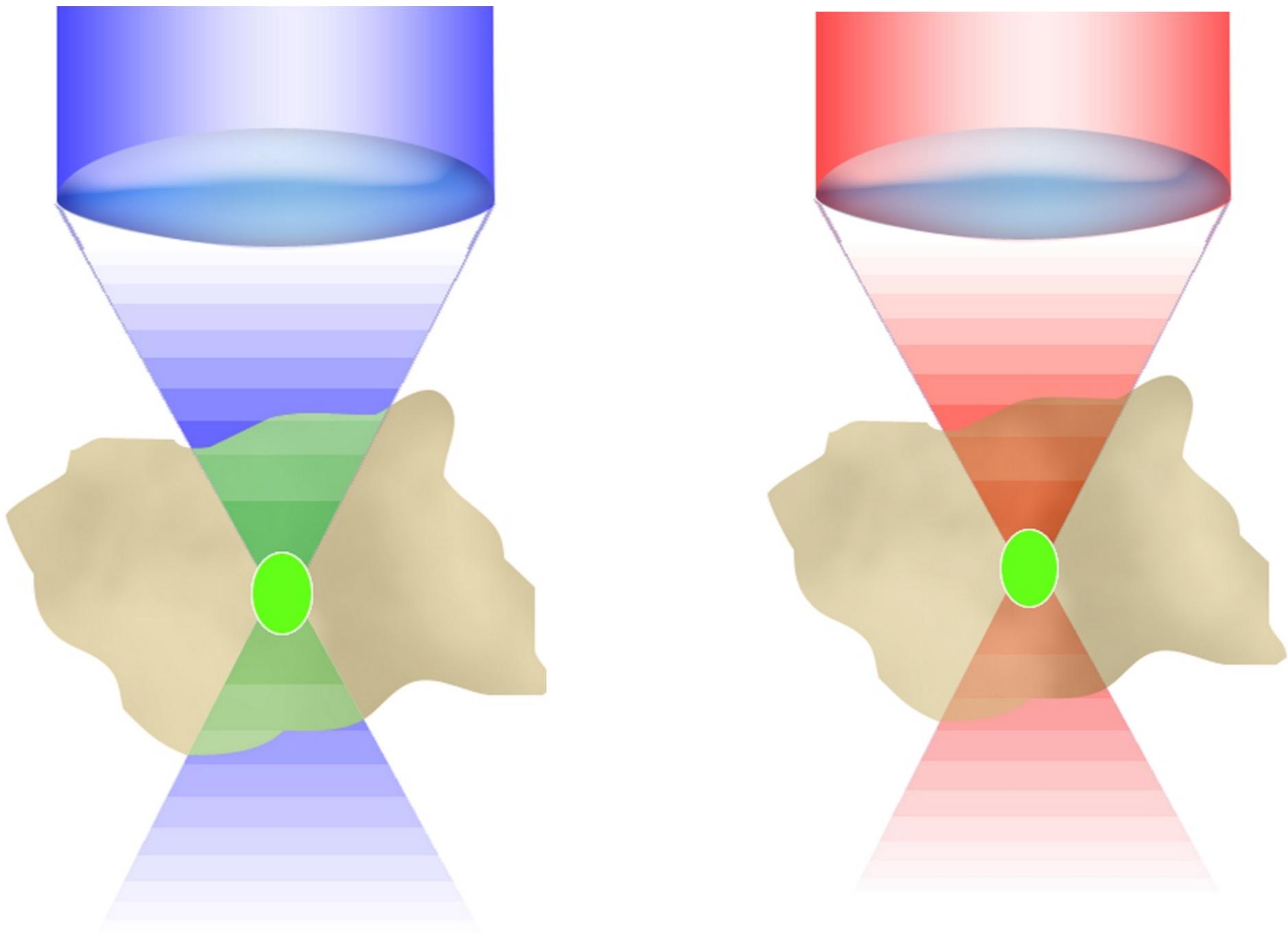
Slide credit: Brad Amos, MRC, Cambridge, UK



TWO-PHOTON EXCITATION MICROSCOPY

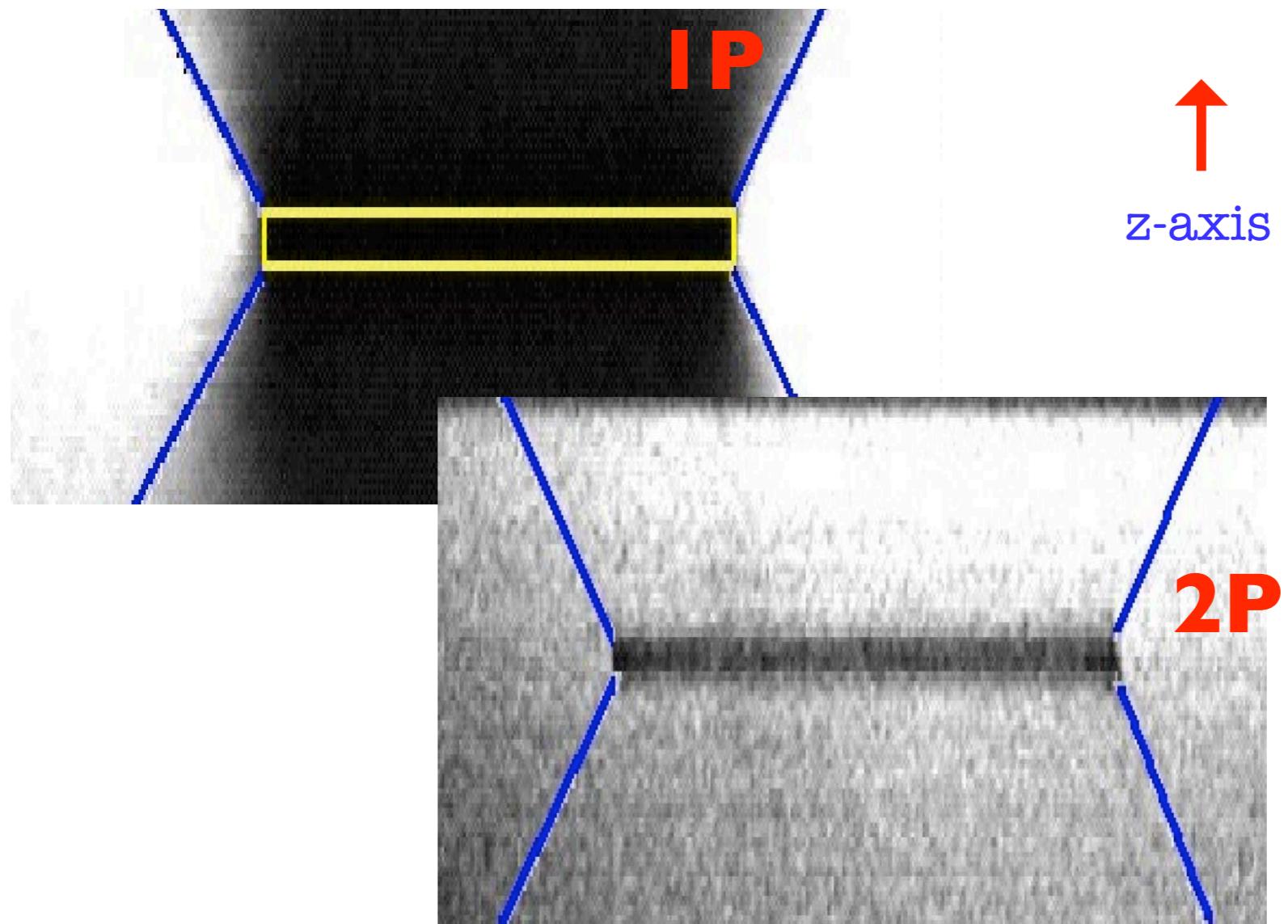


M.D.Cahalan et al. (2002) Nat.Rev.Immunol., 2: 872



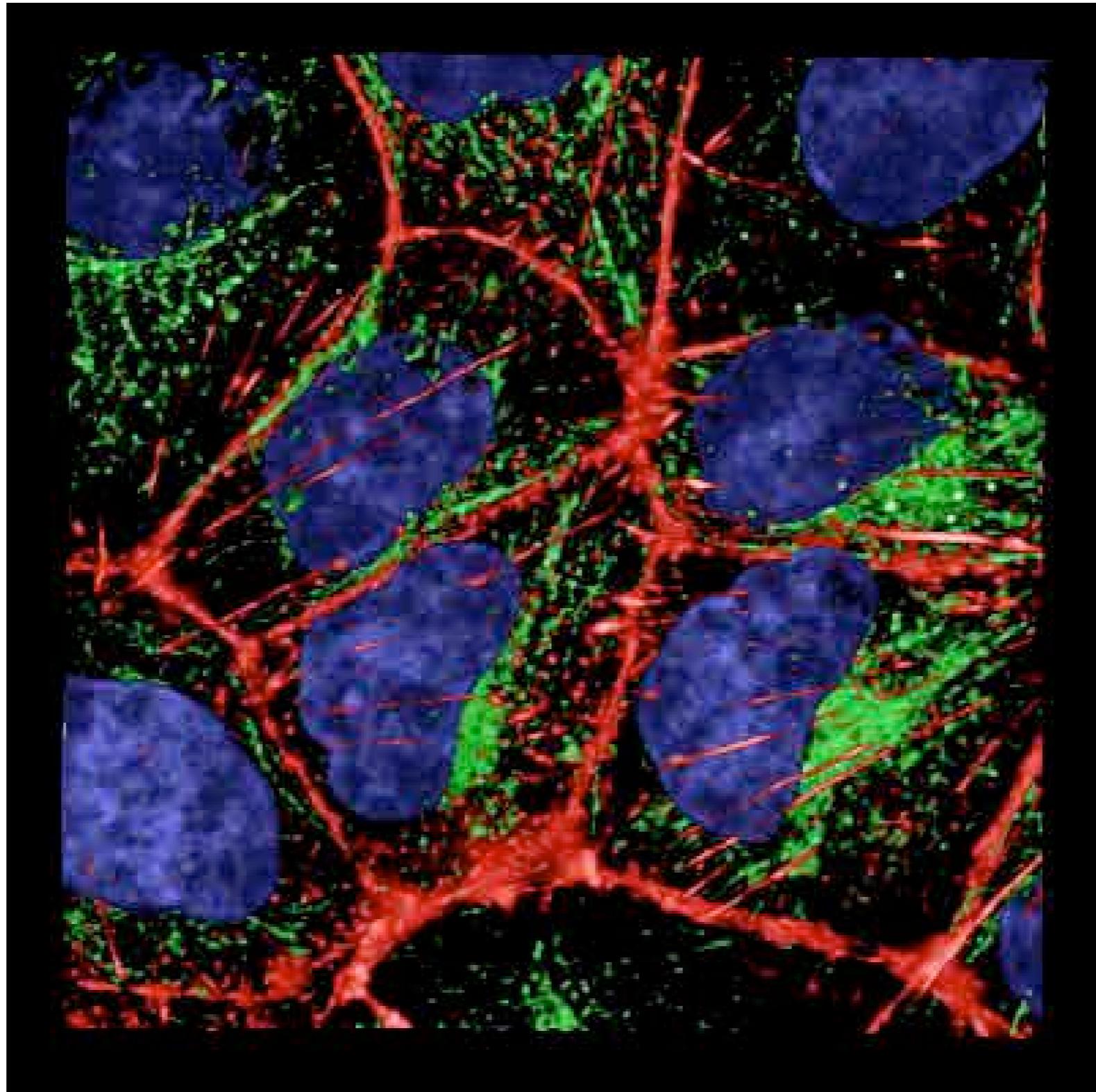
Slide credit: Paolo Bianchini, LAMBS-MicroScoBio, Genova

TWO-PHOTON EXCITATION MICROSCOPY



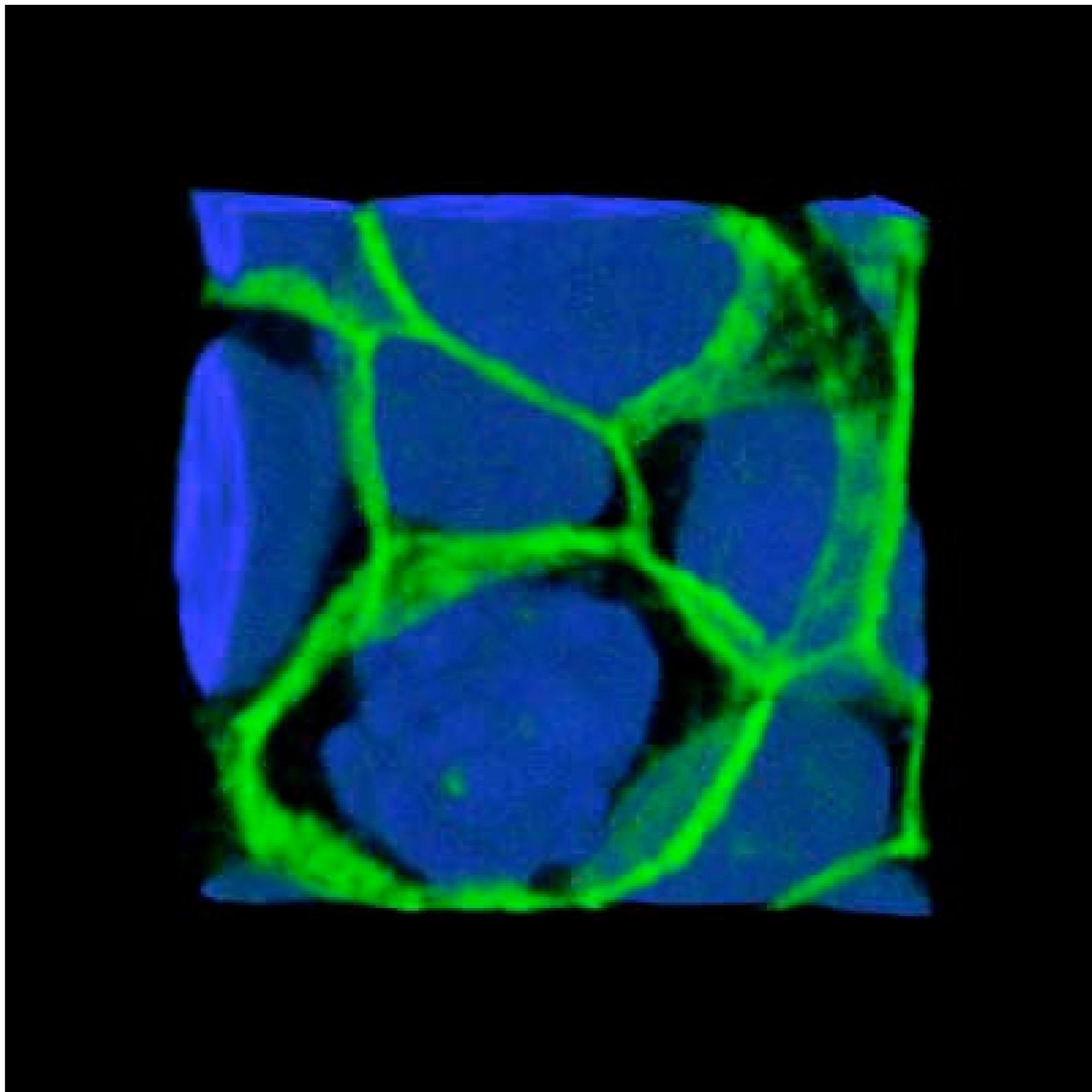
Slide credit: David Piston, Vanderbilt University

TWO-PHOTON EXCITATION MICROSCOPY



Tissue cultured PTR cells, TexasRed-Phalloidin, FITC- α tubulin, and DAPI. (Drs. Wang and Dunn, ICBM)

TWO-PHOTON EXCITATION MICROSCOPY

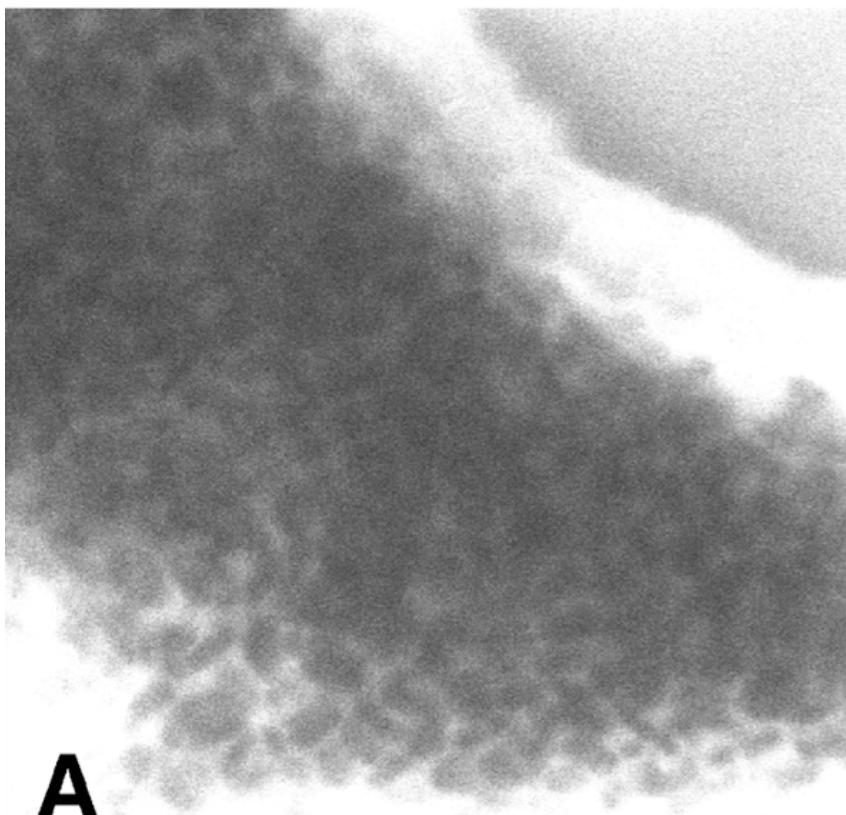


Two photon volume of dapi labeled nuclei and fluorescein labeled anti-cadherin. (Dr. Dunn, ICBM)

TWO-PHOTON EXCITATION MICROSCOPY

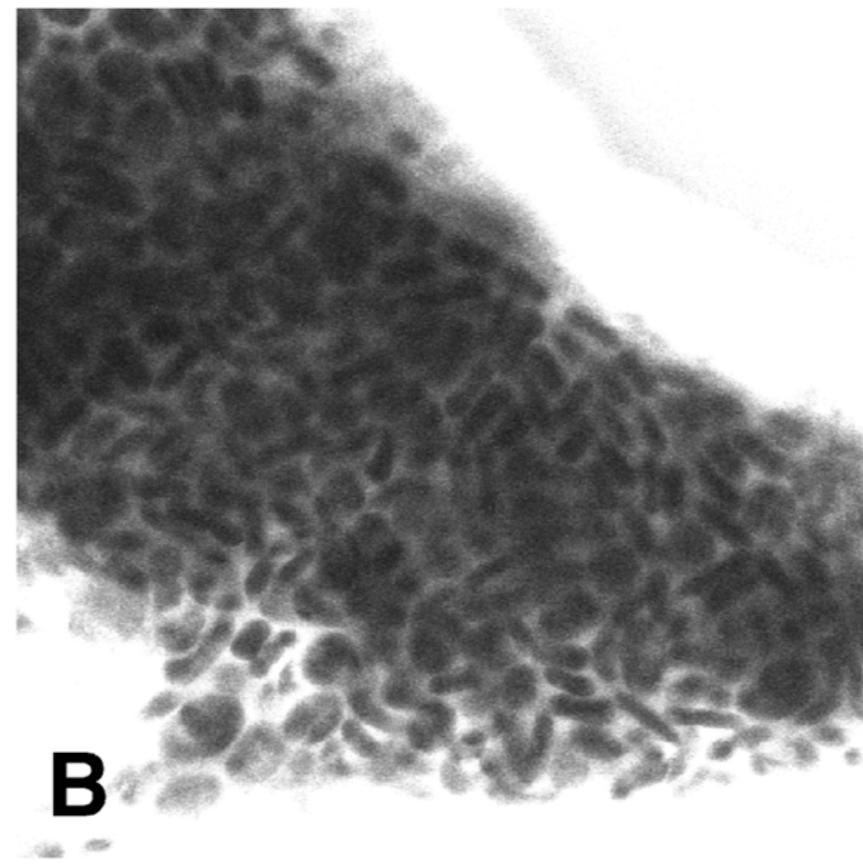
Penetration depth

One-Photon



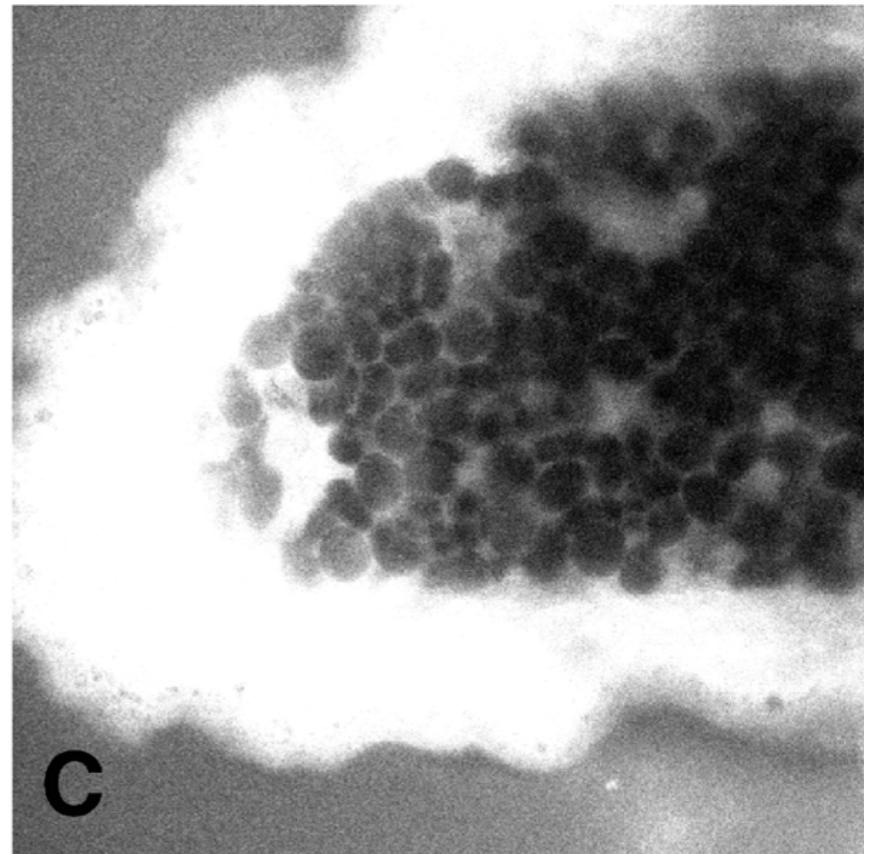
A

70 μm Deep



B

Two-Photon



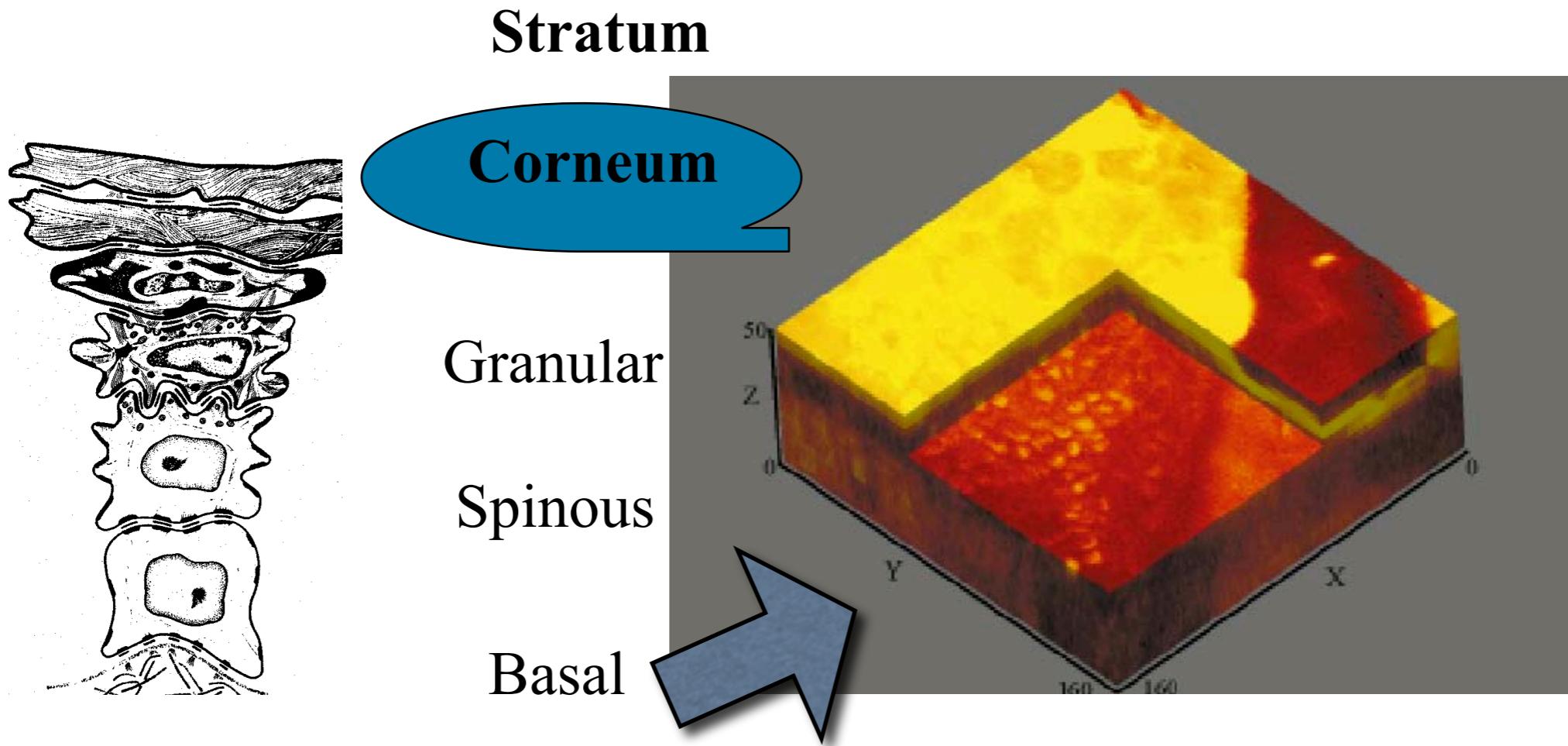
C

140 μm Deep

Images of a Shark Choroid Plexus Stained with Fluorescein

Piston DW (2005), PLoS Biol 3(6): e207

Penetration depth

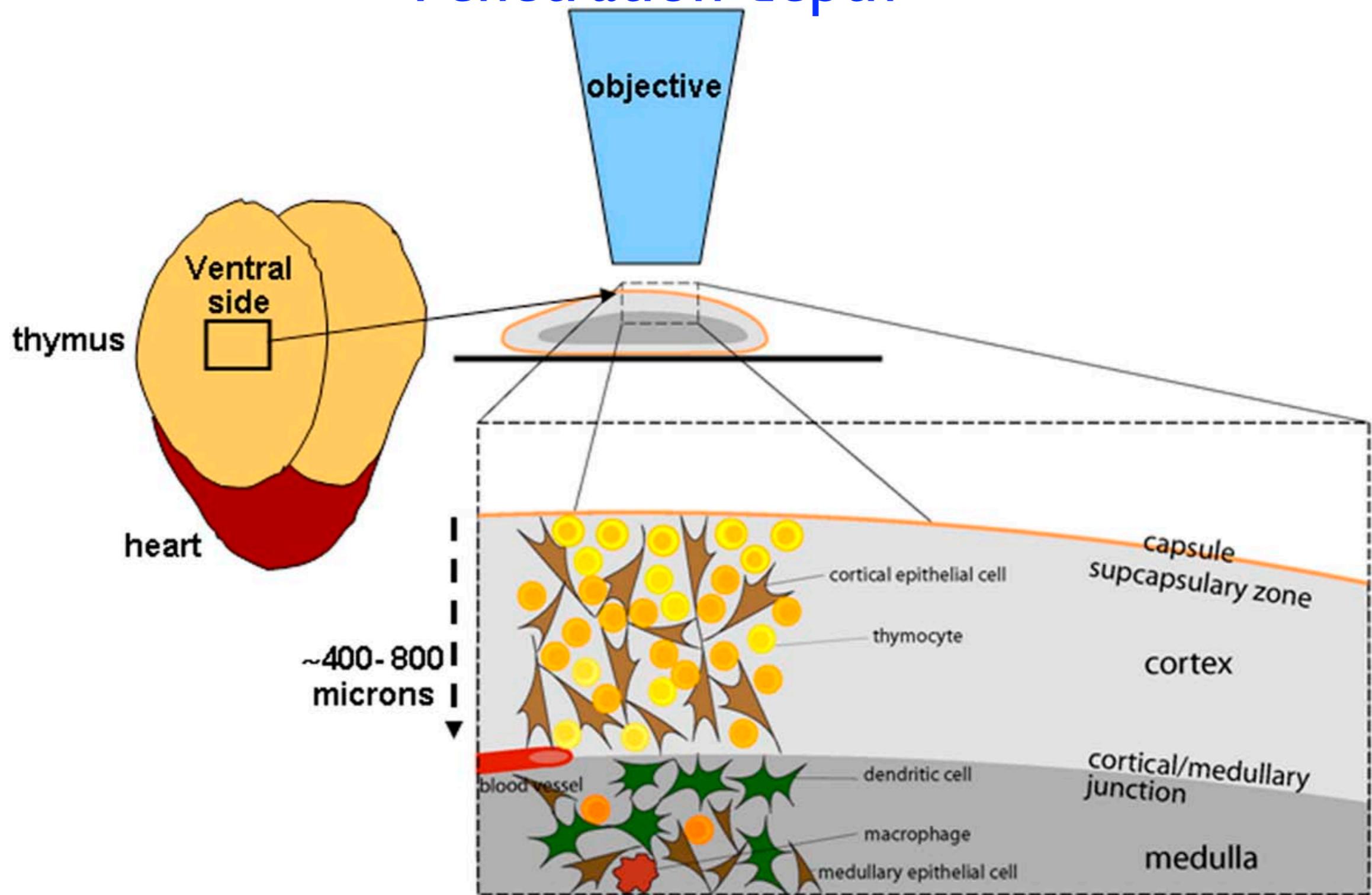


Three-dimensional reconstructed two-photon images of *in vivo* human skin. The strata corneum and the basal layers are clearly visible.

(Image credit: PETER SO, BARRY MASTERS)

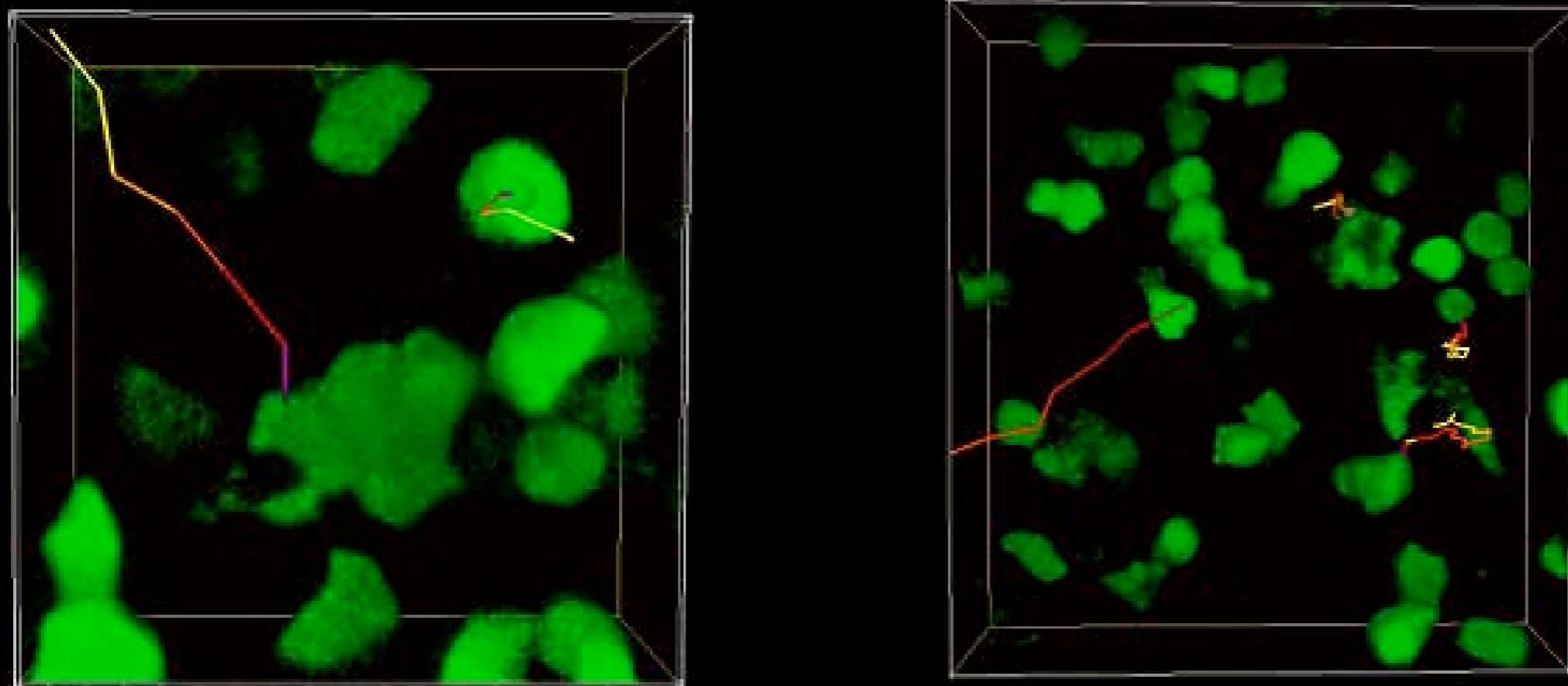
TWO-PHOTON EXCITATION MICROSCOPY

Penetration depth



Real-time visualization of thymocytes within intact thymic lobes using two-photon microscopy (2005) PLoS Biol 3(6): e205

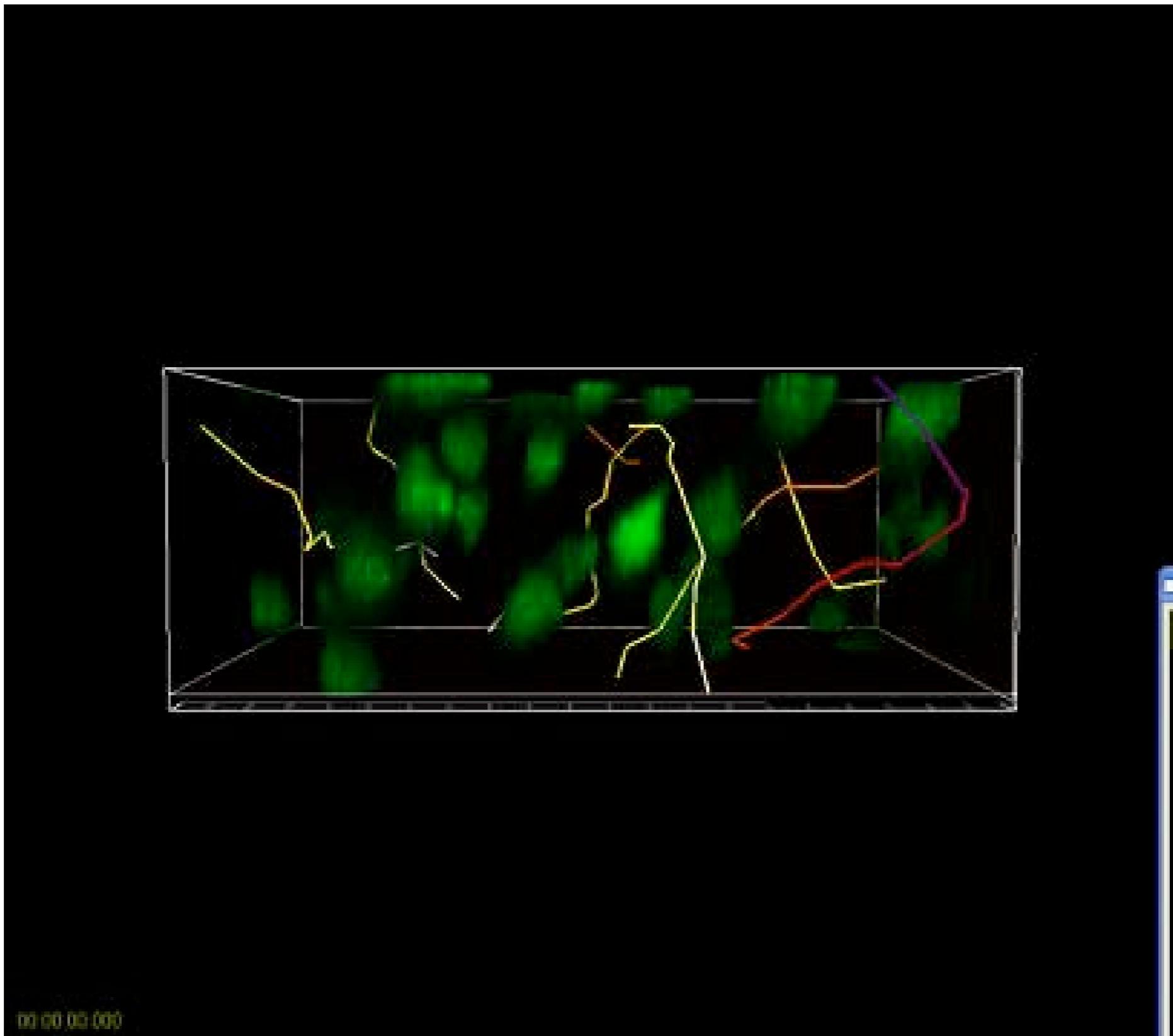
TWO-PHOTON EXCITATION MICROSCOPY



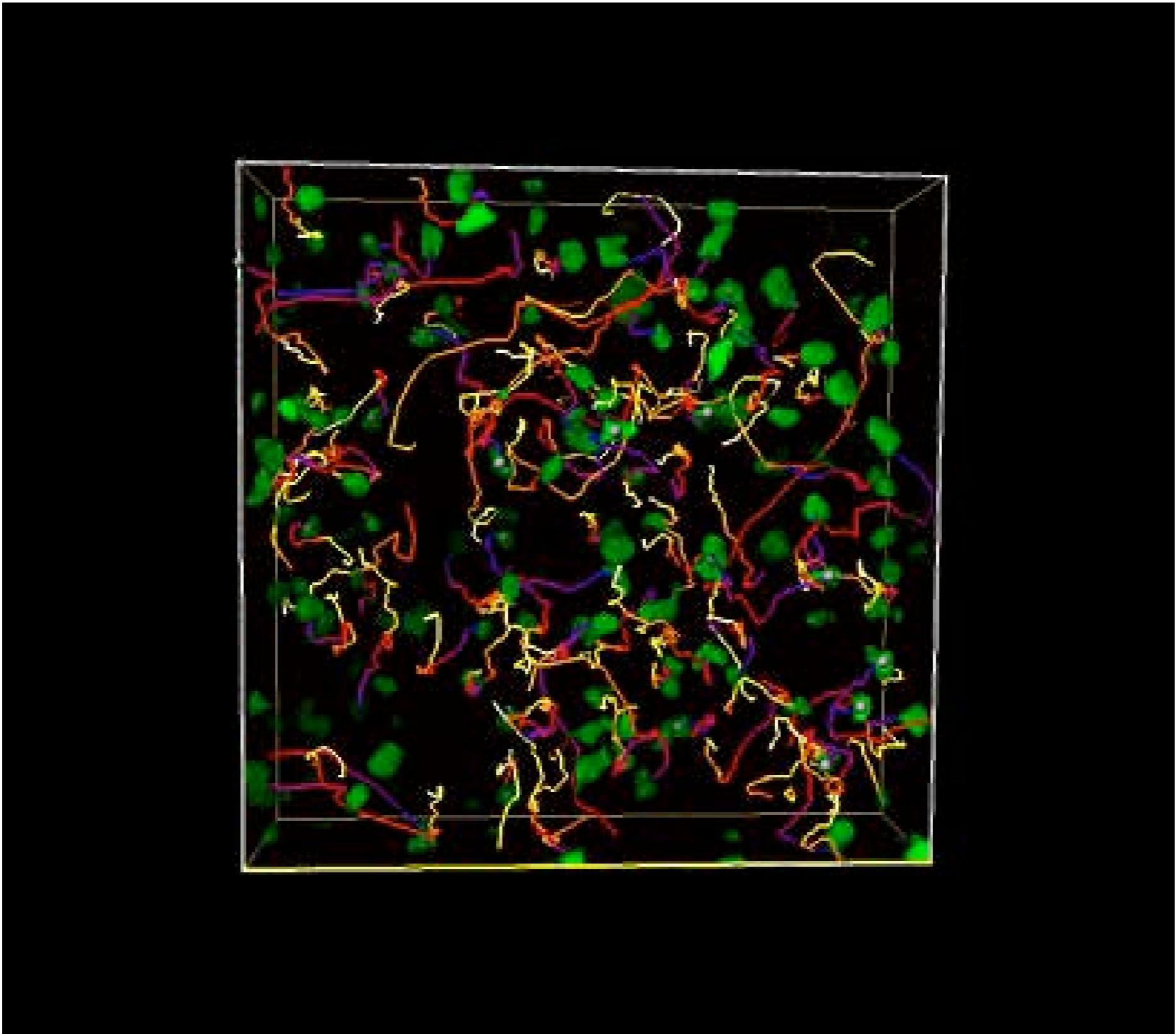
Two-photon microscopy reveals the three-dimensional dynamics of B cell (red) and T cell (green) conjugate within lymph tissue in real time

Citation: (2005) Tracking the Details of an Immune Cell Rendezvous in 3-D. PLoS Biol 3(6): e206

TWO-PHOTON EXCITATION MICROSCOPY

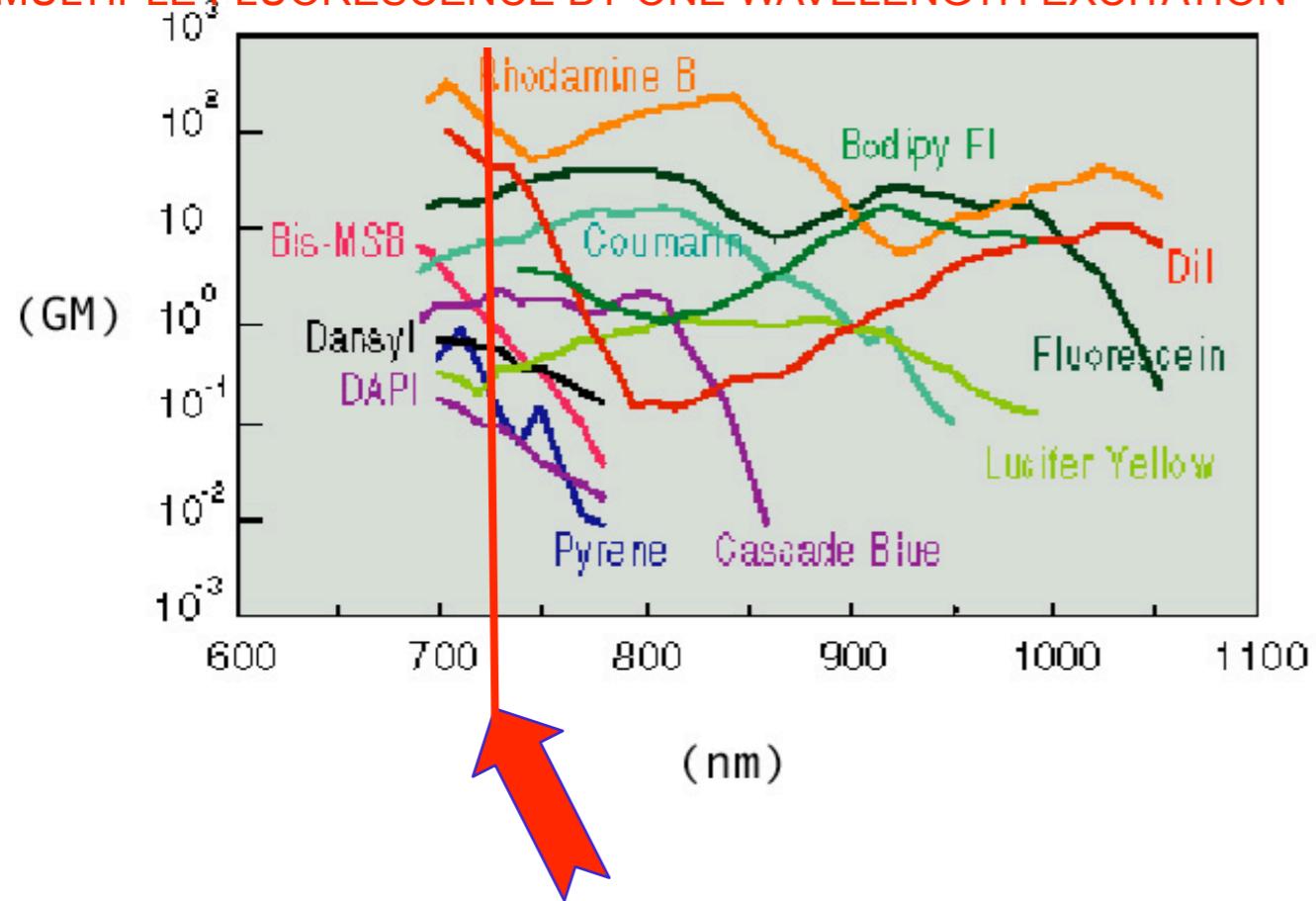


TWO-PHOTON EXCITATION MICROSCOPY

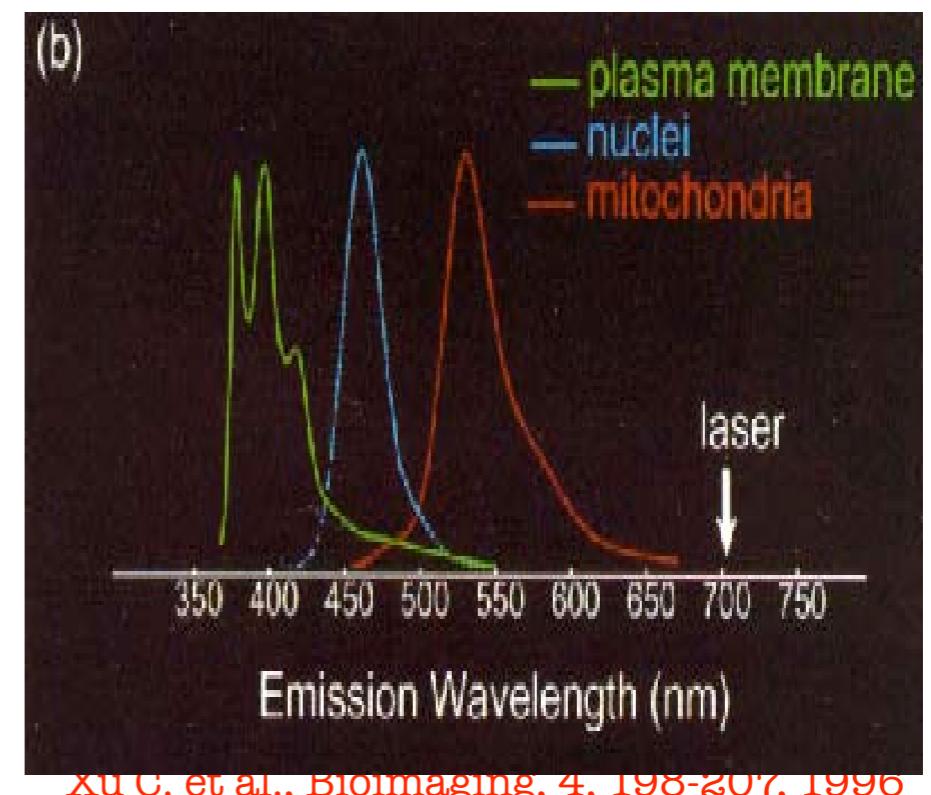
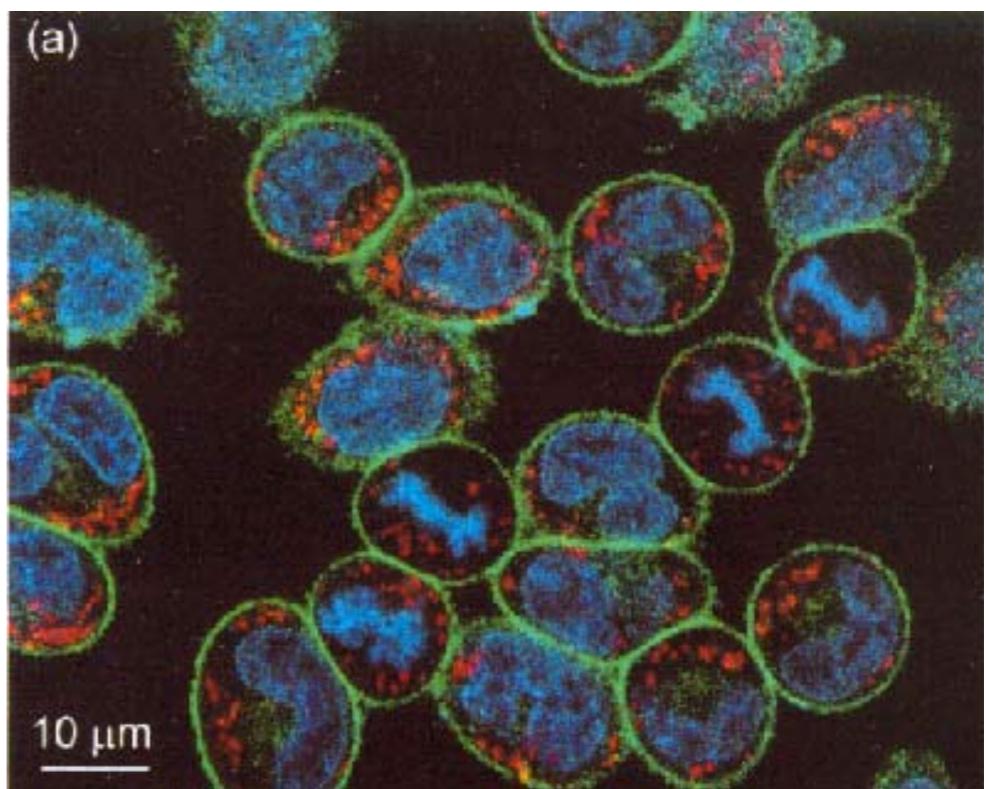
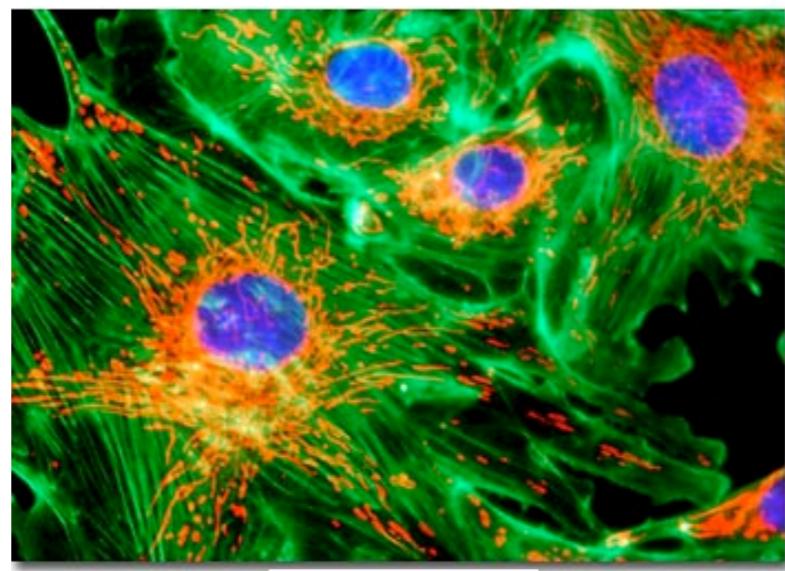
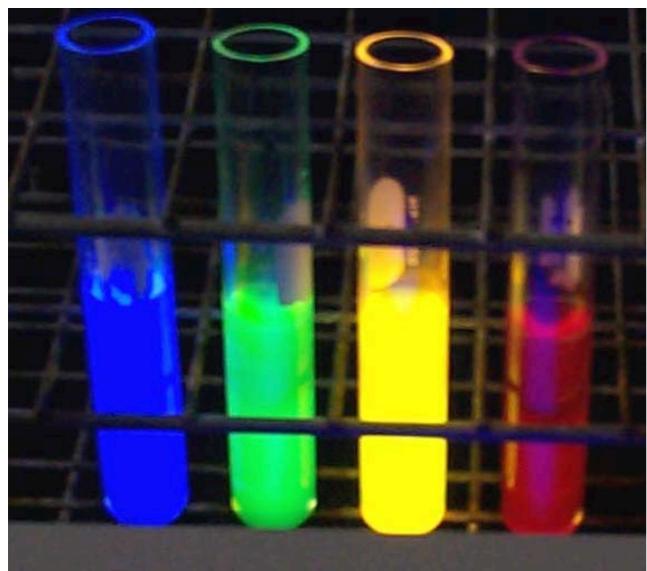


TWO-PHOTON EXCITATION MICROSCOPY

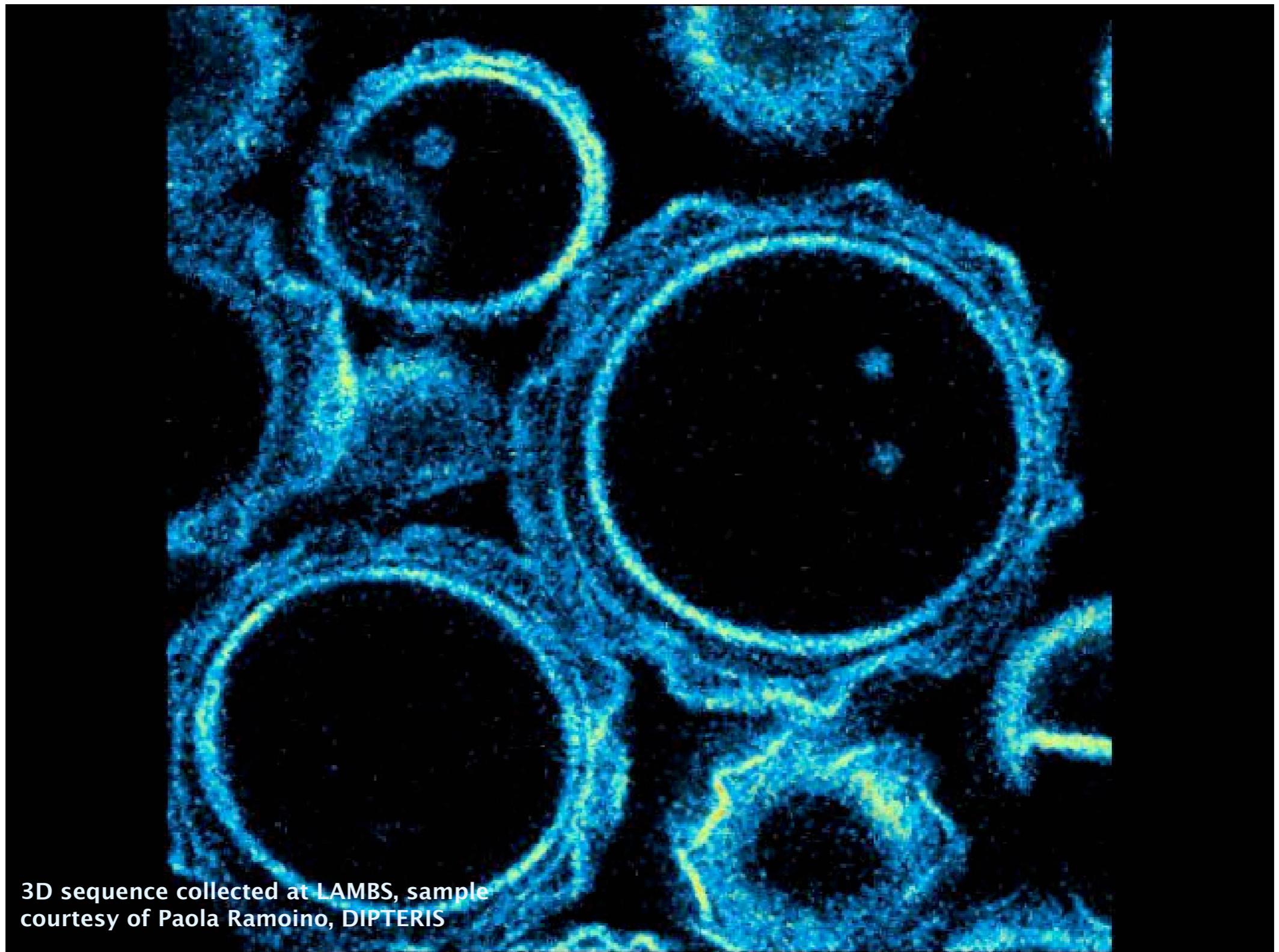
MULTIPLE FLUORESCENCE BY ONE WAVELENGTH EXCITATION



One dice...two dices...to get 5!

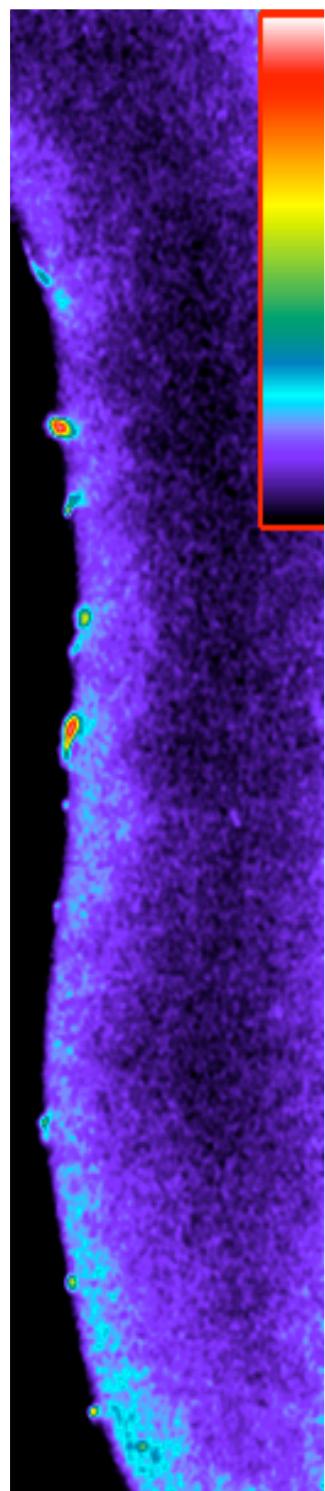


TWO-PHOTON EXCITATION MICROSCOPY



A.DIASPRO (2005) TRENDS IN BIOTECHNOLOGY, IN PRESS

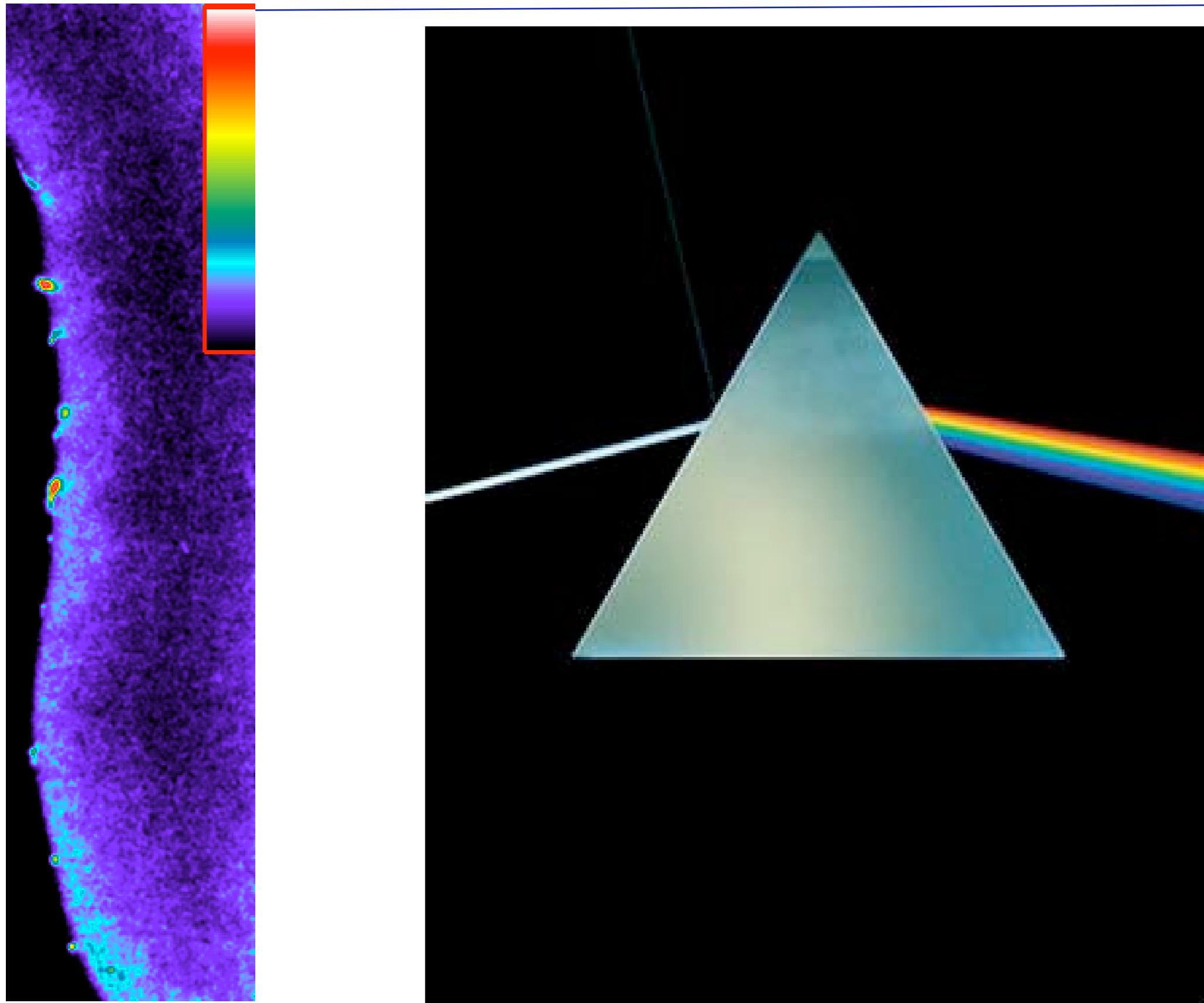
SPECTRAL INFORMATION



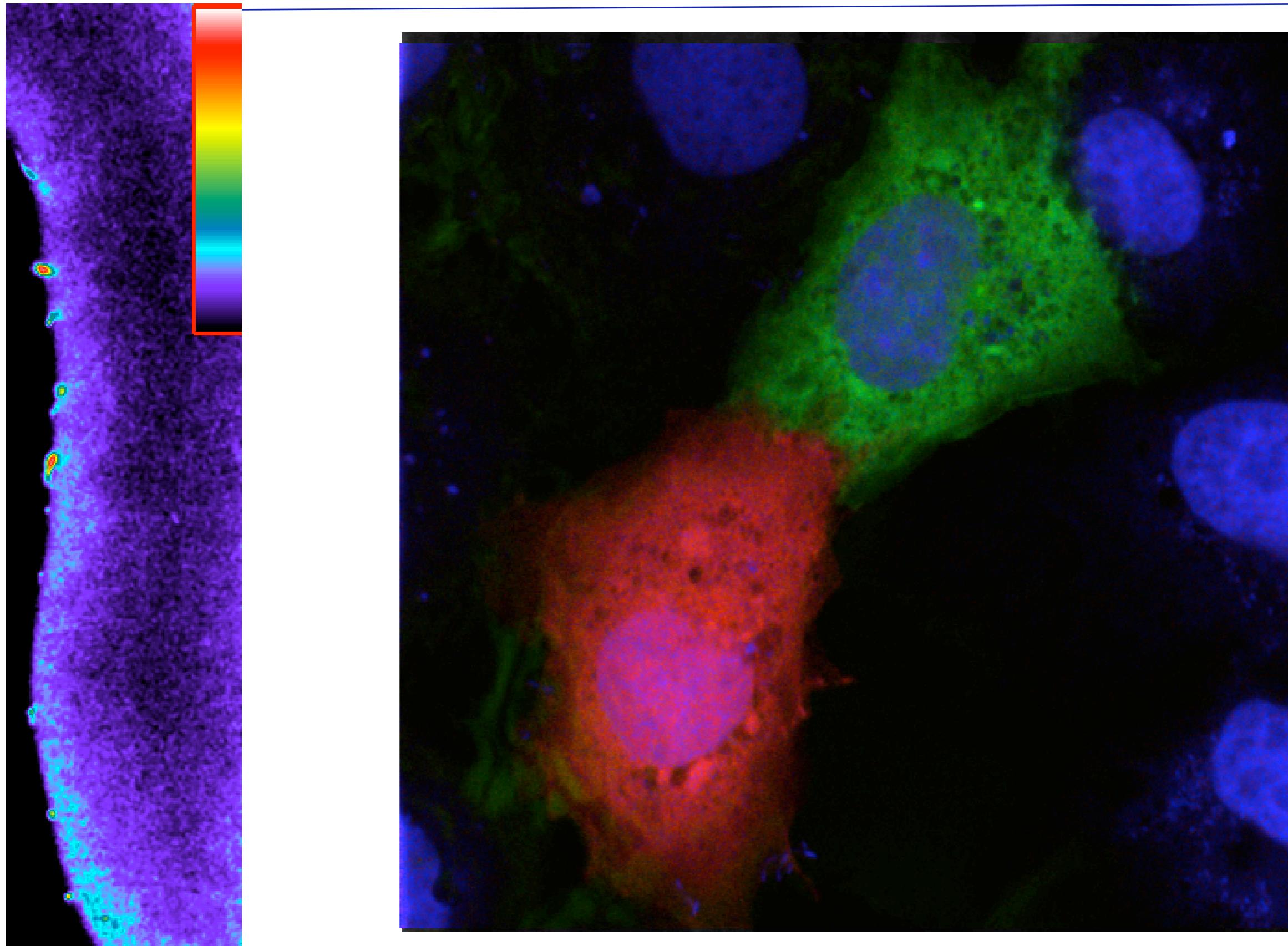
Two different stained neurons in Gaudi image

TCS SP5 resonant scanner 50 Hz framerate. Simultaneous record of CFP, GFP, YFP, DsRed, DiD and transmitted light.
Credits: LEICA Microsystems and Dr. Harald Hutter, Max Planck Institute (MPI) for Medical Research, Heidelberg, Germany

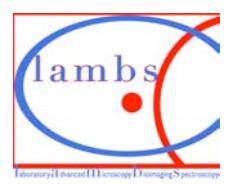
SPECTRAL SEPARATION PROBLEM



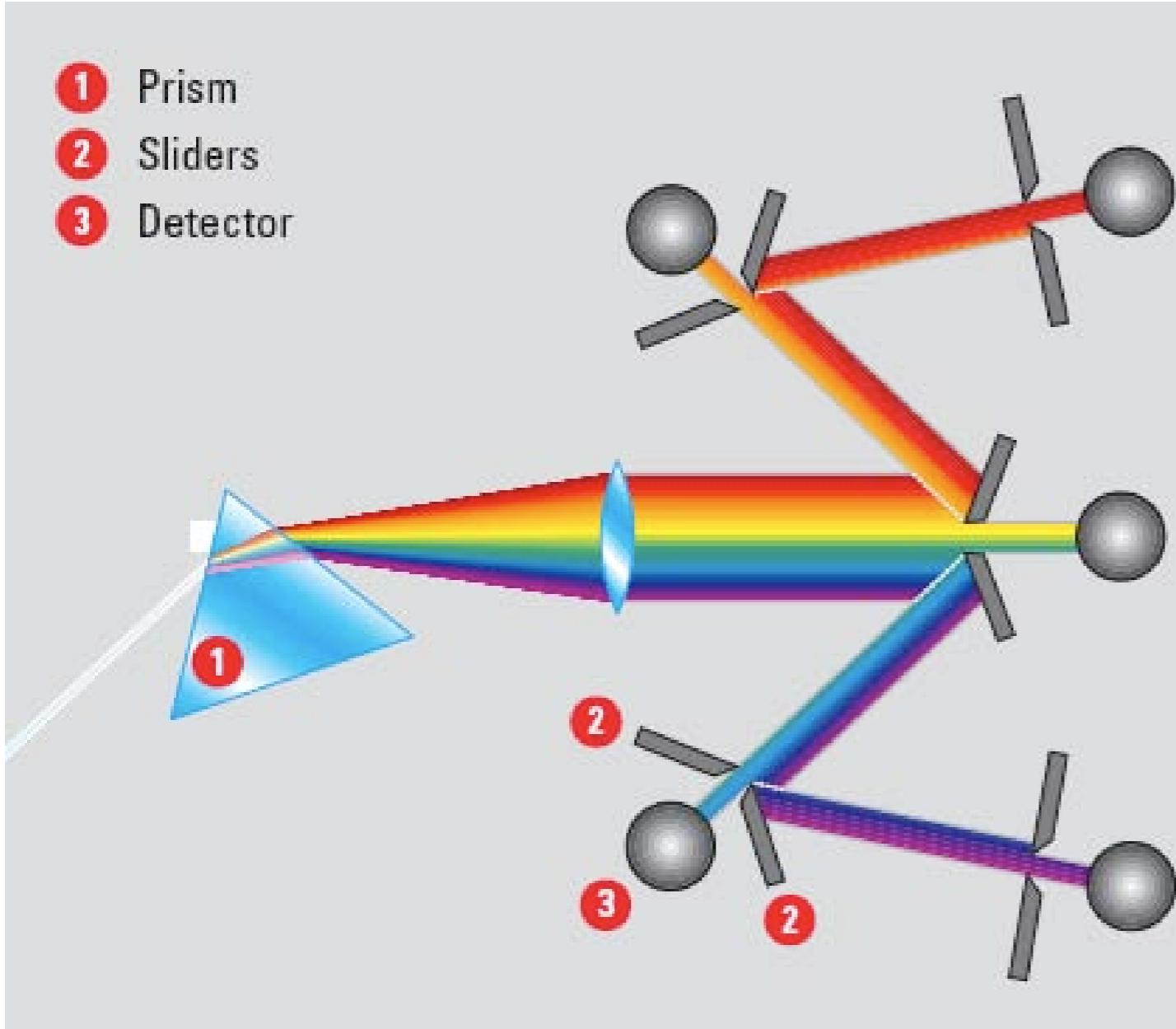
SPECTRAL SEPARATION PROBLEM



SPECTRAL SEPARATION PROBLEM



- 1 Prism
- 2 Sliders
- 3 Detector



TCS SP5 resonant scanner 71 Hz framerate.
Beating cilia in reflected light and Dil-Fluorescence together with transmitted light.

Courtesy of Kristin Tessmar und Detlev Arendt
European Molecular Biology Laboratory (EMBL)
Heidelberg, Germany

Leica
MICROSYSTEMS

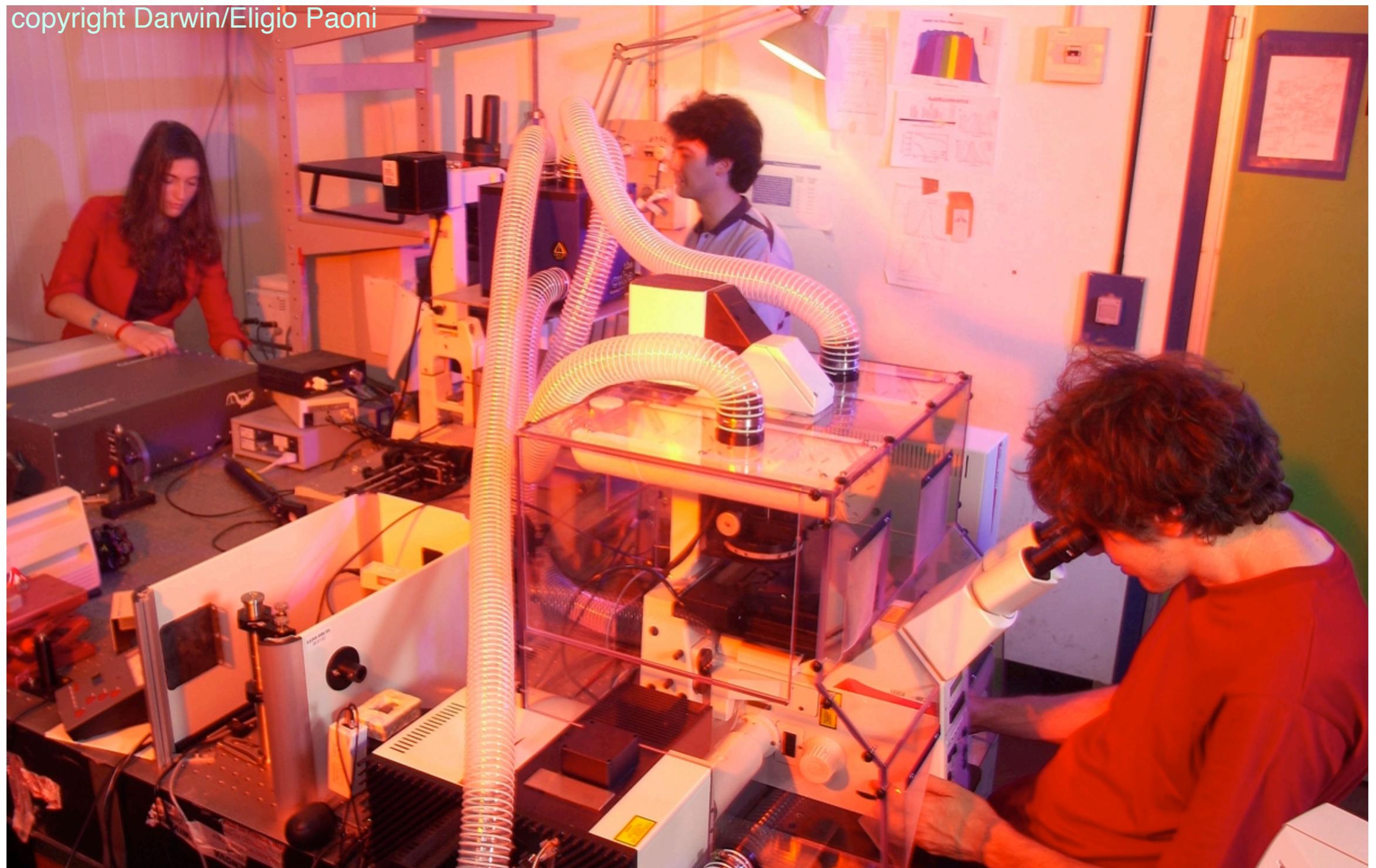
TWO-PHOTON EXCITATION MICROSCOPY

copyright Darwin/Elvio Paoni

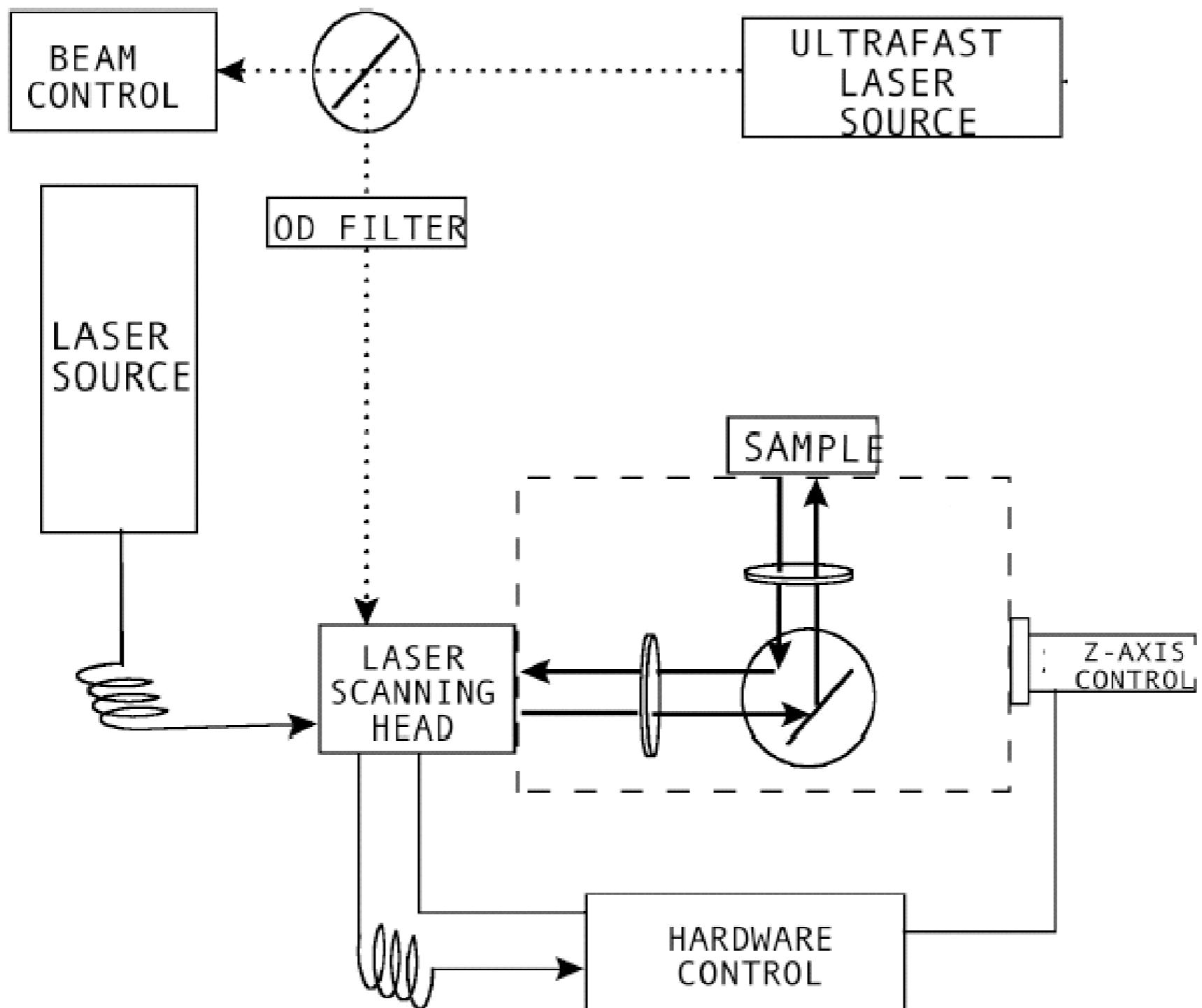


TWO-PHOTON EXCITATION MICROSCOPY

copyright Darwin/Elvio Paoni

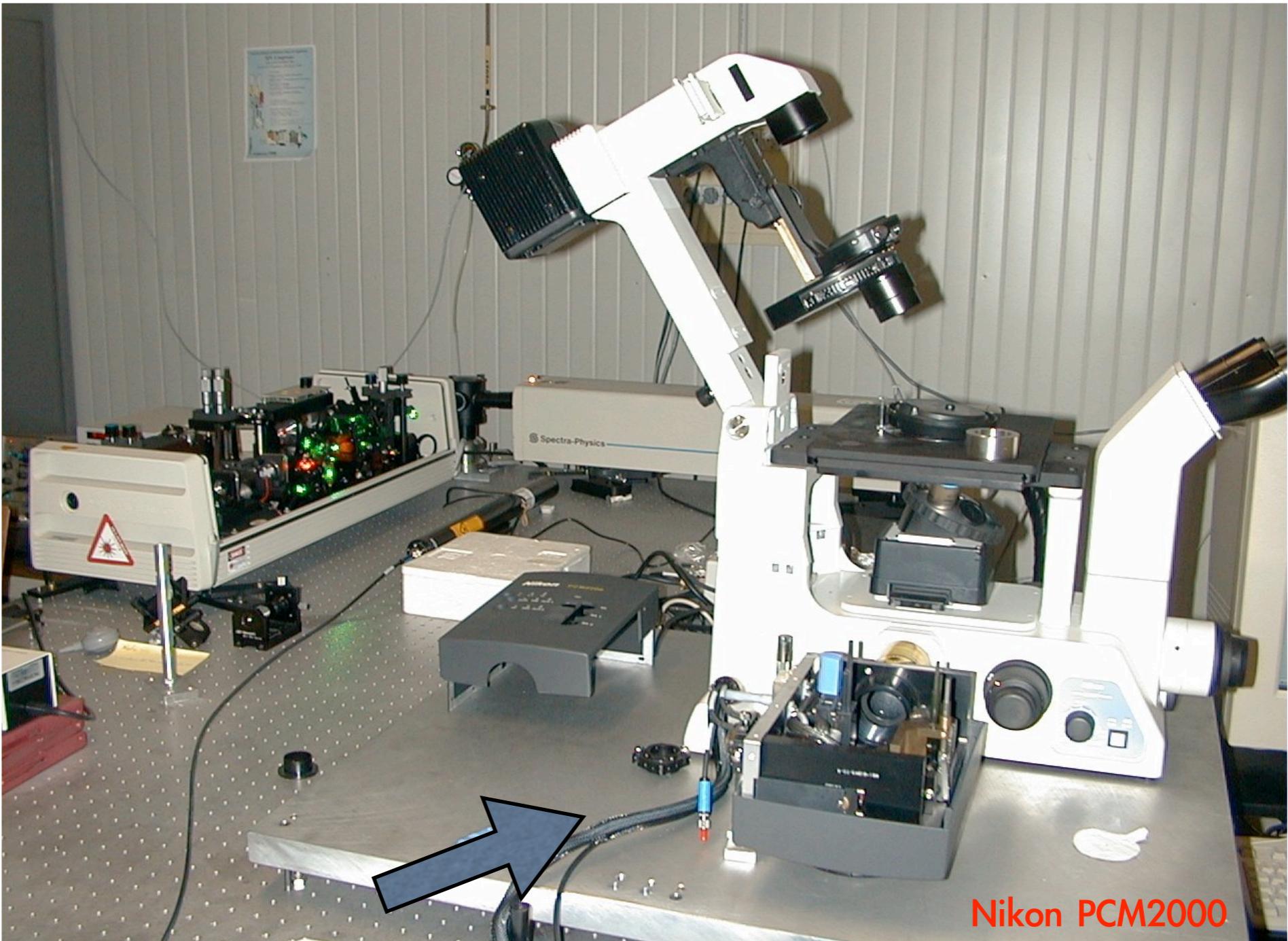


TWO-PHOTON EXCITATION MICROSCOPY



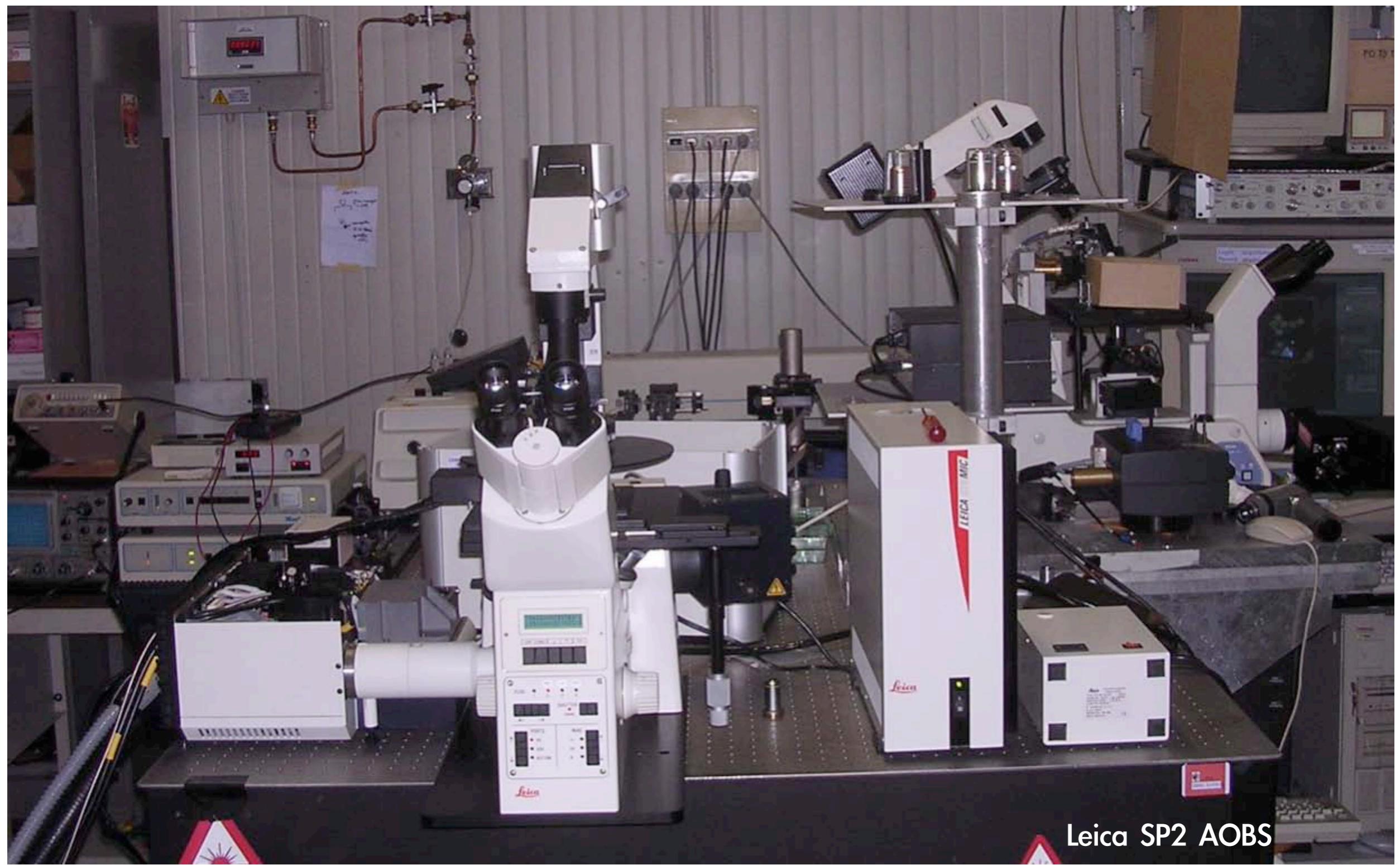
DIASPRO ET AL., MICROSC. RES. TECH., 47(3), 196-205, 1999

TWO-PHOTON EXCITATION MICROSCOPY



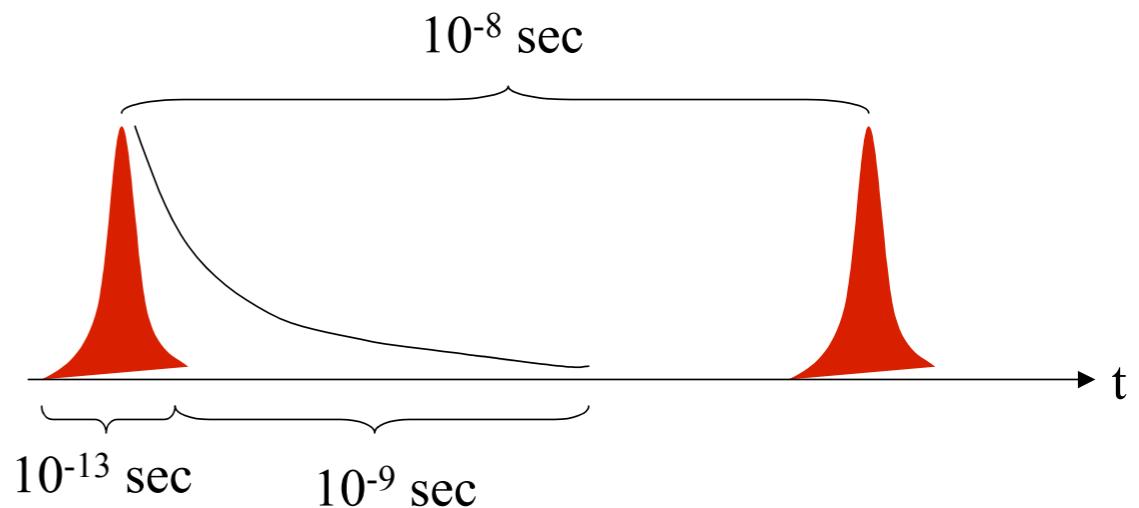
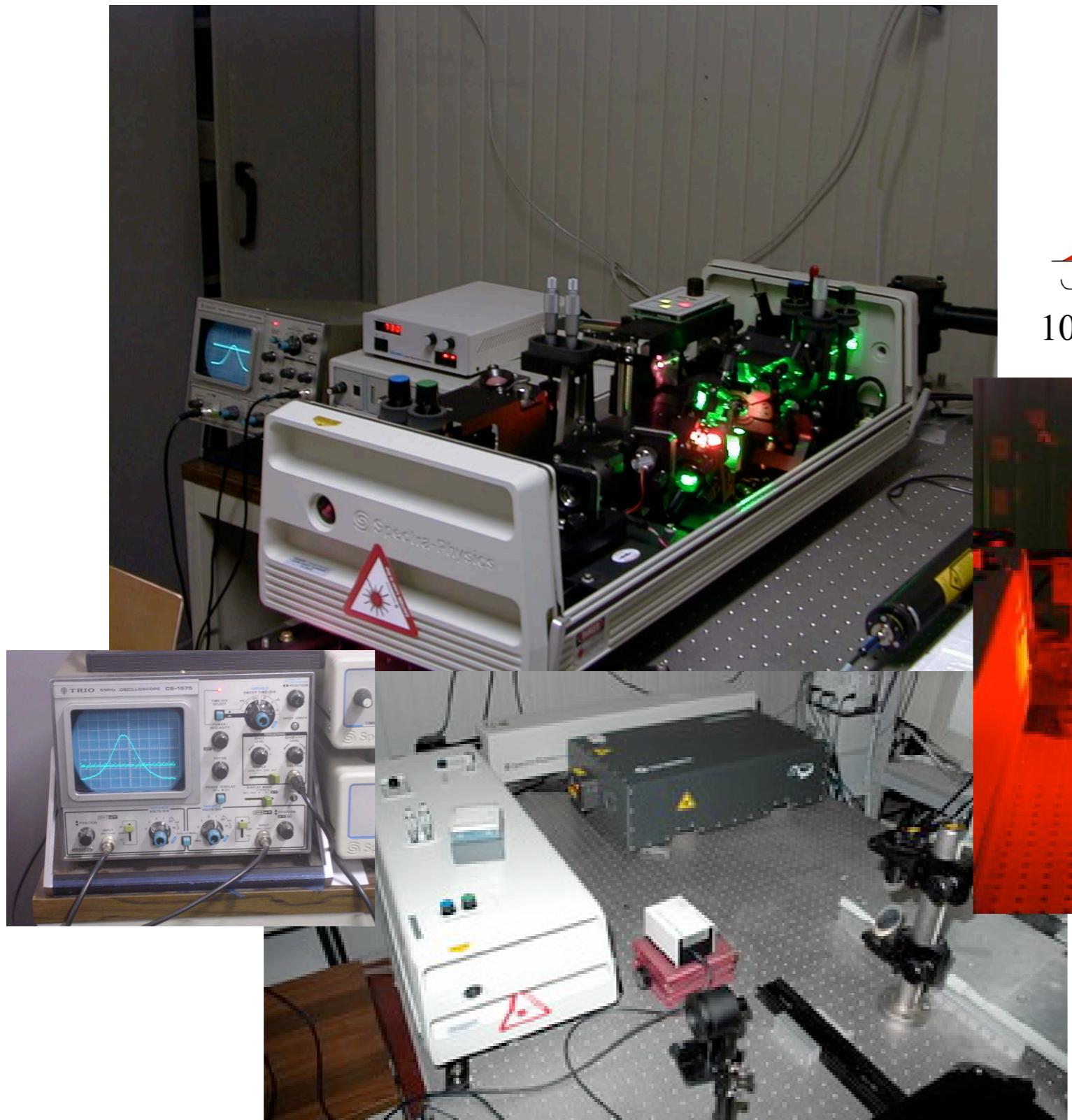
DIASPRO ET AL., MICROSC. RES. TECH., 47(3), 196-205, 1999

TWO-PHOTON EXCITATION MICROSCOPY



A.DIASPRO, G.CHIRICO, M.COLLI (2004) QUART. REV. BIOPHYS., VOL.38, NR.2, PP.1-72 (2006).

TWO-PHOTON EXCITATION MICROSCOPY



**Tsunami-Millennia
(680-830/780-930),
Chameleon XR (720-980)**

TWO-PHOTON EXCITATION MICROSCOPY

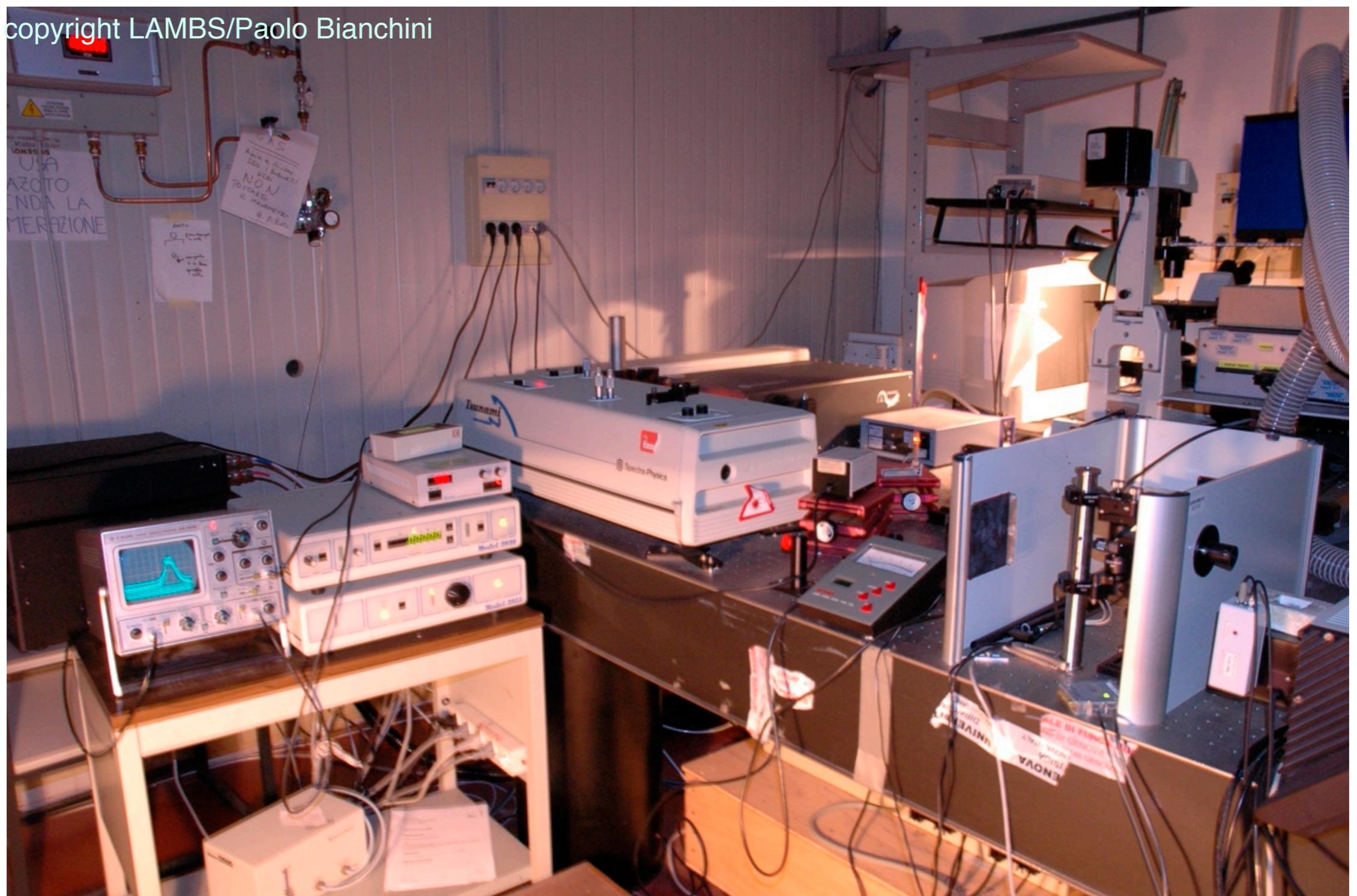
copyright LAMBS/Paolo Bianchini



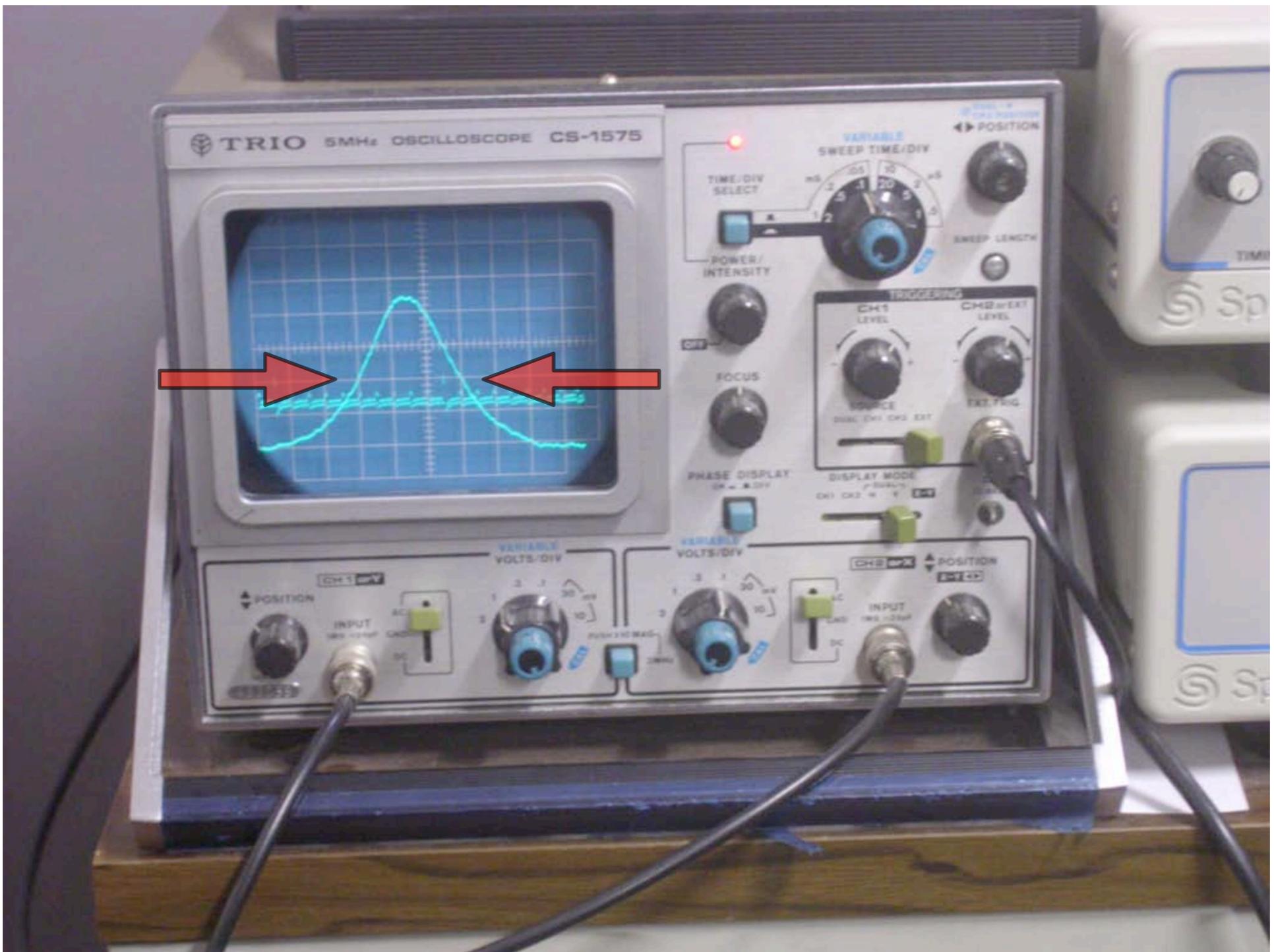
A.DIASPRO, G.CHIRICO, M.COLLIINI (2004) QUART. REV. BIOPHYS., VOL.38, NR.2, PP.1-72 (2006).

TWO-PHOTON EXCITATION MICROSCOPY

copyright LAMBS/Paolo Bianchini



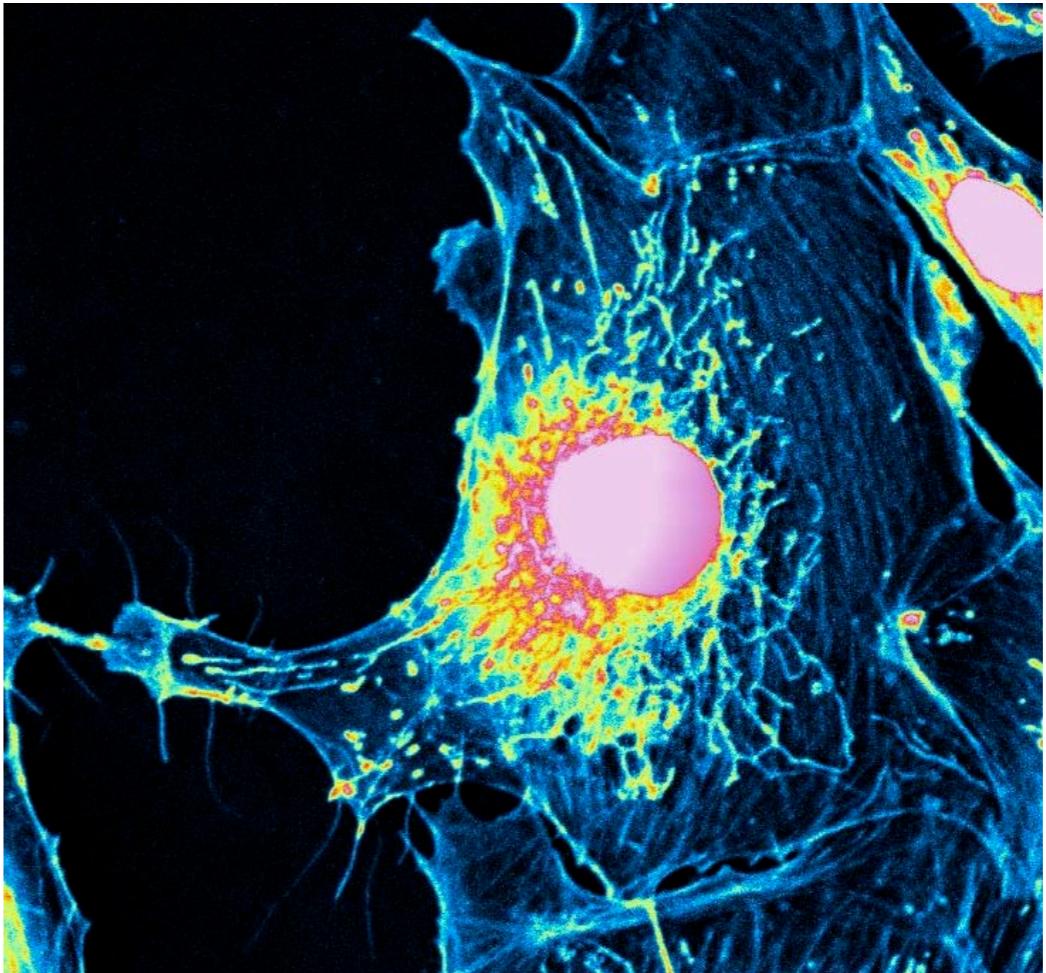
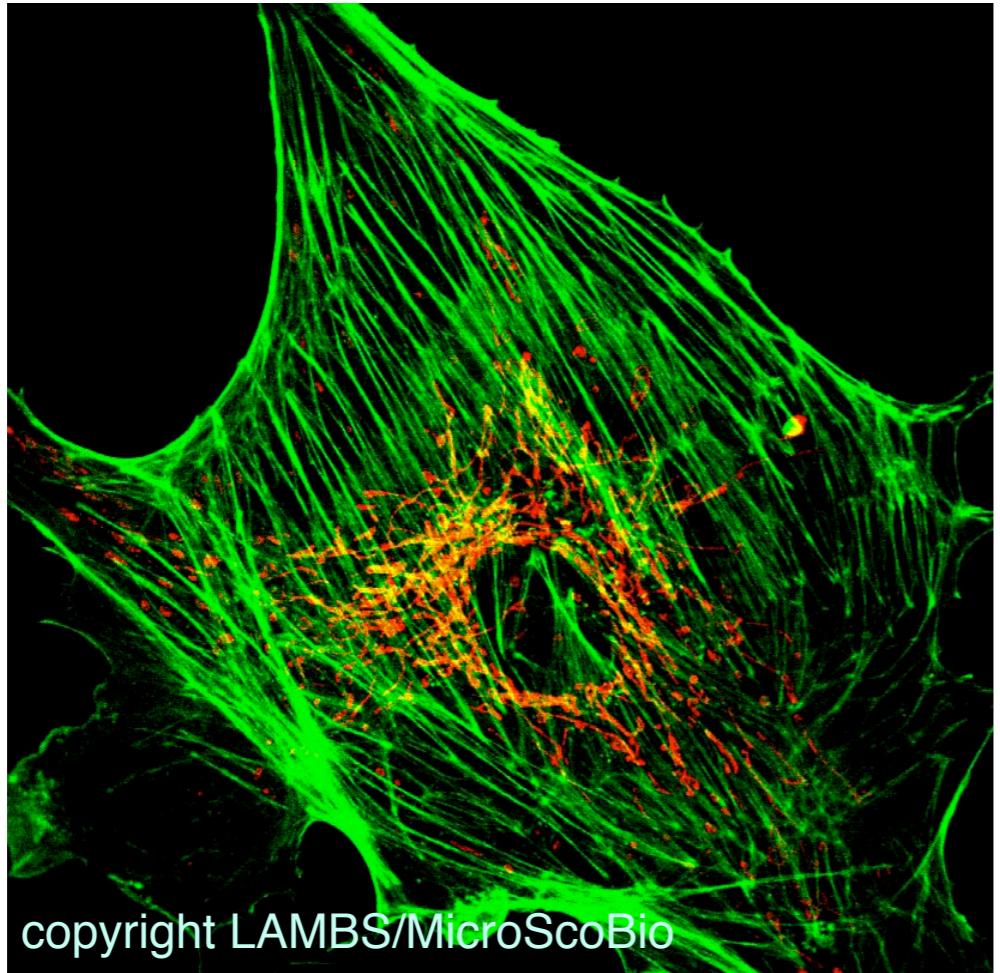
TWO-PHOTON EXCITATION MICROSCOPY



Slide credit: Mario Arace, 1978

TWO-PHOTON EXCITATION MICROSCOPY

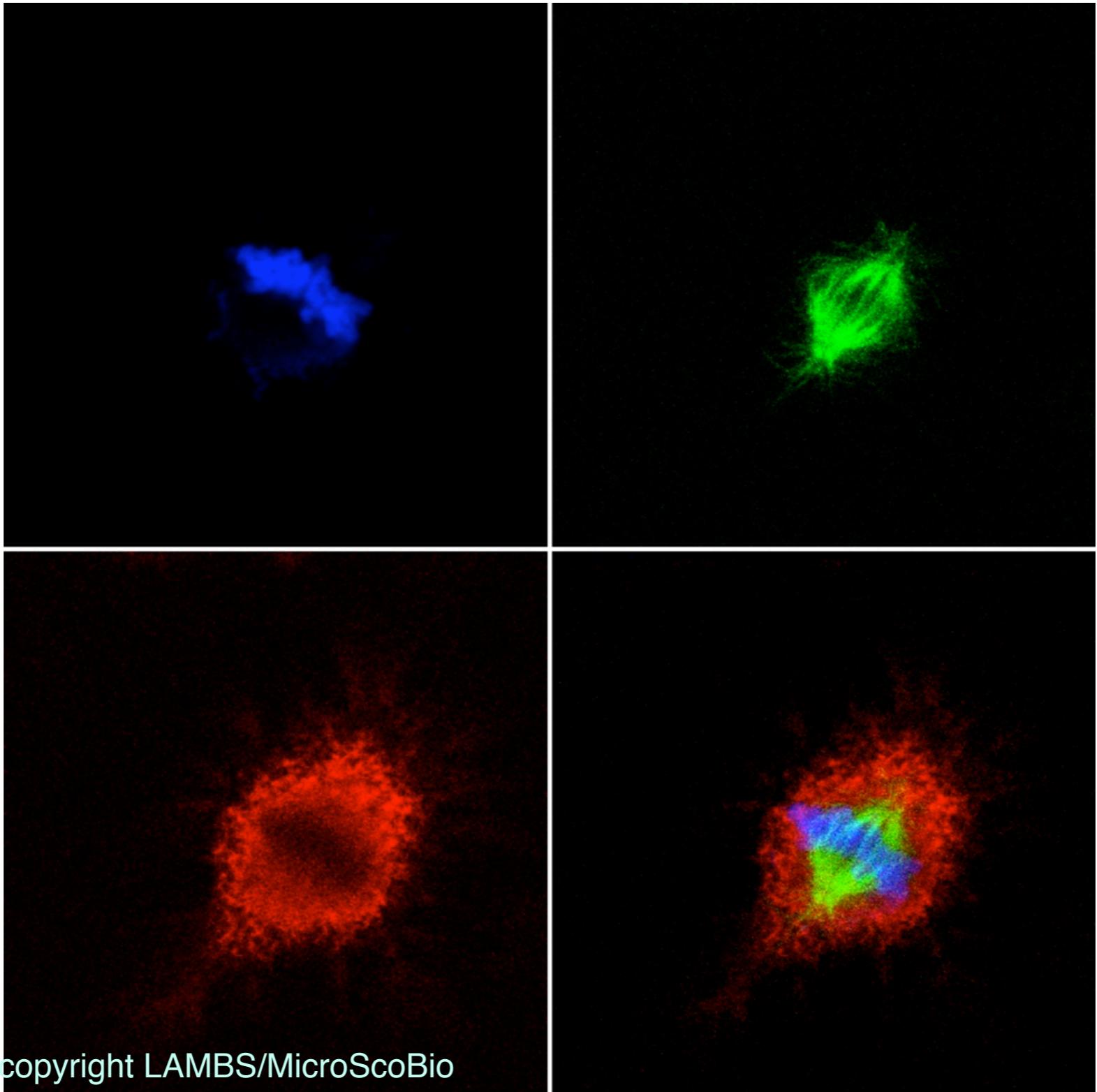
Confocal (left) and TPE (right) multiple fluorescence.



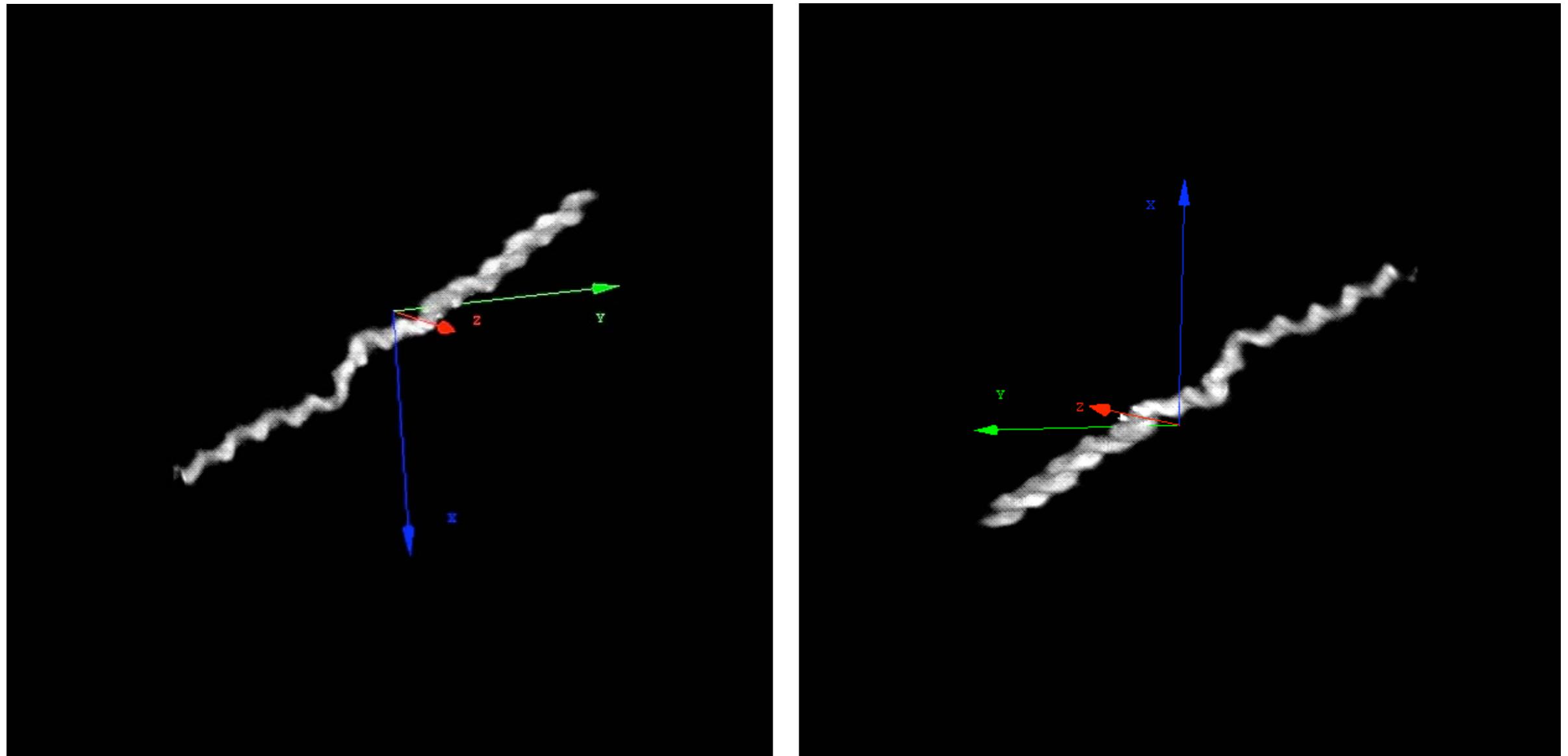
Cells are labelled with DAPI (DNA), mitotracker red (mitochondria) and bodipy (actin); F-14780 Molecular Probes slide.

Voce “Cellula”, Supplemento Piccola Treccani

TWO-PHOTON EXCITATION MICROSCOPY



TWO-PHOTON EXCITATION MICROSCOPY

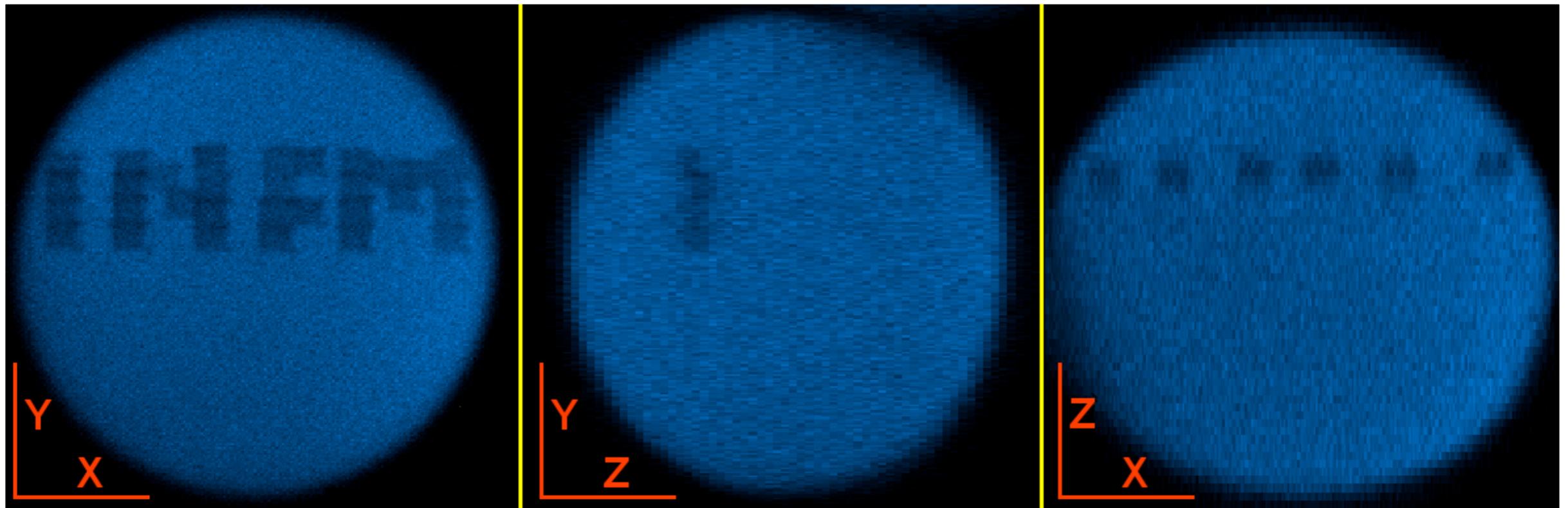


Eledone cirrhosa (octopus) sperm heads

DIASPRO ET AL., MICROSC. RES. TECH., 47(3), 196-205, 1999

TWO-PHOTON EXCITATION MICROSCOPY

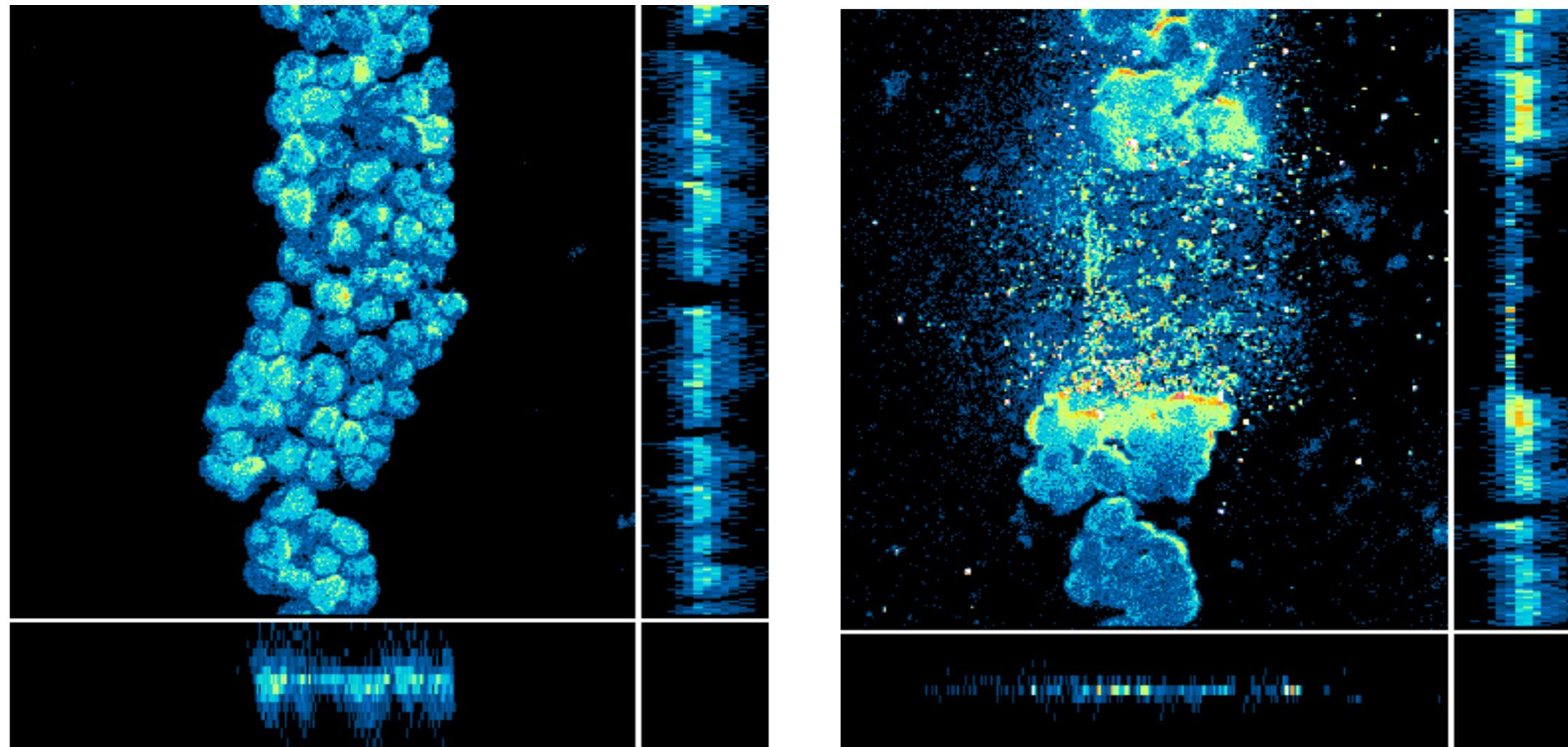
Two-photon micropatterning on fluorescent microspheres ($22 \mu\text{m}$) using photobleaching



DIASPRO ET AL., MICROSC. RES. TECH., 47(3), 196-205, 1999

TWO-PHOTON EXCITATION MICROSCOPY

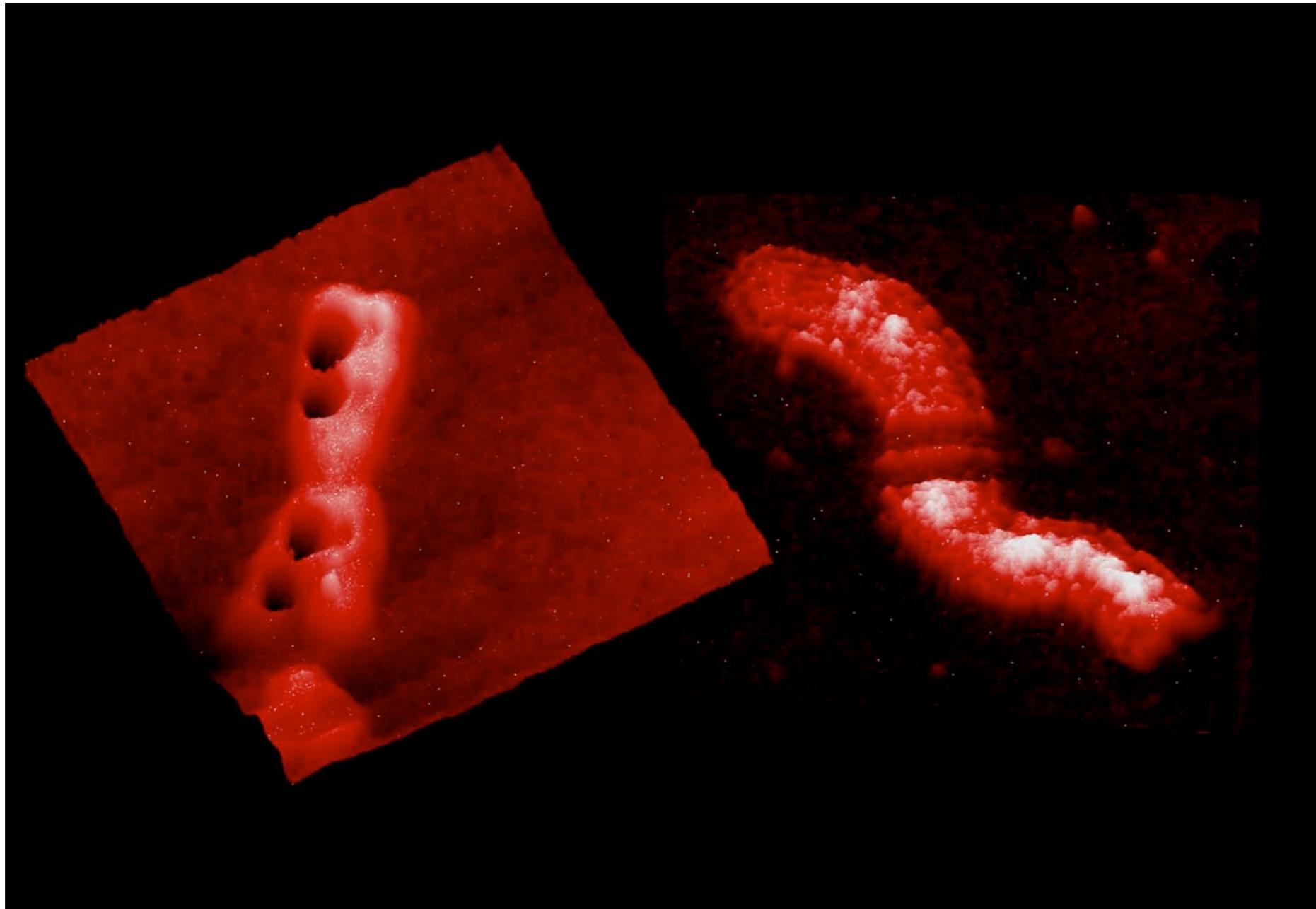
Selective 3D damage on
Saccharomyces cerevisiae cell layers



DIASPRO A., EJH, 1999

TWO-PHOTON EXCITATION MICROSCOPY

From Karsten Konig Lab



Journal of Microscopy, Vol. 200, Pt 2, November 2000, pp. 83–104.
Received 20 December 1999; accepted 16 June 2000

TWO-PHOTON EXCITATION MICROSCOPY

From Karsten Konig Lab

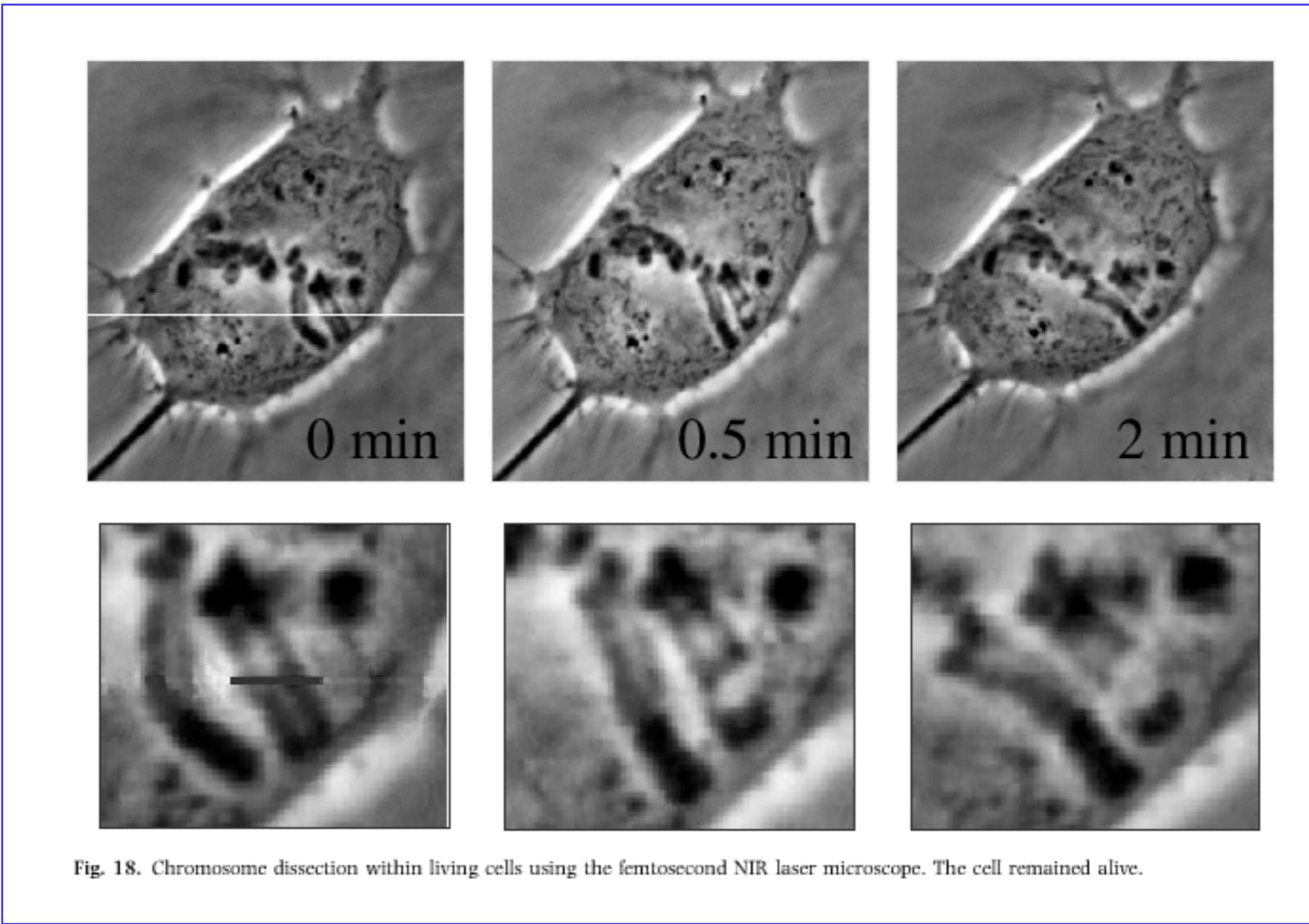
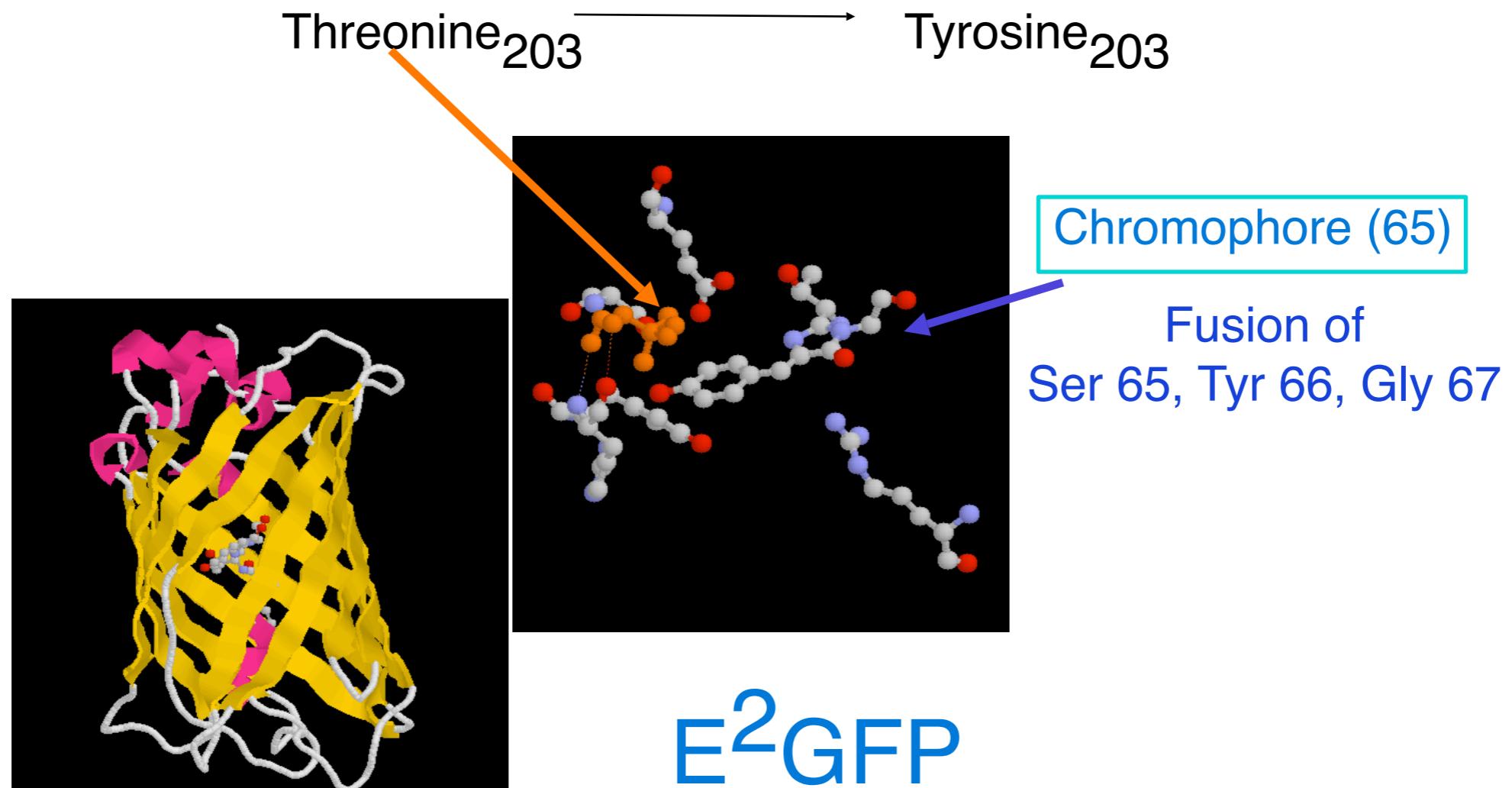


Fig. 18. Chromosome dissection within living cells using the femtosecond NIR laser microscope. The cell remained alive.

Journal of Microscopy, Vol. 200, Pt 2, November 2000, pp. 83–104.
Received 20 December 1999; accepted 16 June 2000

DIASPRO ET AL., *MICROSC. RES. TECH.*, 47(3), 196-205, 1999

SINGEL FLUORESCENT PROTEINS

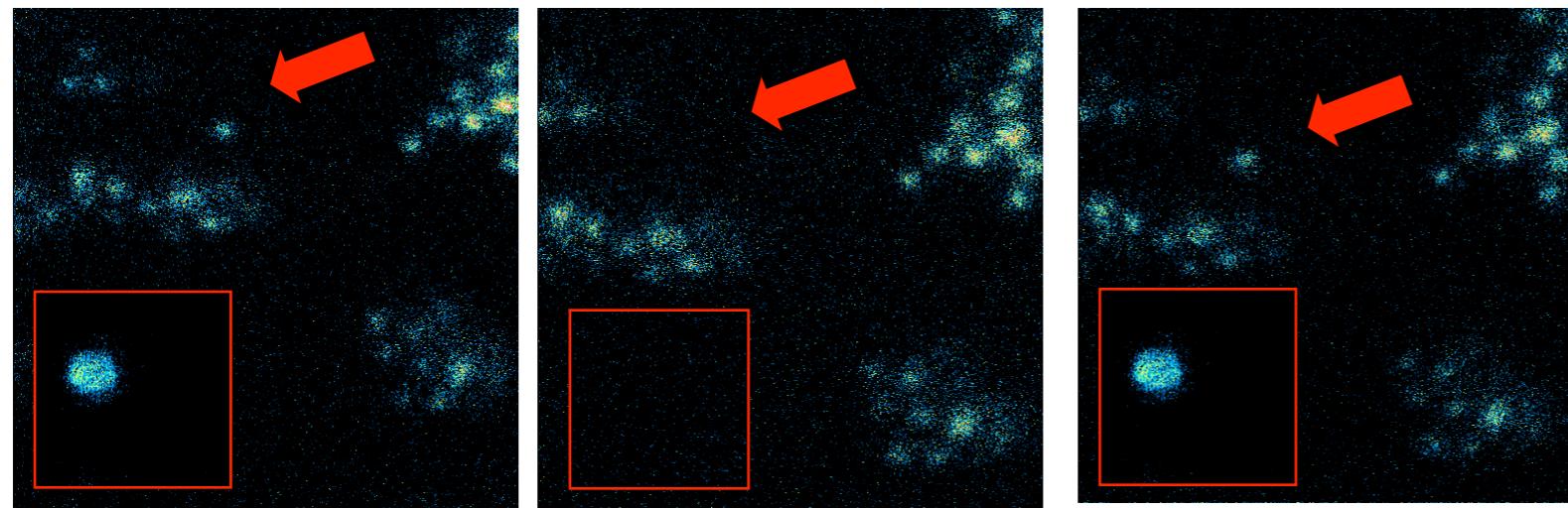


CINELLI RA, FERRARI A, PELLEGRINI V, TYAGI M, GIACCA M, BELTRAM F.,
 PHOTOCHEM PHOTOBIOLOGY. 2000 JUN;71(6):771-6.

CINELLI RA, TOZZINI V, PELLEGRINI V, BELTRAM F, CERULLO G, ZAVELANI-ROSSI M, DE SILVESTRI S, TYAGI M, GIACCA M.,
 PHYS REV LETT. 2001 APR 9;86(15):3439-42.

PHYS REV E STAT NONLIN SOFT MATTER PHYS. 2004 SEP;70(3 PT 1):030901

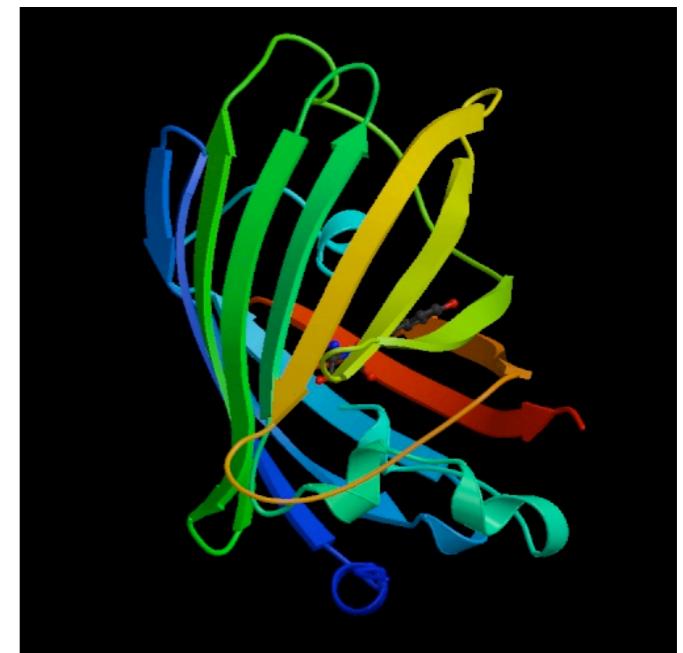
SINGEL FLUORESCENT PROTEINS



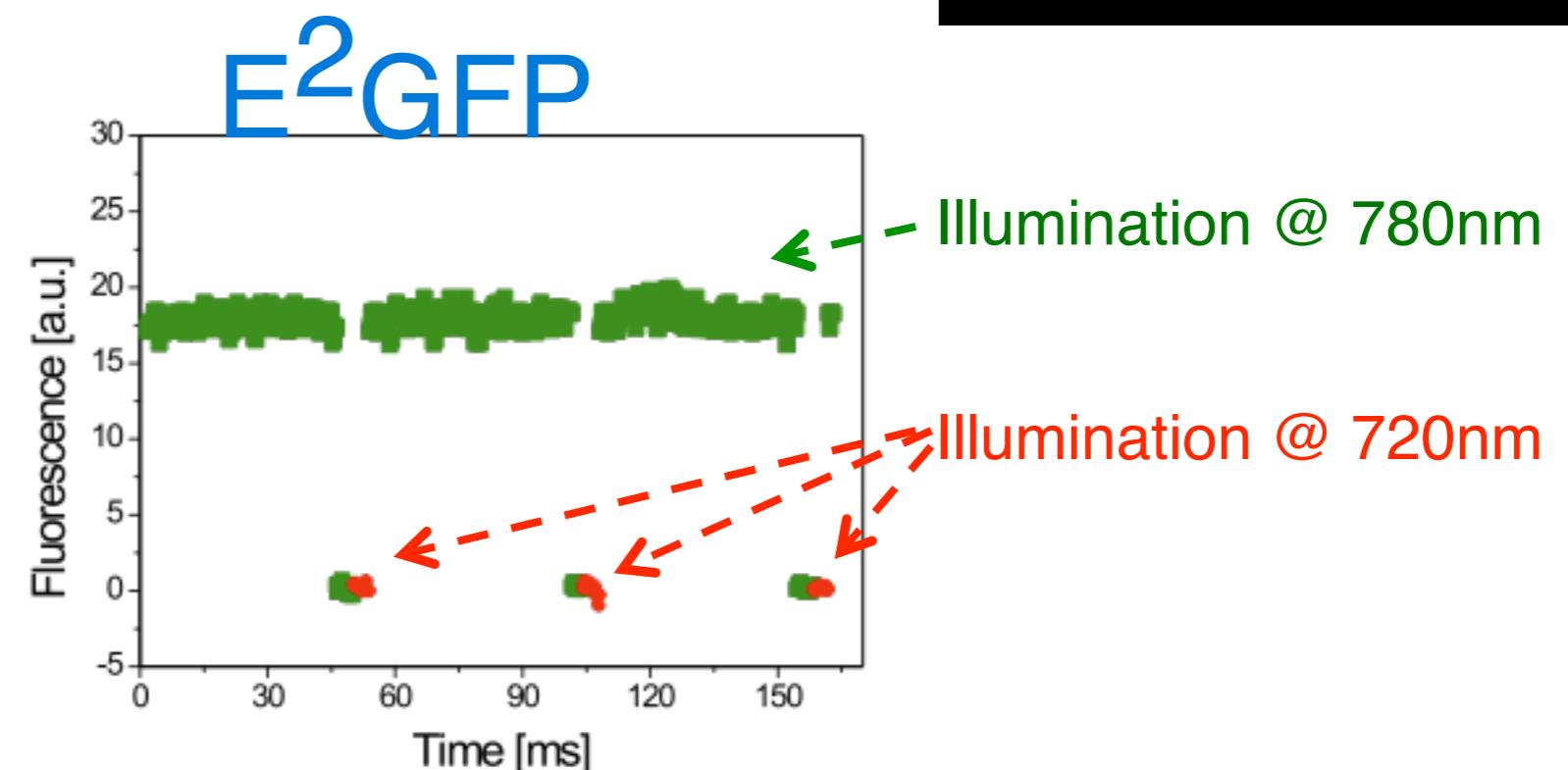
TPE @ 785 nm

TPE @ 720 nm

TPE @ 785 nm



E^2 GFP in silica GEL,
maximum excitation
within 700-800 nm range
@ 785; after imaging
photobleaching @ 785;
photocycling @ 720 nm -
no photorecovery @
710/730 -; final imaging
@ 785.

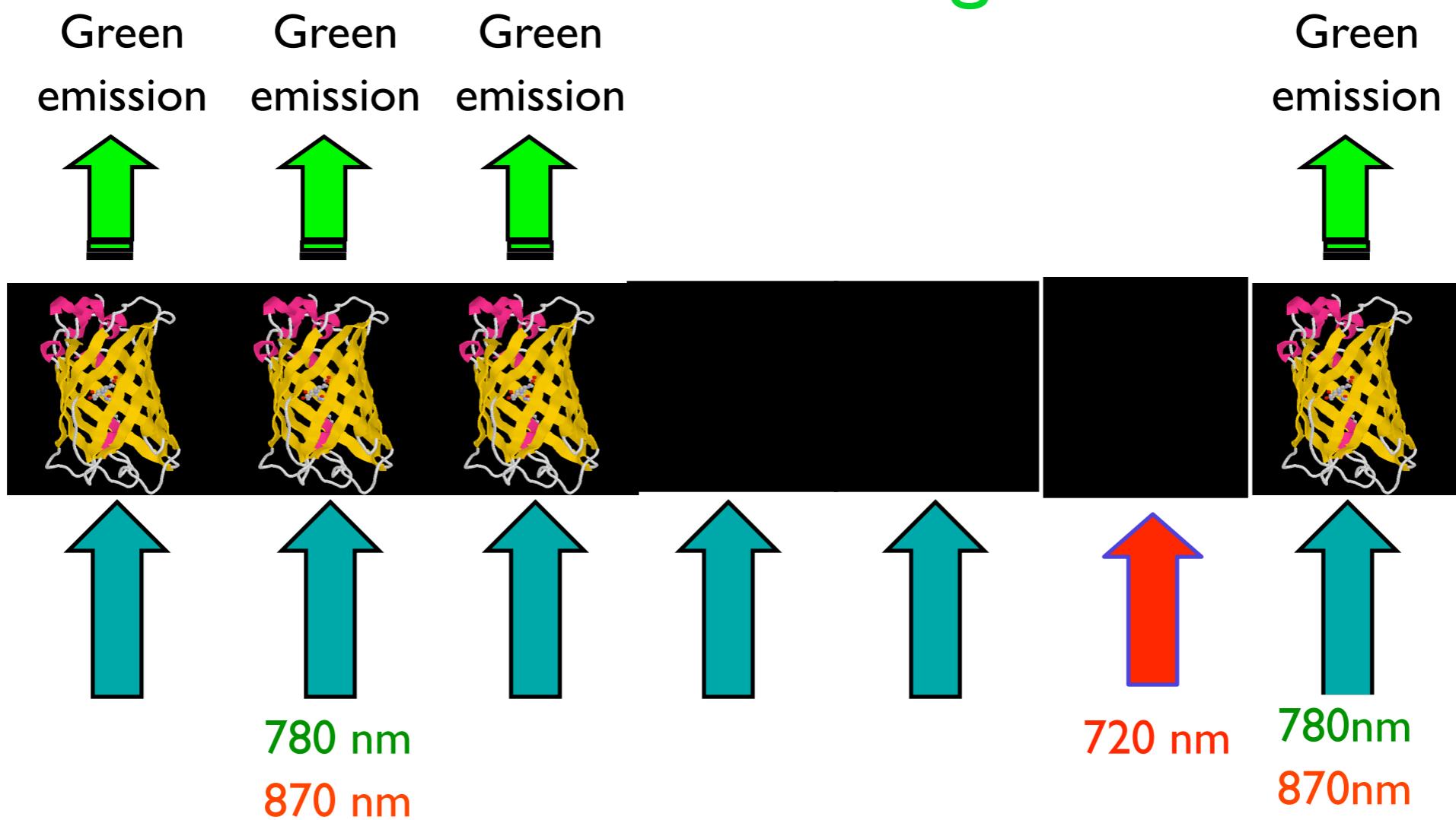


PHYS REV E STAT NONLIN SOFT MATTER PHYS. 2004 SEP;70(3 PT 1):030901

SINGEL FLUORESCENT PROTEINS

E^2GFP

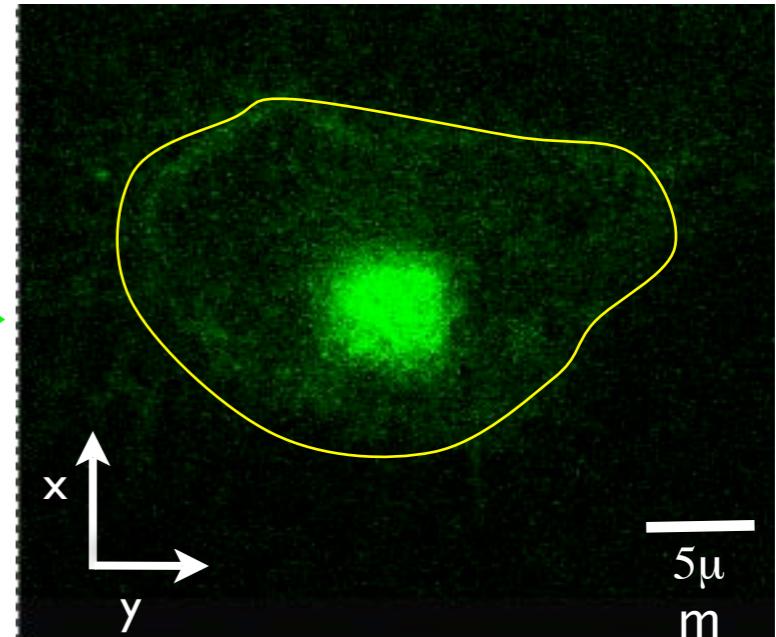
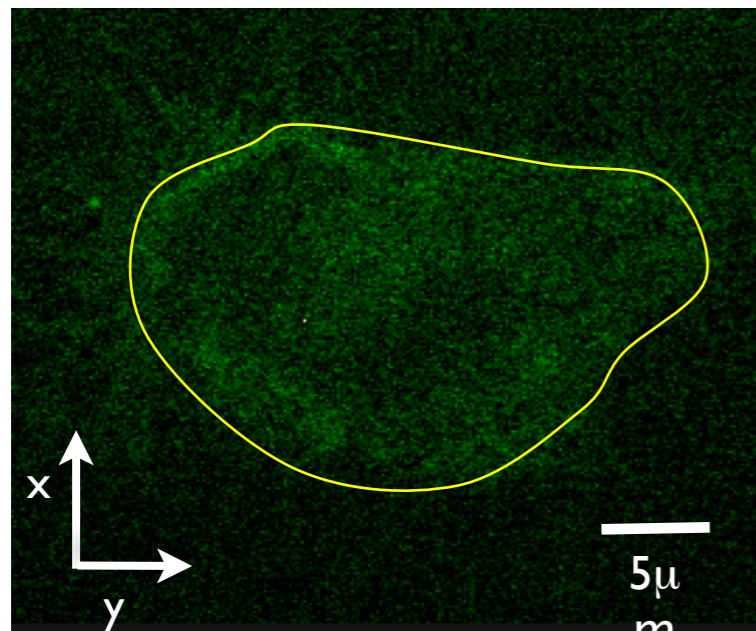
Two Photon Switching Protein



PHYS REV E STAT NONLIN SOFT MATTER PHYS. 2004 SEP;70(3 PT 1):030901

GFP-PHOTOACTIVATION

Switching on a subset of the expressed protein



PRE ACTIVATION

$\langle P_{EXCITATION} \rangle = 0,04 \text{ mW}$, $\lambda_{EXCITATION} = 488\text{nm}$
Pixel time = $4,9 \mu\text{s}$, 512×512 pixels

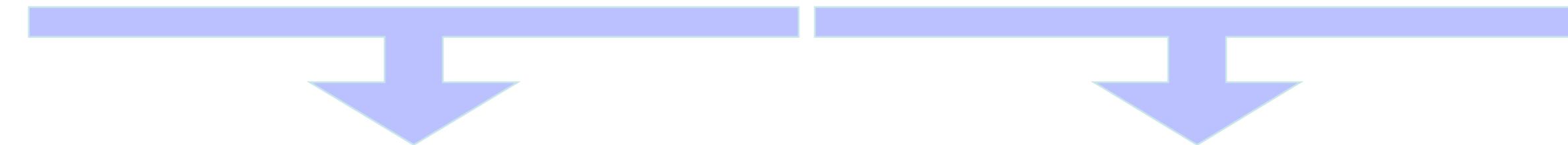
POST ACTIVATION

$\langle P_{EXCITATION} \rangle = 0,04 \text{ mW}$, $\lambda_{EXCITATION} = 488\text{nm}$
Pixel time = $4,9 \mu\text{s}$, 512×512 pixels

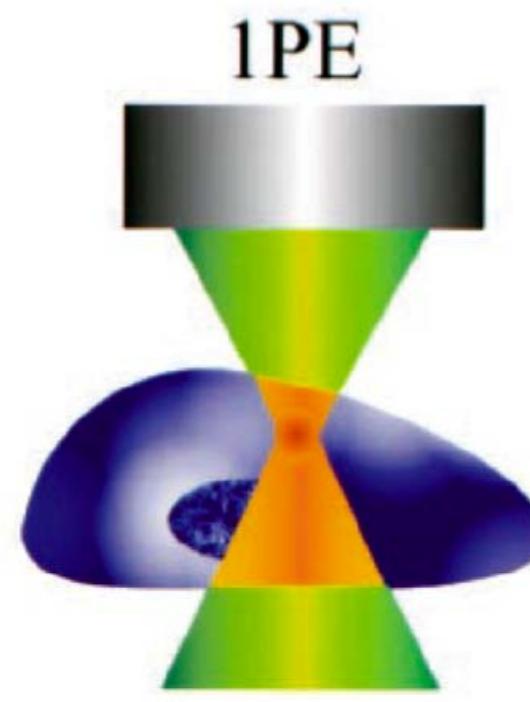
Photoactivation allows you to create a population of labeled proteins in a **desired volume** inside the cell

I.Testa, S.Barozzi, M.Faretta, A.Diaspro (2005) - LAMBS-IFOM Research Note, 7.

GFP-PHOTOACTIVATION

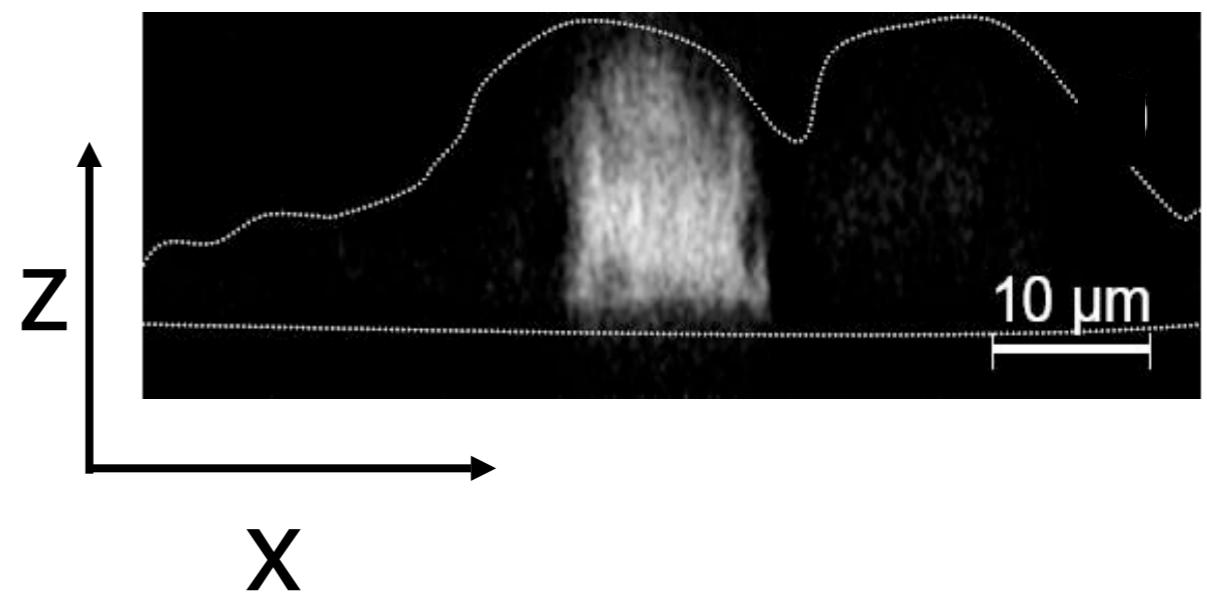


FLUORESCENCE MICROSCOPY



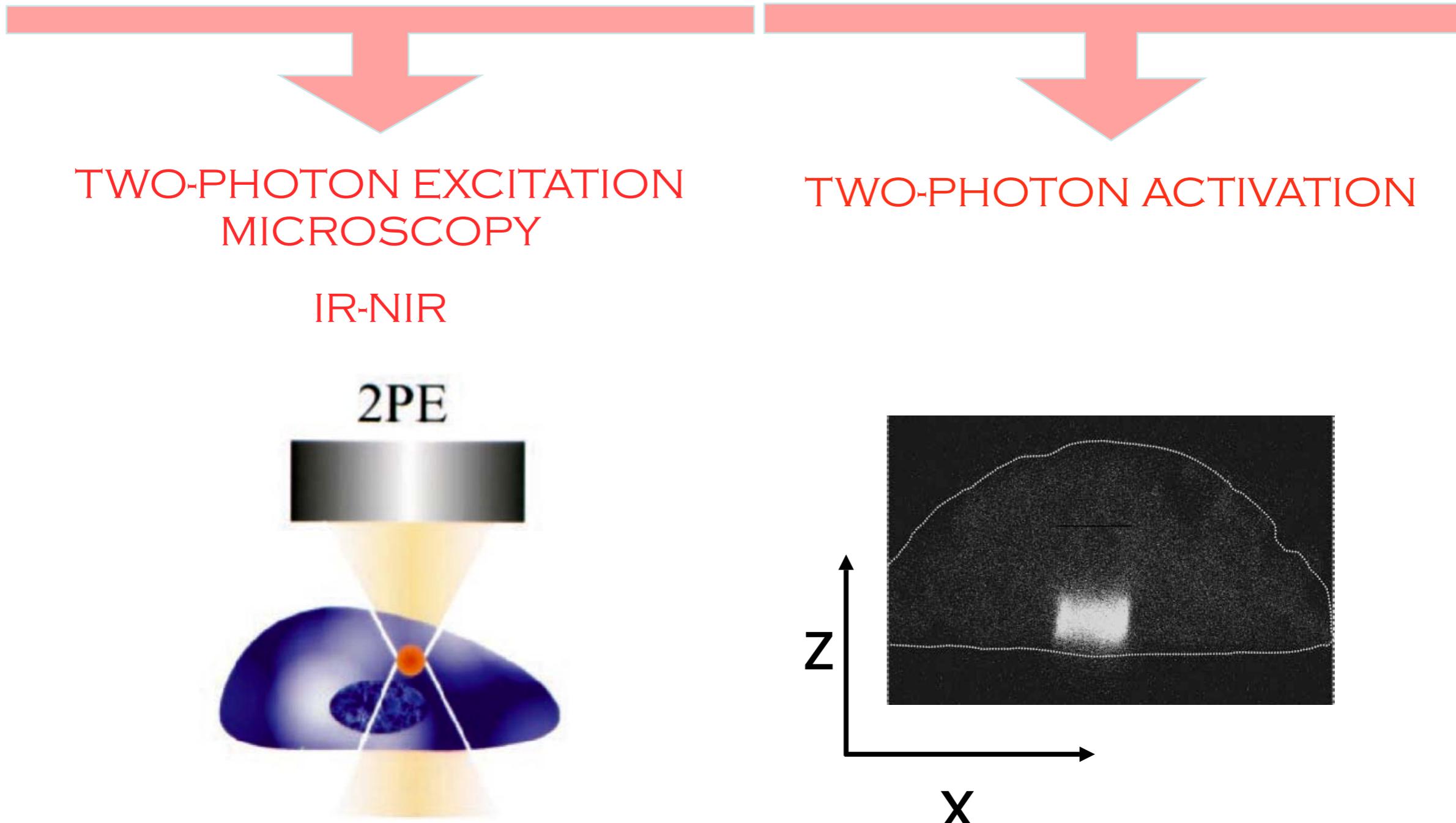
SINGLE-PHOTON ACTIVATION

**UV-Vis
405-413 nm**



SCHNEIDER M., BAROZZI S., TESTA I., FARETTA M., DIASPRO A.
(2005) BIOPHYS. J., VOL.89(2)

GFP-PHOTOACTIVATION



Denk, W., Strickler, J.H. & Webb, W.W. *Science* **248**, 73–76 (1990).

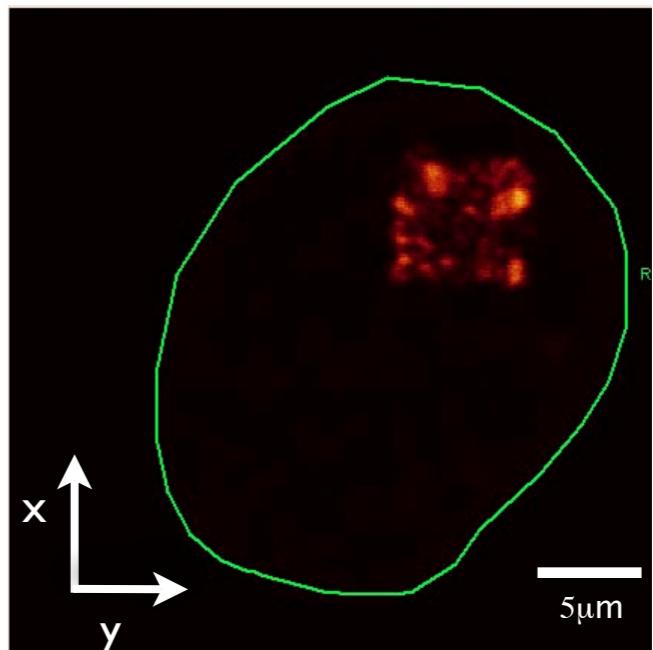
SCHNEIDER M., BAROZZI S., TESTA I., FARETTA M.,
DIASPRO A. (2005) *BIOPHYS. J.*, VOL.89(2)

TWO PHOTON ACTIVATION IN LIVING CELLS

PA-GFP + H2B
He-La cell

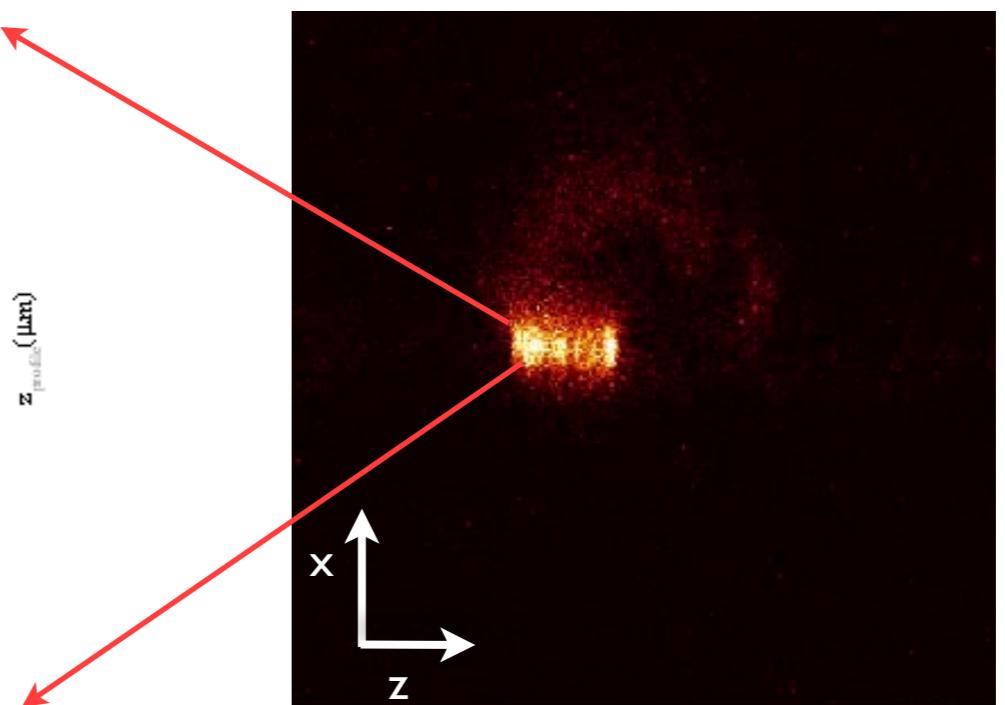
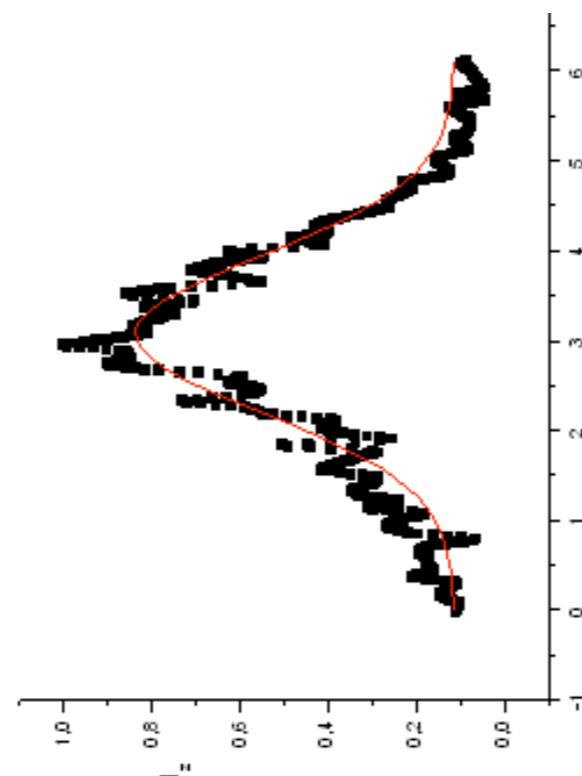


$\langle P_{\text{ACTIVATION}} \rangle = 10 \text{ mW}$
 $\lambda_{\text{ACTIVATION}} = 720 \text{ nm}$
Pixel time = $300 \mu\text{s}$

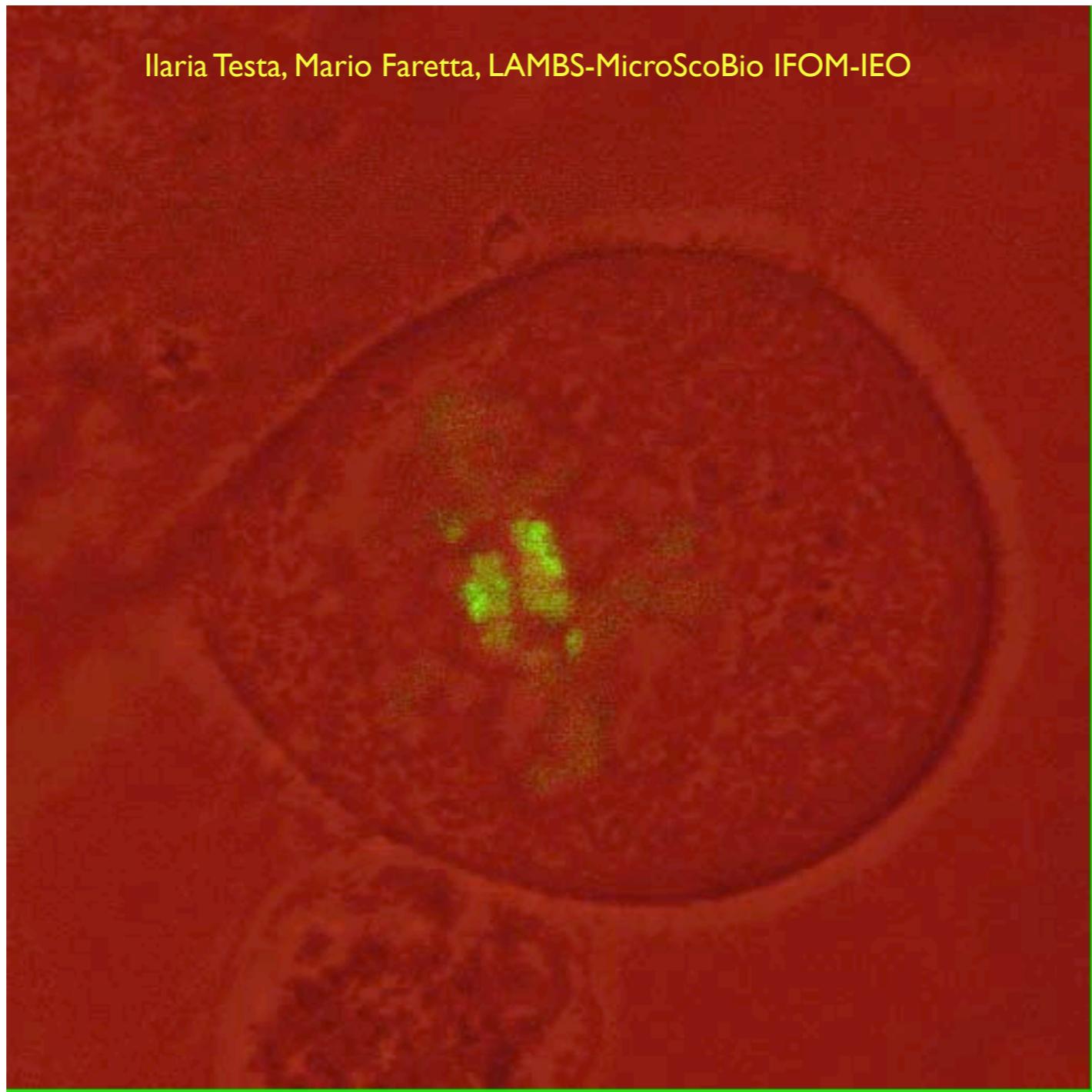


$\langle P_{\text{EXCITATION}} \rangle = 0,04 \text{ mW}$, $\lambda_{\text{EXCITATION}} = 488 \text{ nm}$
Pixel time = $4,9 \mu\text{s}$, 512×512 pixels

LOCALIZED
INTERACTION



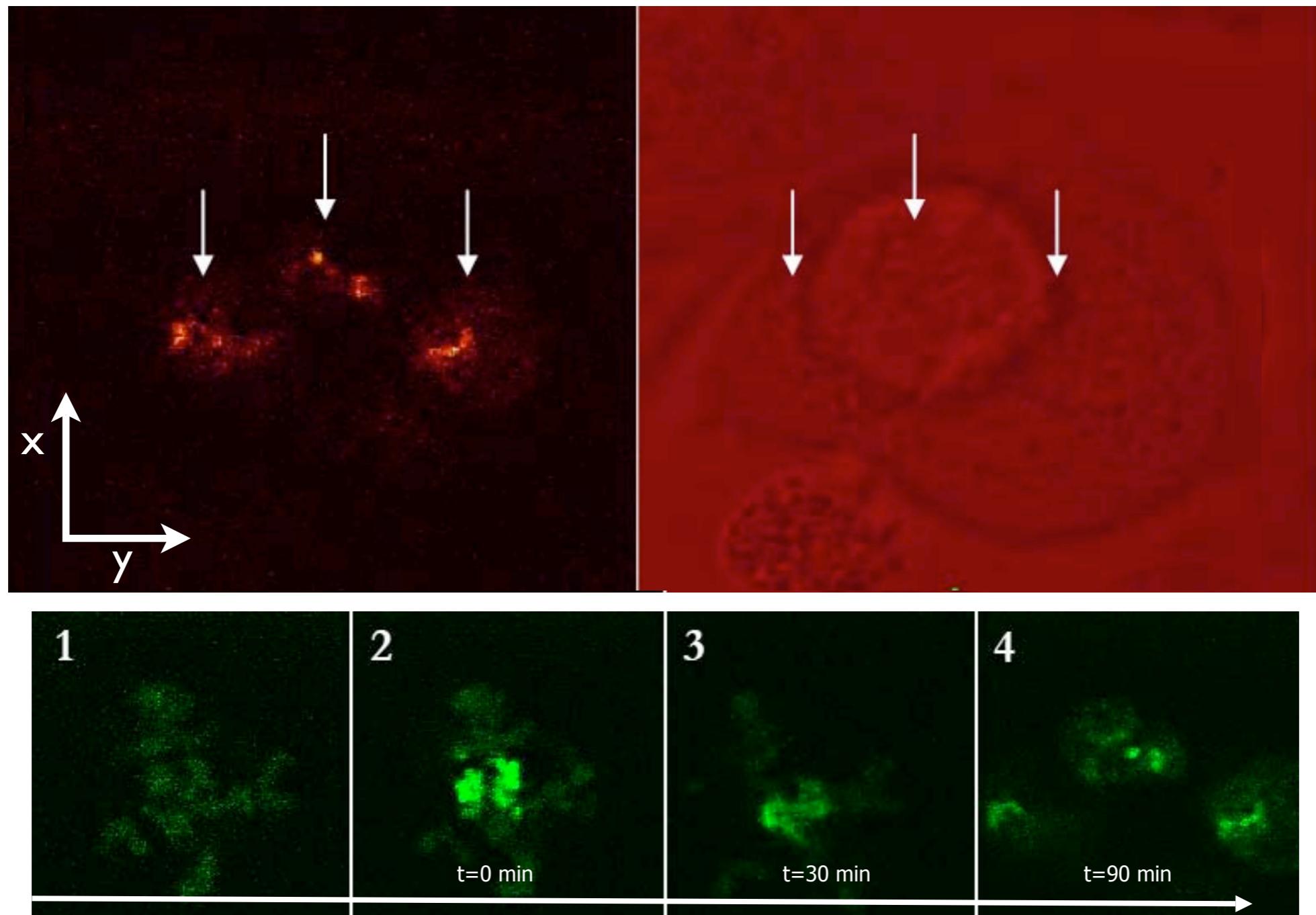
TWO PHOTON ACTIVATION IN LIVING CELLS



Ilaria Testa, Mario Fareta, LAMBS-MicroScoBio IFOM-IEO

I.Testa, S.Barozzi, M.Fareta, A.Diaspro (2005) - LAMBS-IFOM Research Note, 7.
PA-GFP, LEICA SP2 AOBS spectral system, Chameleon-XR Coherent tunable Ti-Sapphire laser

TWO PHOTON ACTIVATION IN LIVING CELLS



I.Testa, S.Barozzi, M.Fareta, A.Diaspro (2005) - LAMBS-IFOM Research Note, 7.
PA-GFP, LEICA SP2 AOBS spectral system, Chameleon-XR Coherent tunable Ti-Sapphire laser

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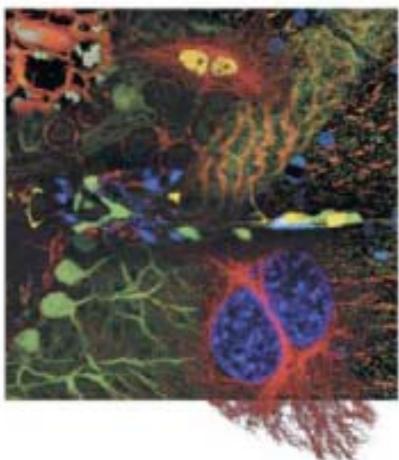
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